



The Importance of Standardized Toxicological Methods for Aquatic Organisms



Combined with New Approach
Methodologies (NAMs)

Sandy Raimondo
US Environmental Protection Agency
24 January 2023

The views expressed in this presentation are those of the author and do not necessarily reflect the views or policies of the US EPA

A Little Background

- The NASEM literature review found extremely limited data on the toxicity of UV-filters on various aquatic organisms (animals and plants)

- Oxybenzone = most data rich (Ecamsule = most data poor, N=4)

- 123 toxicity values
- 38 species
 - 4 fish
 - 21 invertebrates (8 coral)
 - 11 algae
 - 2 bacteria



Looks better on paper
than in application

- NASEM report to spark a flurry of research, let's ensure it produces data we can use in a consistent, defensible, and reproducible manner
 - Species lacking standard test protocols will be investigated
 - Standardized data can still be obtained

Discussion Goals

- Brief overview of what it means to be a “standardized” test method
- How are data from these protocols used?
 - Tier 2 Ecological Risk Assessment
- Limitations of standardized tests
- Looking toward the future:
 - Higher tiered assessments
 - Standardized endpoints + New Approach Methodologies (NAMs)



Availability of Standard Methods

- Internationally accepted standards
 - US EPA
 - Organisation for Economic Co-operation and Development (OECD)
 - American Society for Testing and Materials (ASTM)
- Test Types:
 - Acute & chronic aqueous (freshwater & saltwater)
 - Chronic sediment (freshwater & saltwater)
 - Terrestrial (worms, plants, mammals, birds)
 - Microbial
- Aquatic taxa:
 - Fish
 - Amphibians
 - Crustaceans
 - Molluscs
 - Algae

Don't see what you need?
Testing a species without a formal
“standard method” doesn't have to
completely re-invent the wheel



United States
Environmental Protection
Agency

Office of Chemical Safety
and Pollution Prevention
(7101)

EPA 712-C-16-013
October 2016

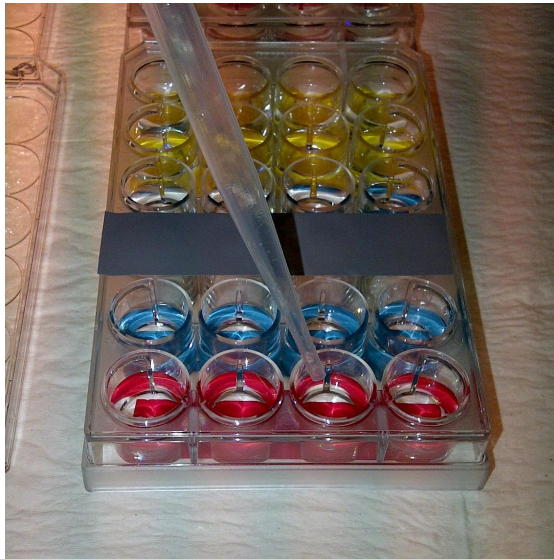
Ecological Effects Test Guidelines

OCSPP 850.1010:
Aquatic Invertebrate
Acute Toxicity Test,
Freshwater Daphnids



What Do Aquatic Toxicity Tests Look Like?

- Replicated, controlled laboratory studies
- Single stressor (i.e., UV-filter)
- Expose organism to range of concentrations
- Static, static renewal, or flow through
- Ideally analytical chemistry to confirm each concentration



Single replicate 24-well plate small-scale exposures with fish embryos. Each well row housed fish exposed to a unique concentration.



Single replicate table of large-scale fish exposures at the EPA Gulf Breeze Laboratory. Each tank housed fish exposed to a unique concentration.

What is a Test “Endpoint”?

- Survival, growth, reproduction?
- LC50, EC50, EC20 (Lethal/Effect concentration of given percentage)
- NOEC/LOEC (No/lowest observable effect concentration)?

Endpoint = Exposure duration + Statistic + Biological effect

Endpoint examples:

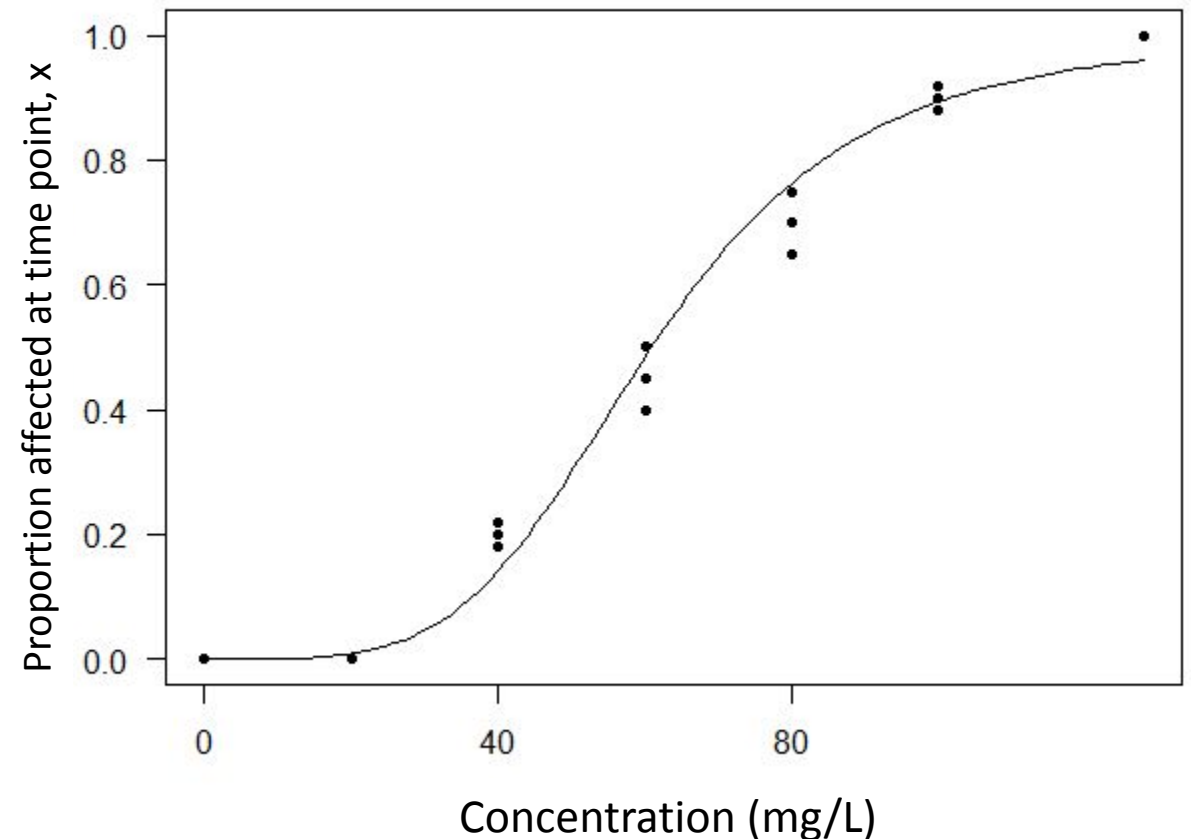
- 96 h LC50 (lethality)
- 28 d EC20 for growth
- 270 d NOEC/LOEC for reproduction

Aquatic Test Data

Concentration Response Curve

The relationship of biological response with concentration and time

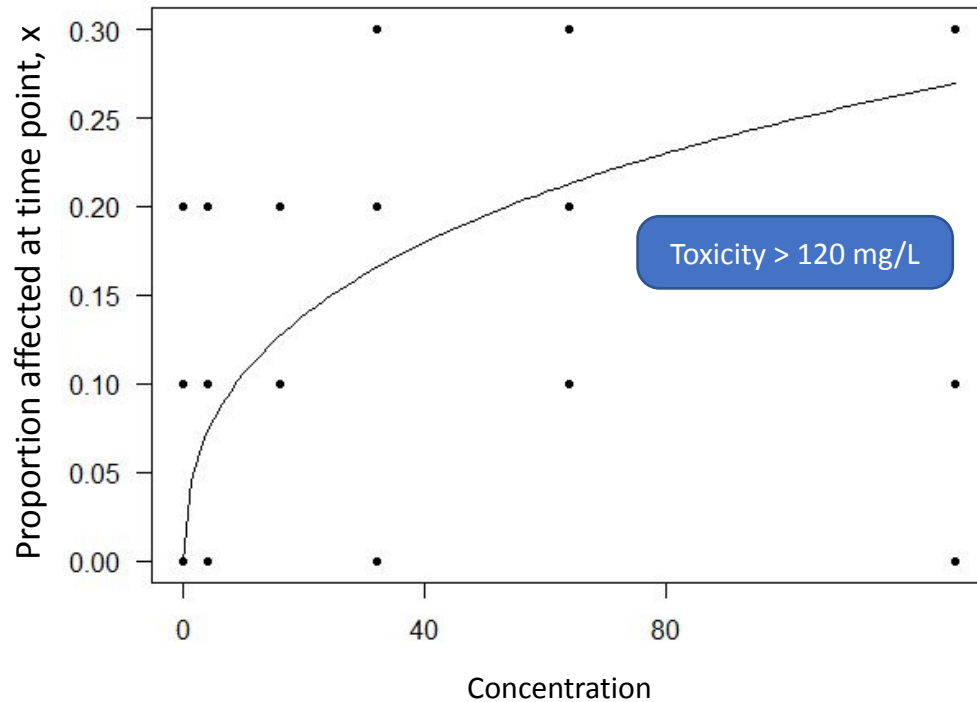
- Influenced by:
 - Exposure duration
 - Concentration ranges
 - Test design
- Data needs to be conclusive
- Clear and significant relationships



Pilot, Range-finding, and Inclusive Studies

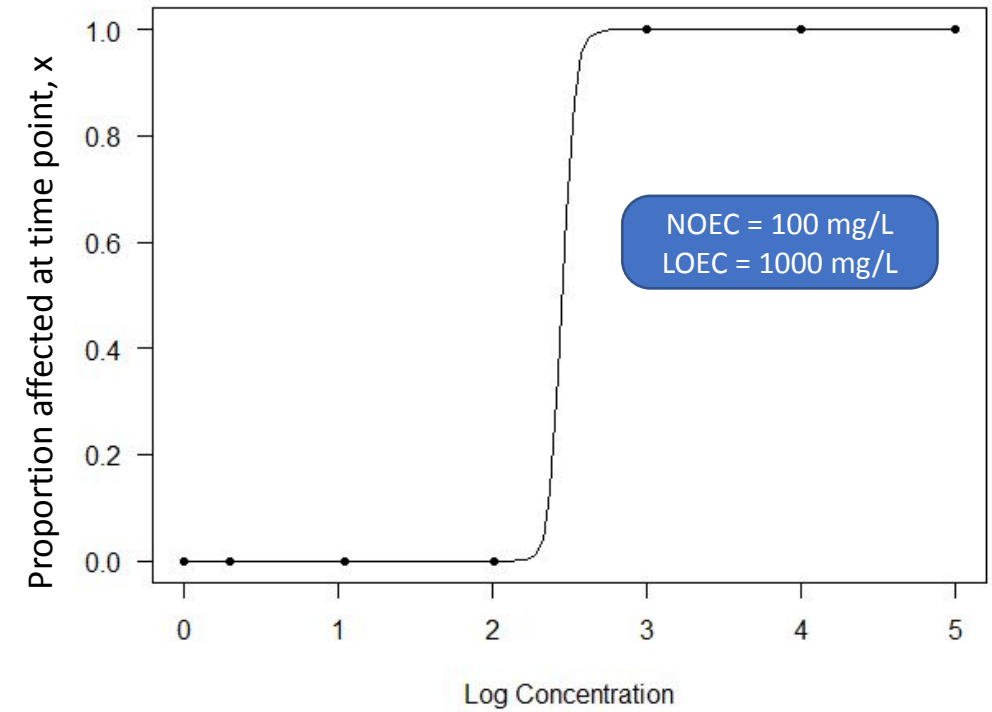
Pilot study:

Work out kinks in the methods to reduce variability across replicates and ensure definitive test



“Range finding” study:

Conducted to identify a range in which effects are most likely to occur



What does it mean to be “standardized”?

- Exposure duration
- Biological response
 - Can range from immobilization to population growth rate, depending on species
- Age/life stage
- Test conditions
 - Temperature
 - Dissolved oxygen
 - Salinity
 - Hardness
 - pH

Within a species/taxa

Within a chemical

- Active ingredient (%)
- Analytical methodologies
- Normalizations

Across test reports

- What constitutes a “successful” test?
- Data analysis = statistic
- Open-ended toxicity values (i.e., > 100 mg/L) are way too common

What Does it Mean to be “Standardized”?

Acute vs Chronic

Acute toxicity

- Short term (24 - 96 hr*)
- Endpoint
 - Lethal (LC) or effect concentration (EC) of 50%
 - e.g., 96-h LC50, 48-h EC50
- Pros:
 - Quick, resource effective
 - Most common & abundant toxicity test data
- Cons:
 - Minimal information
 - Lack ecological relevance

Chronic toxicity

- Longer term (> 7 days*)
 - Early life stage (ELS)
 - Partial life cycle (PLC)
 - Life cycle (LC)
- Traditional statistics
 - NOEC/LOEC – not associate with % effect
 - EC20
- Biological responses
 - Growth, reproduction, lethality, biomass
 - Histopathology, behavior
 - Multi-omics, Adverse Outcome Pathways (AOPs)
- Pros: exposure durations and endpoints more ecologically relevant
- Cons: more resource intensive

Common Test Species

Species/Taxa

- Amenable to culture in laboratory systems
- Bias towards smaller individuals and life stages
- Not always the most sensitive

Data source:
www3.epa.gov/webice

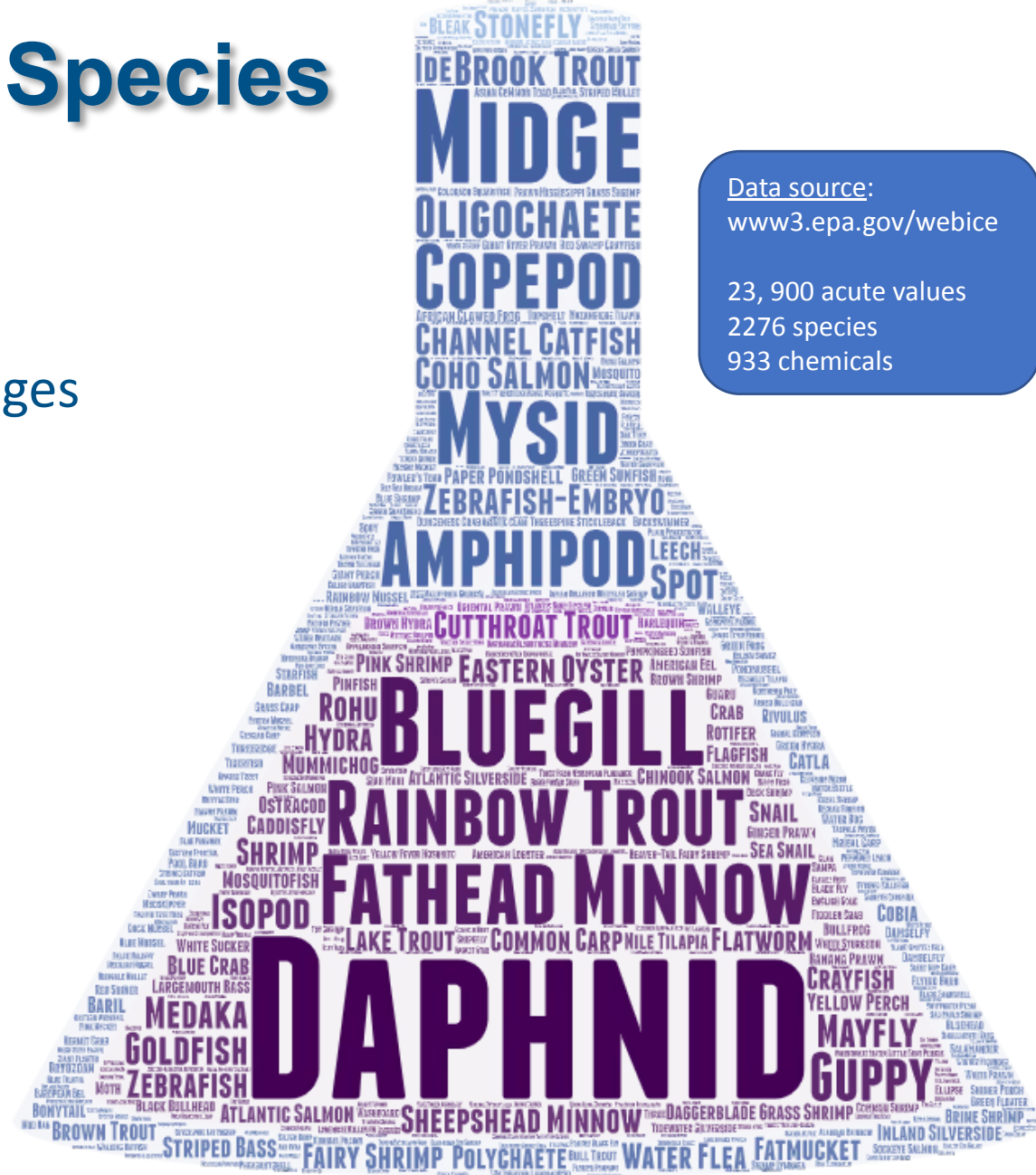
23, 900 acute values
2276 species
933 chemicals



≠



from the gallery of [szymek drobniak](#)



Discussion Goals

- Brief overview of what it means to be a “standardized” test method
- How are data from these protocols used?
 - Tier 2 Ecological Risk Assessment
- Limitations of standardized tests
- Looking toward the future:
 - Higher tiered assessments
 - Standardized endpoints + New Approach Methodologies (NAMs)



Toxicity Data Use: General

Data are used to understand:

- How a chemical may adversely affect an organism
- Which taxa/communities are most at risk
- Where should limited resources be invested for refined understanding of environmental hazards
- What mitigation efforts may reduce environmental impact

Data are applied to:

- Establish registered use of chemical that results in *de minimus* environmental impacts
- Establish water quality standards that guide State/Tribe permits and environmental restoration
- Assess impacts of accidental chemical release into to the environment

Example Application: Oxybenzone

Most data rich of all UV filters: 123 toxicity values, 38 species

Acute values

- 68 values, 28 species
- 7 open ended/inconclusive
- 24 -96 h
- Duration of exposure often, but not always, consistent within a species/taxa

48-h EC10	24-h LC/EC50	60-h EC50
48-h LC/EC50	72-h LC/EC50	96-h LC/EC50
48-h LC86	72-h EC50	96-h NOEC
48-h LOEC	72-h NOEC	96-h LOEC
	72-h LOEC	

Chronic values

- 54 values, 20 species
- 29 open ended/inconclusive

Exposure duration (d) = 7, 10, 12, 14, 20, 21, 28, 30, 42, 60
Statistic = EC5, EC10, EC20, EC50, ICx*, NOEC, LOEC

Biological response:

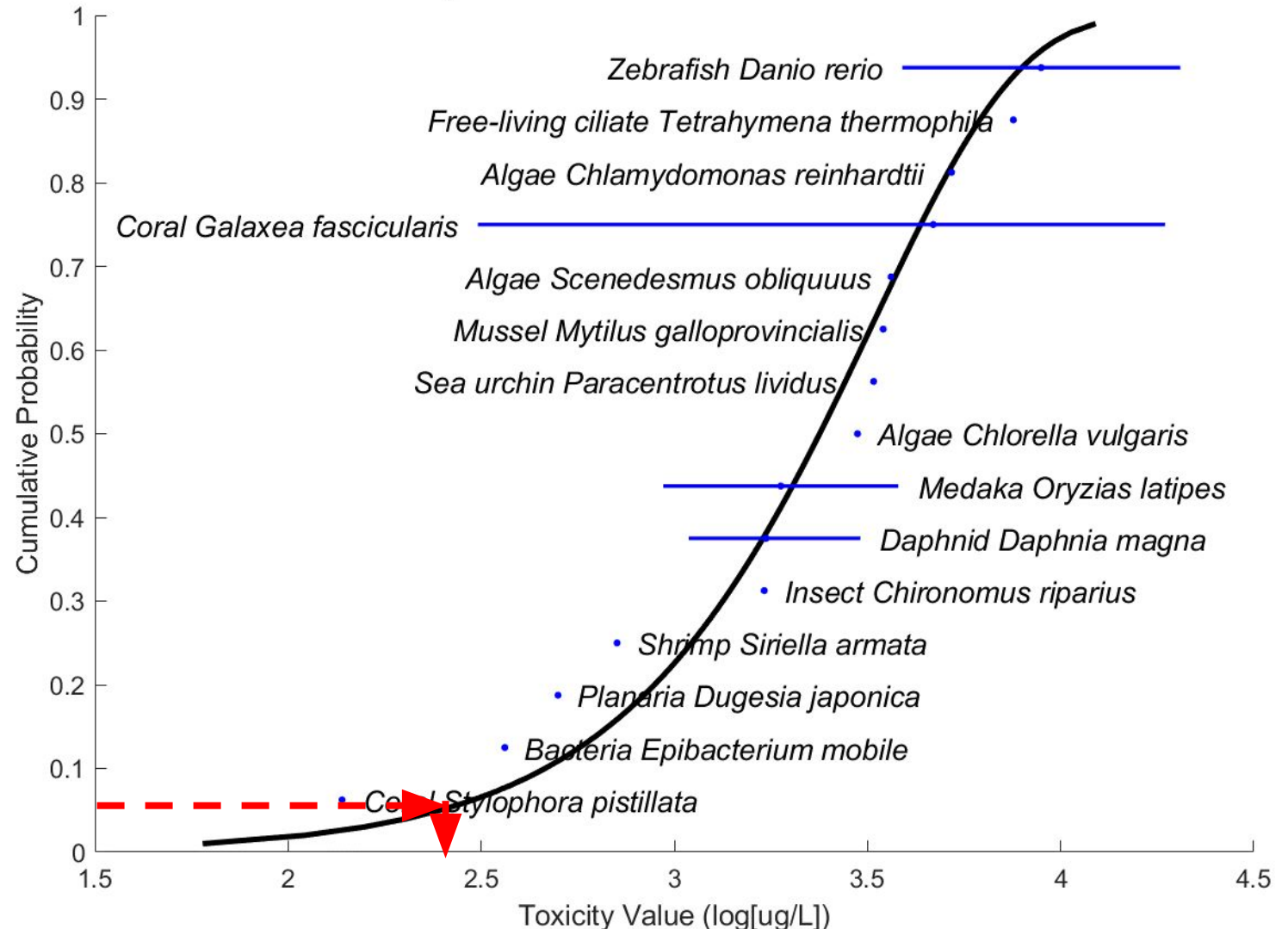
density, settlement, cell morphology, microbiom, bleaching, fecundity, PSII yield, chlorophyll, "Fv/Fm", length, weight, Y mortality, reproduction, hatch rate, growth rate, zooxanthellae density, on "K/HIS factors", hatch time

All introduce variability and uncertainty into data application and interpretation

Toxicity Data Use: Species Sensitivity Distributions (SSDs)

- Cumulative probability distribution
- HC5 = Hazardous concentration of the 5th percentile
 - Metric used as the basis for many risk determinations
 - Compared to measured/estimated environmental concentrations
- Acute and Chronic
- Identify which species are the most (or least) sensitive
- Estimate a concentration at which most taxa would be protected
- Can apply different approaches to adjust for uncertainties

Oxybenzone standardized data



SSD with Standardized Data: Oxybenzone

Standardized endpoints

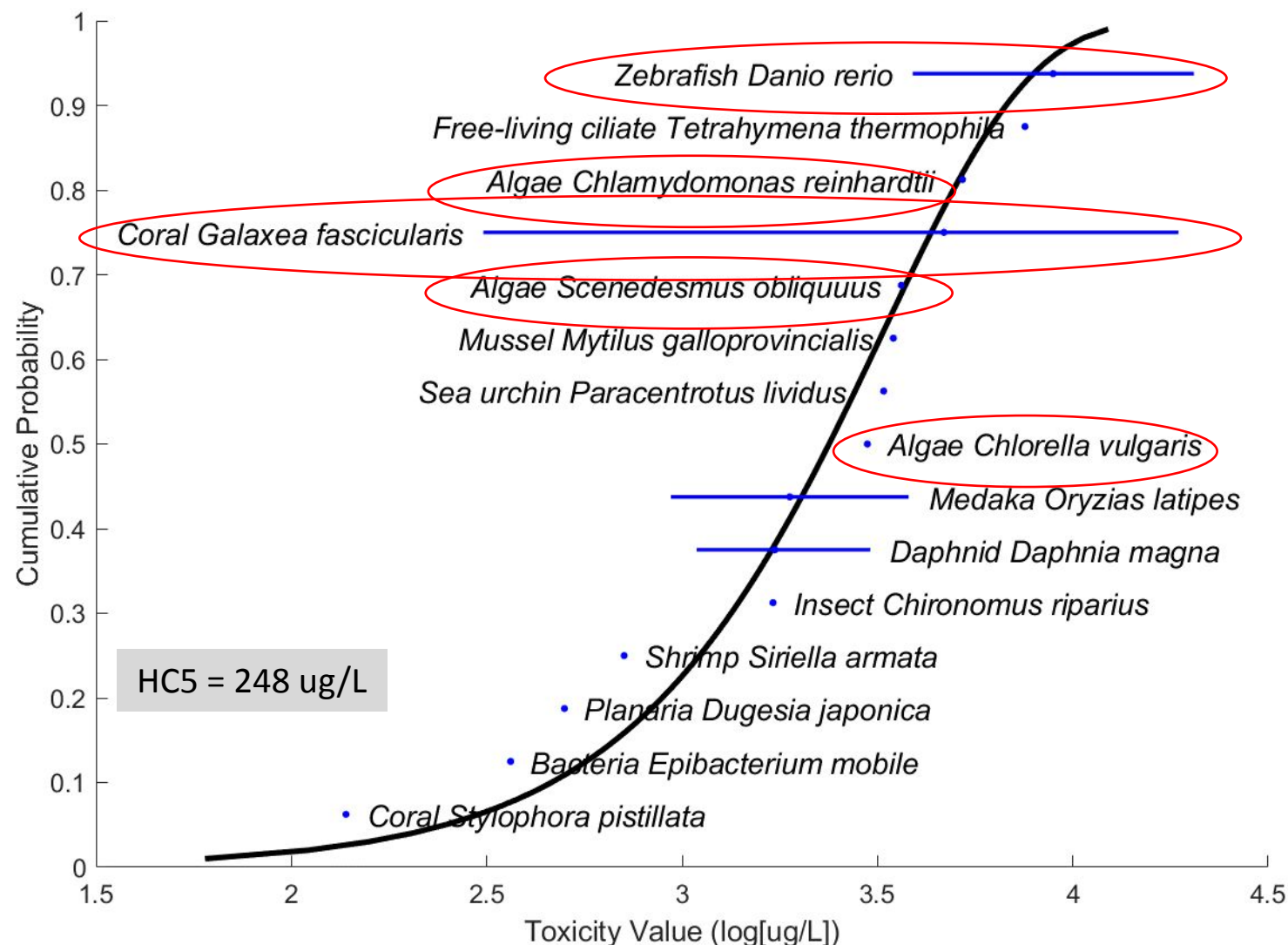
- All values EC/LC50
- 15 species, 36 test results (53% of 68 values)

Standardized species

- Algae: 96 h EC50
- Daphnid: 48 h EC50
- Mollusc larvae: 48 h EC50
- Insects, mysids: 96 h LC50
- Fish: 96 h LC50

Non-standardized species

- Sea urchin: 48 h EC50
- Free-living ciliate: 24 h EC50
- Planaria: 96 h LC50
- Bacterium: 60 h EC50
- Coral:
 - Planulae: 24 h EC/LC50
 - Adult fragments: 96 h EC/LC50



SSD with All Data: Oxybenzone

Included all values except open ended

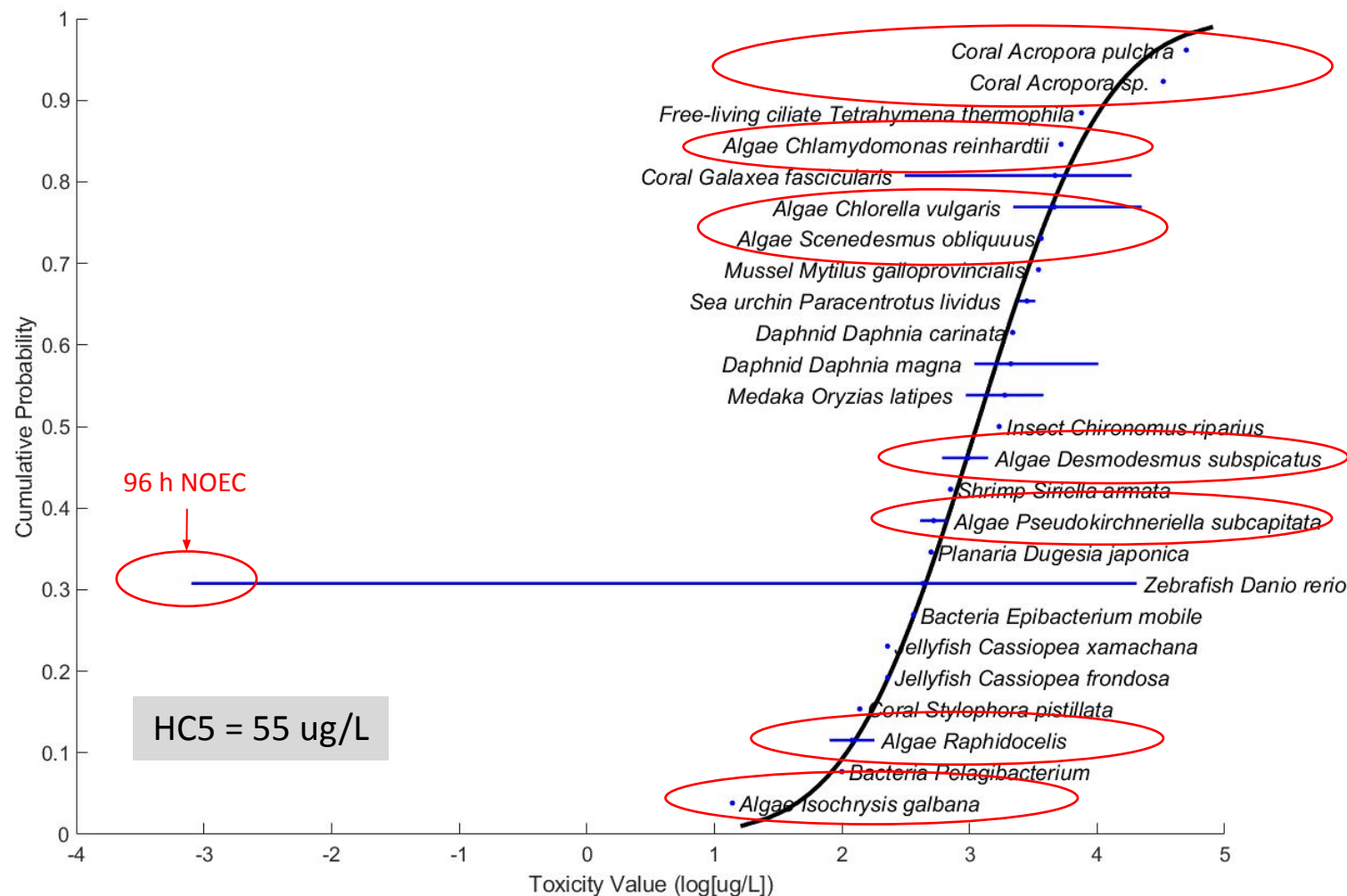
- 25 species, 61 datapoints
- Excluded open-ended values (i.e., > 100 ug/L)

What we added:

- Coral least sensitive = LC86 (48 & 96 h)
- Sea urchin = EC10, EC50
- Zebrafish
 - least sensitive with standardized data
 - 48 - 96 h NOEC, LOECs,
- Jellyfish = 72-h LOEC
- Daphnids = 24 – 48 h
- Algae = NOECs, LOEC, IC10s, IC50s
- Bacteria = 48 h LOEC

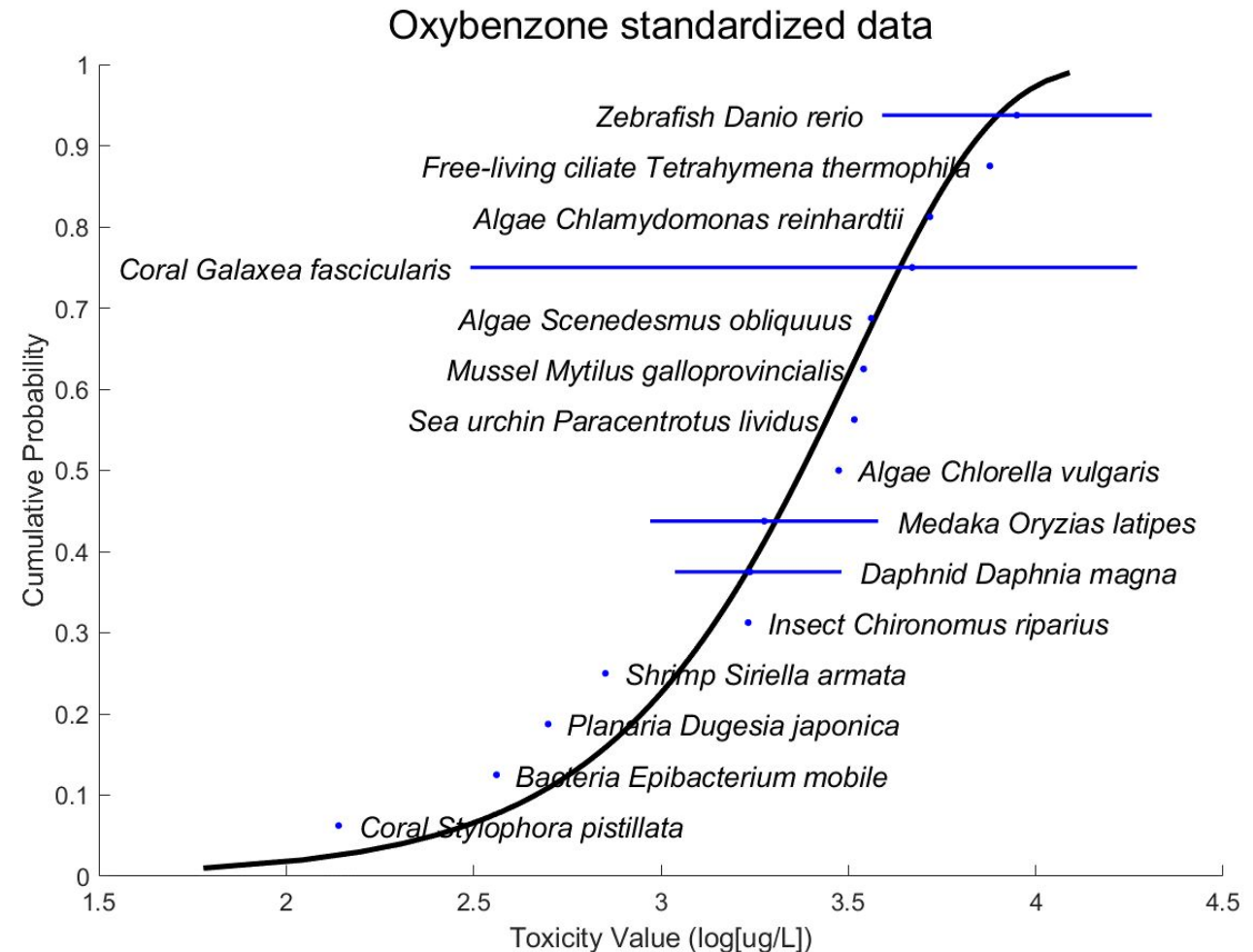
Bottom line:

- We added a lot of noise
- Don't know relative species sensitivity



Why Standardized Data Are Important

- Reduces the variation in interspecies and inter-chemical comparison
- Prioritization of mitigation and management
 - Misrepresentation of relative species sensitivity may misguide decisions
- Standardization within EPA
 - Non-standardized endpoints not often used quantitatively
 - A LOT of thrown out data
- #1 DEFENSIBILITY



Discussion Goals

- Brief overview of what it means to be a “standardized” test method
- How are data from these protocols used?
 - Tier 2 Ecological Risk Assessment
- Limitations of standardized tests
- Looking toward the future:
 - Higher tiered assessments
 - Standardized endpoints + New Approach Methodologies (NAMs)



Limitations of Standardized Tests

- Limited species with methods developed
- Limited interpretation of standardized endpoints
- Limited environmental realism

Nonstandardized lab tests
contain all these limitations
PLUS incompatibility,
incongruence with other data

- Interspecies extrapolation from test species
- Lab-to-field extrapolation
 - Environmental exposure
 - Variability of biotic and abiotic environment
- Individual to population



≠



Photo credit: Charles LoBue/EPA

Discussion Goals

- Brief overview of what it means to be a “standardized” test method
- How are data from these protocols used?
 - Tier 2 Ecological Risk Assessment
- Limitations of standardized tests
- **Next steps**
 - Higher tiered assessments
 - Standardized endpoints + New Approach Methodologies (NAMs)



Toxicity Testing of UV Filters without Formal Standards

- Develop *transferrable* test designs
 - Developing tests for non-standard organisms should be transferrable to other facilities
 - “Standardization” = Reproducibility, Reproducibility, Reproducibility
- Endpoints
 - More is more
 - ECx, NOEC, LOECs for widely accepted exposure durations (acute, chronic)
 - Consensus on endpoints for non-standard species (e.g., coral)
 - Endpoints don’t need to be the same as other species
 - Do need to be consistent across tests
 - Non-apical endpoints without quantitative links to apical endpoints difficult to defend

Higher Tiered Risk Assessments

- Improve environmental realism
- Methods and models for laboratory-to-field extrapolation
 - Includes extrapolation from standard endpoints to relevant ecological endpoints
- Improve inter-species extrapolation
 - Identify species amenable to controlled laboratory studies
 - Develop approaches for robust interspecies extrapolation
 - From 'omics to traits-based approaches
 - Improves understanding of sensitivity of untested species
- Integration of laboratory data with New Approach Methodologies (NAMs)



Photo credit: Cheryl Hankins/EPA

Thoughts for Moving Forward

- Gain applicable toxicity knowledge
 - Screening sensitivity assays
 - Prioritization of chemicals and organisms
- Avoid “minimal publishable unit”
 - Repeat studies if needed
 - Publish useable data
- Develop NAMs compatible with toxicity tests
 - Marine environments
 - Abiotic variability
 - Interspecies extrapolations
 - Look ahead to higher tiered assessments



Photo credit: Cheryl Hankins/EPA

Questions?

Raimondo.Sandy@epa.gov

The views expressed in this presentation do not necessarily reflect those of the US Environmental Protection Agency

