

Assessing readiness of an omics signature for use in a clinical trial

Improving Cancer Diagnosis and Care: The Clinical Application of Computational Methods in Precision Oncology

National Academies of Sciences, Engineering, and Medicine
Washington, DC

Lisa M McShane, PhD
Biometric Research Program, Division of Cancer Treatment and Diagnosis

Disclaimers

- The views expressed represent my own and do not necessarily represent views or policies of the U.S. National Cancer Institute.
- Examples I cite are all true, but in some cases I have concealed details to protect identities.

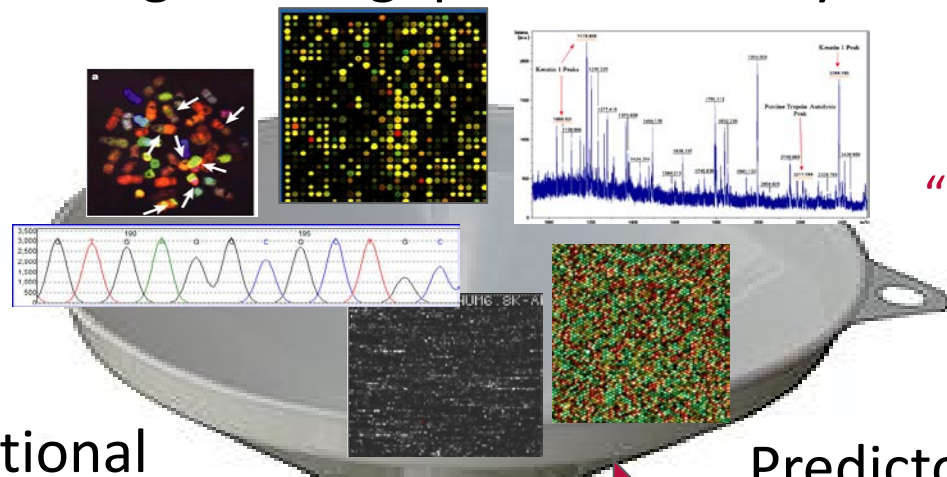
My perspective

- Statistical/scientific reviewer of NCI-sponsored clinical trials and studies for development and validation of biomarker-and omics-based tests
- Scientific Advisory Board (*Science Translational Medicine*) and Editorial Board (*BMC Medicine*)
- Statistical reviewer for numerous biomedical journals
- Statistical collaborator in research projects involving biomarkers and omics tests

Translation from omics discoveries to clinically useful omics-based tests

Discovery

High-throughput omics assays

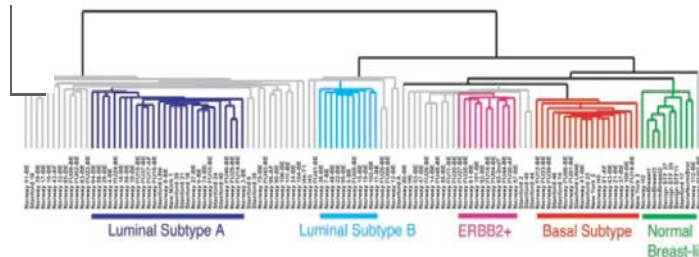
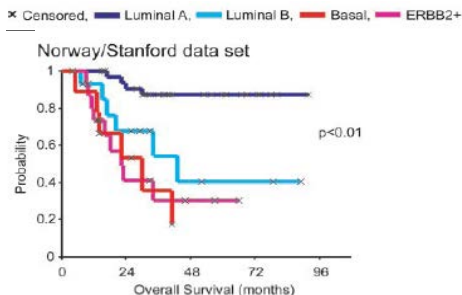


“Omics predictor”

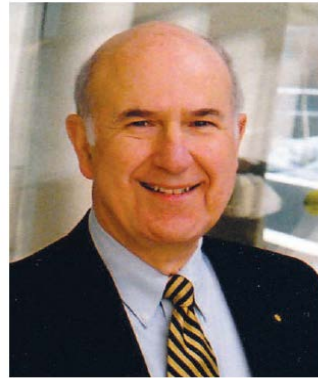
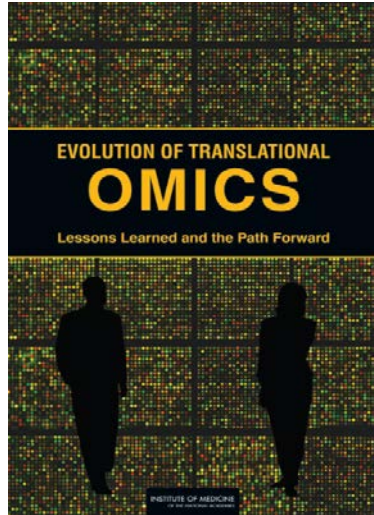
Computational models

Predictors, classifiers, risk scores

Clinical Utility?



Institute of Medicine reviews field of translational omics



"There are a lot of lessons here that surely apply to other places."

—GILBERT S. OMENN,
UNIVERSITY OF
MICHIGAN,
ANN ARBOR



"There are a lot of lessons here that surely apply to other places."

<http://www.iom.edu/Reports/2012/Evolution-of-Translational-Omics.aspx>

NCI criteria for the use of omics-based predictors in clinical trials

McShane et al. Nature 2013;502:317-320 (checklist)

McShane et al. BMC Medicine 2013;11:220 (explanation & elaboration)

5 DOMAINS

✓ Specimens

✓ Assays

} Getting all of the details right is hard work; important but often ignored in discovery stage

✓ Model development, specification & *preliminary* performance evaluation

} Too many researchers still fall into traps of high-dimensional data analysis

■ Clinical trial design

■ Ethical, legal, and regulatory issues

} No time to discuss today

Domain 1: Specimens

- Collection, processing & storage (**pre-analytic factors**)
- Specimen **quality** screening
- Minimum required **amount**
- **Enrichment** (e.g., micro- or macro-dissect for tumor cells)
- **Feasibility** of collecting needed specimens
 - Specimen collection (e.g., biopsy) is safe
 - Achievable in standard clinical settings
 - Percent useable for assays is sufficiently high

Domain 1: Specimen pre-analytic requirements and quality monitoring plans

- Example 1: Only 20% of first 100 specimens collected in a diagnostics study were of adequate quality for immediate assay due to failure to freeze immediately upon collection
- Example 2: Only 50% of frozen samples collected on a trial were fit for assay upon thawing a few years later

Moore et al. Biospecimen Reporting for Improved Study Quality (BRISQ). Cancer Cytopathology 2011;119:92-101

Domain 2: Assay considerations

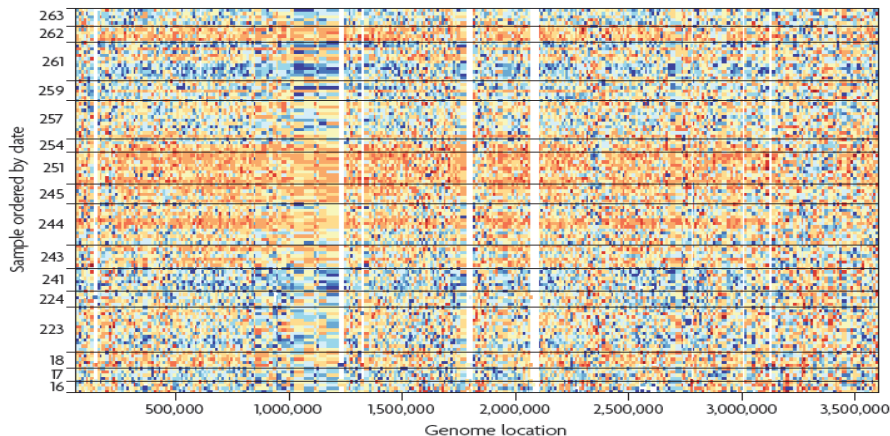
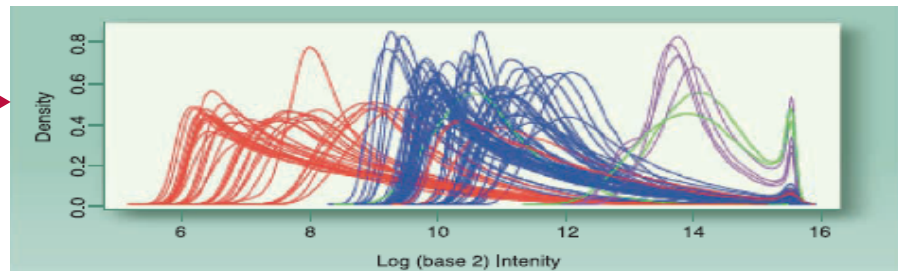
- Lock down standard operating procedures (SOPs)
- Impact of changes in assay procedures
- Quality monitoring (controls, standards)
 - Equipment malfunction, bad reagent lots, re-calibration needed
- Quality criteria to accept/reject assay values
 - Bad specimens, batch effects
- Analytical performance evaluation (sensitivity, specificity, bias, accuracy, precision, reproducibility)
 - Becker R, Analytical validation of in vitro diagnostic tests. In *Design and Analysis of Clinical Trials for Predictive Medicine*, Matsui, Buyse, Simon (eds.), Chapman and Hall/CRC, 2015, pp. 33-50.
 - Jennings et al, *Arch Pathol Lab Med* 2009;133: 743–755

Domain 2: Assay artifacts & batch effects

(Owzar et al, *Clin Cancer Res* 2008;14:5959-5966)

Density estimates of PM probe intensities (Affymetrix CEL files) for 96 NSCLC specimens

Red = batch 1, Blue = batch 2
Purple & Green = outliers?



Batch effects for 2nd generation sequence data (std. coverage data). Same facility & platform.

Horizontal lines divide by date
(Leek et al, *Nature Rev Genet* 2010;11:733-739)

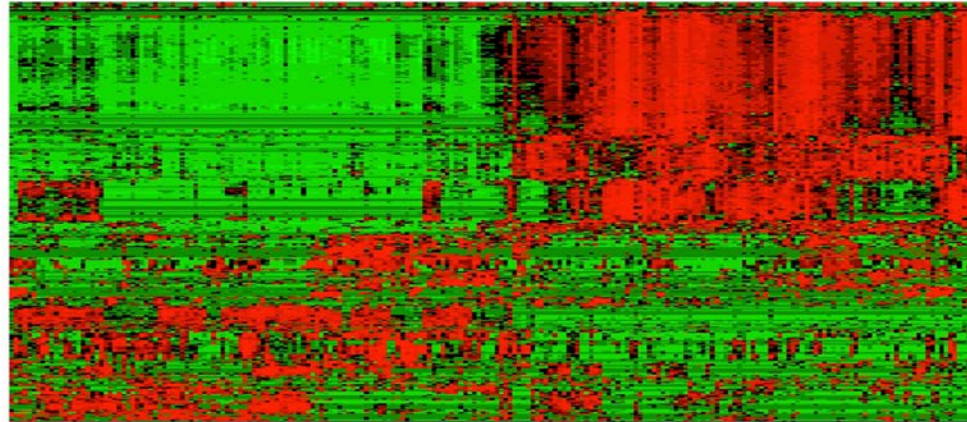
BATCH EFFECTS ARE ESPECIALLY PROBLEMATIC IF CONFOUNDED WITH KEY EXPERIMENTAL FACTORS OR ENDPOINTS.

Domain 2: Assay artifacts & batch effects

- Impact of changes in assay procedures, reagents, equipment, or technician during predictor development

Dramatic effect of change in RNA extraction procedure & reagents on tumor gene expression microarray profiles

Extraction method 1 | Extraction method 2



116 genes
included in a
genomic
predictor of
treatment
response

(Shown with
permission from
an NIH grantee)

215 tumor samples

Domain 2: Assess impact of changes in assay reagents or procedures before implementing

MINDACT Trial

Cardoso F et al., *N Engl J Med*
2016;375:717-729



70-Gene Signature as an Aid to Treatment Decisions in Early-Stage Breast Cancer

F. Cardoso, L.J. van't Veer, J. Bogaerts, L. Slaets, G. Viale, S. Delaloge, J.-Y. Pierga, E. Brain, S. Causeret, M. DeLorenzi, A.M. Glas, V. Goulinopoulos, T. Goulioti, S. Knox, E. Matos, B. Meulemans, P.A. Neijenhuis, U. Nitz, R. Passalacqua, P. Ravdin, I.T. Rubio, M. Saghatelyan, T.J. Smit, C. Sotiriou, L. Stork, C. Strahle, G. Thomas, A.M. Thompson, J.M. van der Hoeven, P. Vuysteke, R. Bernards, K. Tryfonidis, E. Rutgers, and M. Piccart, for the MINDACT Investigators*

“A **change in the RNA-extraction solution** that was used in the calculation of the 70-gene signature (a change that was not communicated by the manufacturer) **caused a temporary shift in the risk calculation** from May 24, 2009, to January 30, 2010, at which time the issue was discovered and rectified . . .

Because of this shift, **162 patients who had been identified as being at high genomic risk were subsequently identified as being at low genomic risk** with the use of the correct . . .

The clinical effect of this risk revision was that an **additional 28 patients received chemotherapy before the results were corrected, although no patient was undertreated.**”

Domain 3: Model development & preliminary evaluation

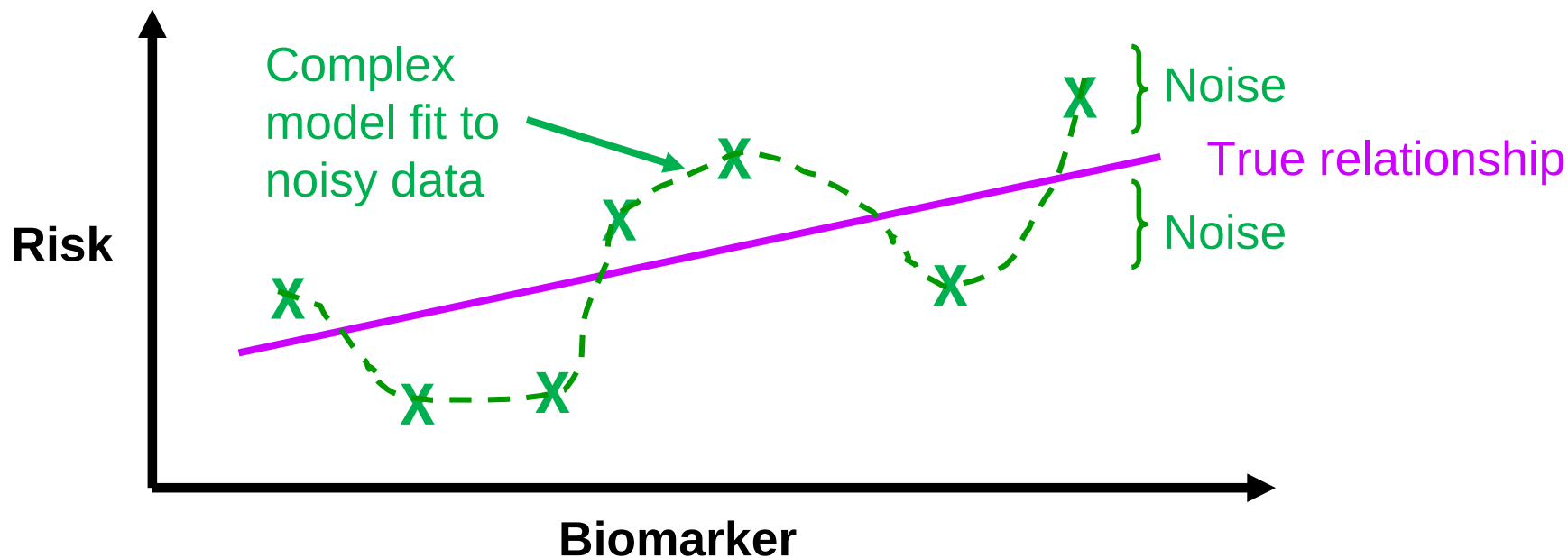
- **Quality of data** (clinical & omics) used to develop and validate predictor models (might not be “clinical trials grade” data)
- **Appropriate statistical approaches** for model development and performance assessment
- **Appropriate “validation”**
 - Define clinical context and use
 - Patient population
 - Clinical use - prognostic, predictive, etc.
 - “Locked down” test
 - Pre-specified evaluation criteria (not just a significant p-value)

Domain 3: Model development & preliminary evaluation common pitfalls

- A statistical model is **OVERFIT** when it describes random error or noise instead of the true underlying relationship
 - Excessively complex (too many parameters or predictor variables)
 - Will have poor predictive performance on independent data set
 - Naively fit omics predictors will always be overfit
- **RE-SUBSTITUTION** is the naïve evaluation of model performance by “plugging in” same data used to build it
 - Other more subtle forms of re-substitution (combining training & test, with covariates, comparative, partial)

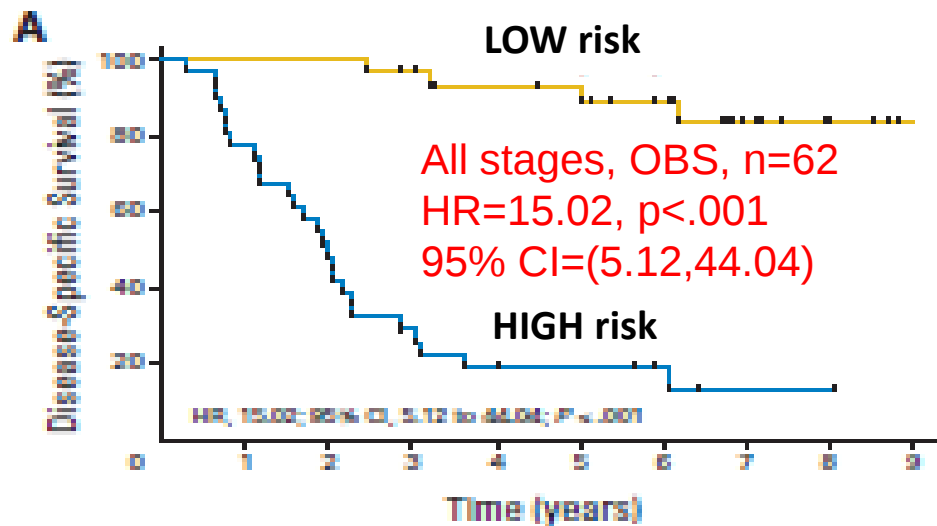
(J Biopharm Statistics 2016;26(6):1098-1110)

Domain 3: Model over-fitting



- Evaluation of a model's fit by data re-substitution will suggest fit is perfect
- In high dimensions (e.g., omics data), naively fit models are almost always over-fit and such models will rarely validate on an independent data set

Domain 3: Avoid “re-substitution” pitfall



(J Clin Oncol 2010;28:4417-4424)

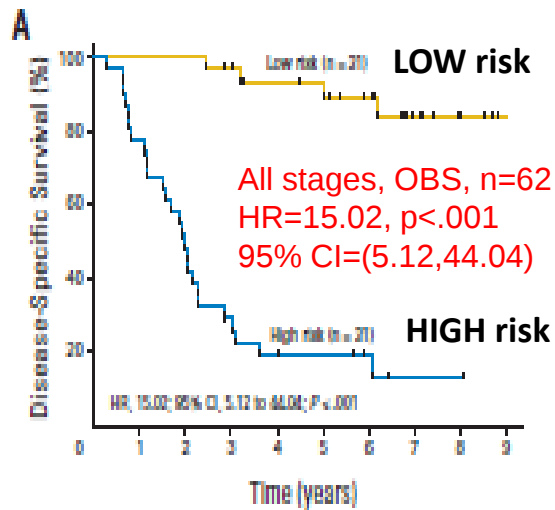
“A 15-gene signature [for lung cancer] separated OBS patients [no chemotherapy after surgery] into high-risk and low-risk subgroups with significantly different survival (hazard ratio [HR], 15.02; 95% CI, 5.12 to 44.04; $P < .001$.”



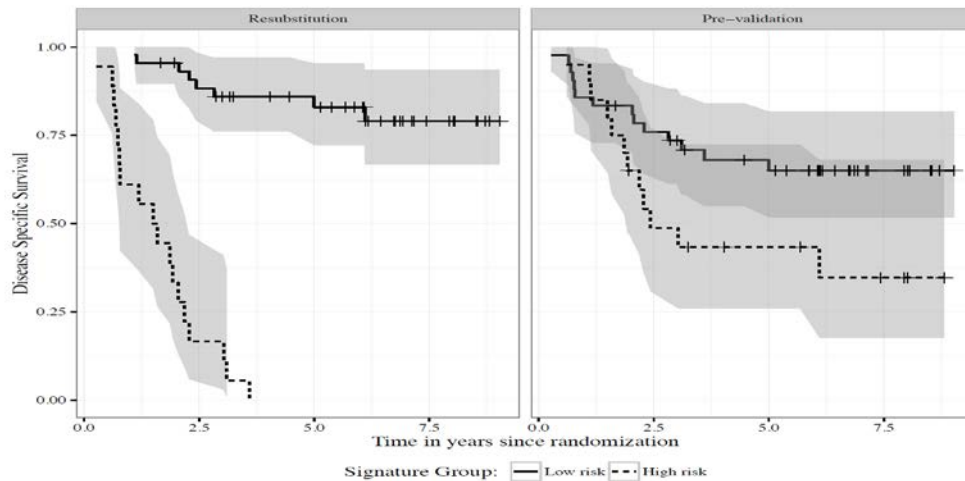
RE-SUBSTITUTION!

If this large separation in survival curves was real, the signature would have clinical utility. Patients designated as low risk could confidently avoid toxic chemotherapy.

Domain 3: Internal validation (e.g., cross-validation) to correct for re-substitution bias



Original Kaplan-Meier curves (DSS) showing prognostic ability of 15-gene signature in OBS arm, using re-substitution (J Clin Oncol 2010;28:4417-4424)

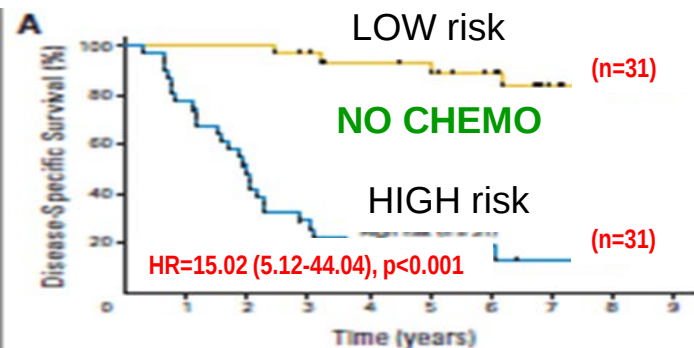


RE-SUBSTITUTION

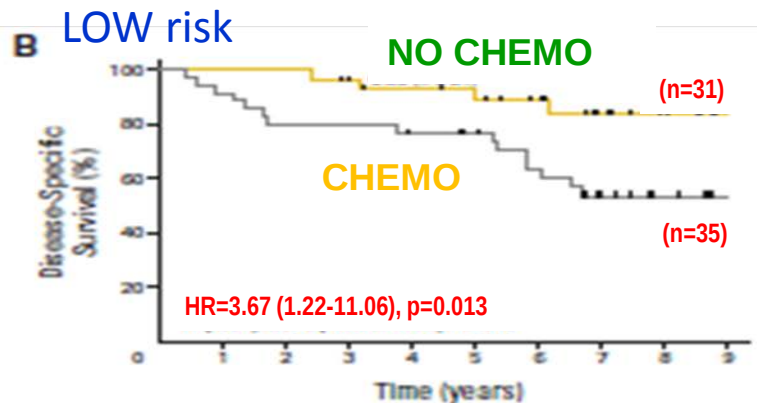
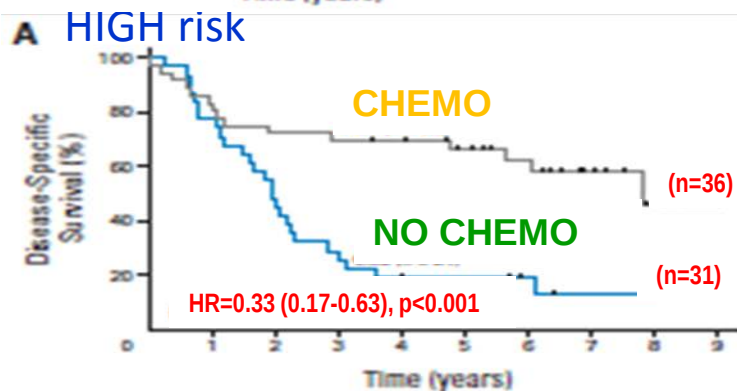
CROSS-VALIDATION

Reproduced (approx.) Kaplan-Meier curves (DSS) showing prognostic ability of 15-gene signature in OBS arm, using re-substitution (LEFT) and cross-validation (RIGHT) (J Biopharm Statistics 2016;26(6):1098-1110)

Domain 3: *Avoid comparisons with re-substitution estimates*



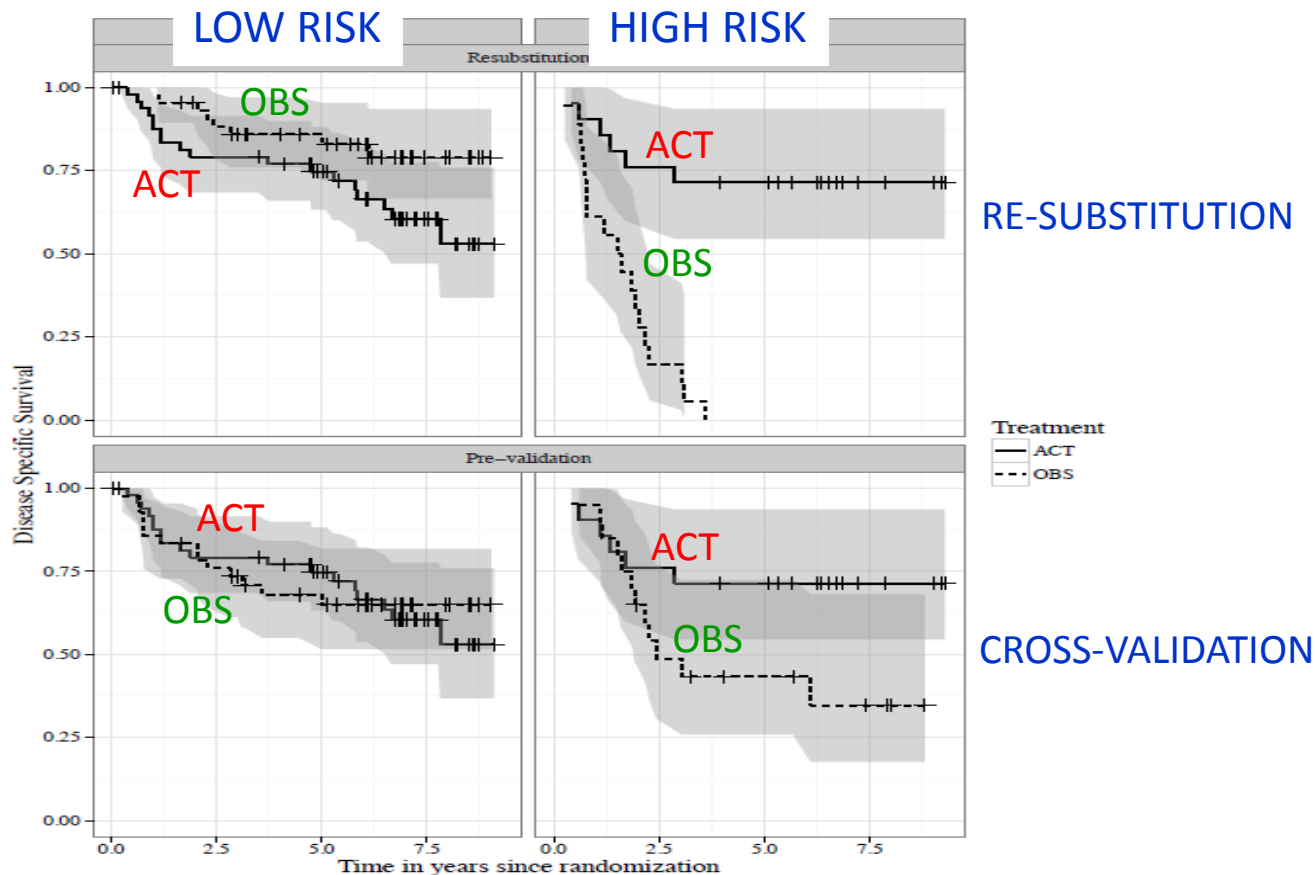
Prognostic classifier fit using gene expression microarray data from clinical trial arm on which patients received **no adjuvant chemotherapy** (re-substitution)



Does the genomic predictor identify groups of patients who benefit differently from adjuvant chemotherapy? **Can't conclude anything.**

Domain 3: Internal validation (e.g., resampling)

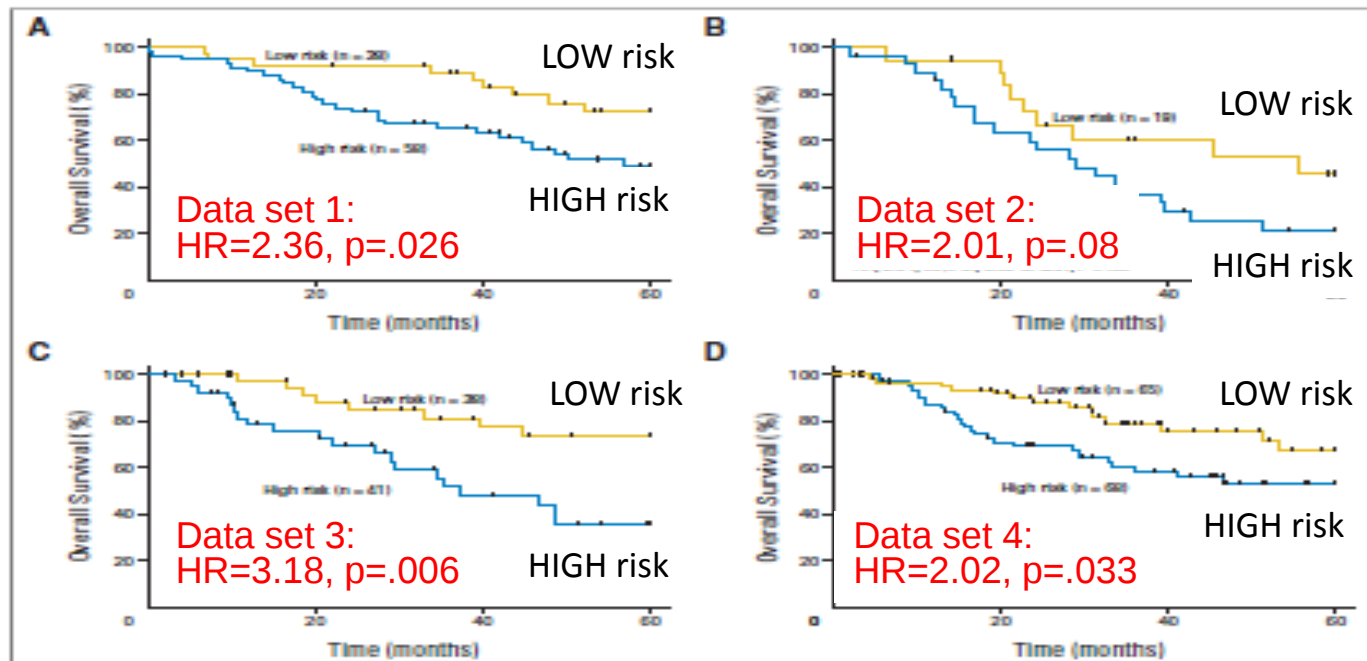
Use of cross-validation to correct for re-substitution bias



Reproduced (approx.)
Kaplan-Meier curves
(DSS) suggesting
predictive ability (for
treatment-selection) of
15-gene signature
using re-substitution
(TOP) and cross-
validation (BOTTOM)

(J Biopharm Statistics
2016;26(6):1098-1110)

Domain 3: What do we mean by validation?



(J Clin Oncol 2010;28:4417-4424)

“ . . . prognostic effect [of 15-gene signature] was **VALIDATED** consistently in four separate microarray data sets (total 356 stage IB to II patients without adjuvant treatment).”

Endpoint: Disease-specific survival (DSS) → Overall survival (OS)

Timescale: 0 to 9 years → 0 to 60 months (5 years)

HR: 15.02 → $\approx 2-3$ 5-yr DSS $\approx 90\%$ → 5-yr OS $< 80\%$

Mixture of disease stages? Adjustment for standard covariates?

Domain 3: What do we mean by validation?

- “Genetic Basis for Clinical Response to CTLA-4 Blockade in Melanoma”
- Original article: “We elucidated a neoantigen landscape that is specifically present in tumors with a strong response to CTLA-4 blockade. We **validated** this signature in a second set of 39 patients with melanoma who were treated with anti-CTLA-4 antibodies.”
- Correction: “Some readers were confused by our incomplete description of part of the data analysis and our use of the term “validation set . . . **our use of “validation set” was not appropriate** in the context of the search for a neoantigen signature, since information from both data sets was used to derive the results.. . we **did not use “validation set” in the conventional way. In contrast to a formal biomarker analysis, our study design focused on defining a recurrent genetic footprint that occurred in a nonrandom fashion.** (Several other corrections made as well.)

Domain 3: Requirements for a rigorous validation of a predictor

- Predictor must be completely **LOCKED DOWN** and must be a **PRE-SPECIFIED PERFORMANCE METRIC**. The lockdown includes *all steps* in the data pre-processing and prediction algorithm (including computer code).
- Ideally, **INDEPENDENT VALIDATION DATA** generated from specimens collected at a different time, or in a different place, and according to the pre-specified collection protocol.
- Assays for the validation specimen set should be run at a different time or in a different laboratory according to the **PRE-SPECIFIED ASSAY** protocol (including quality rejection criteria).

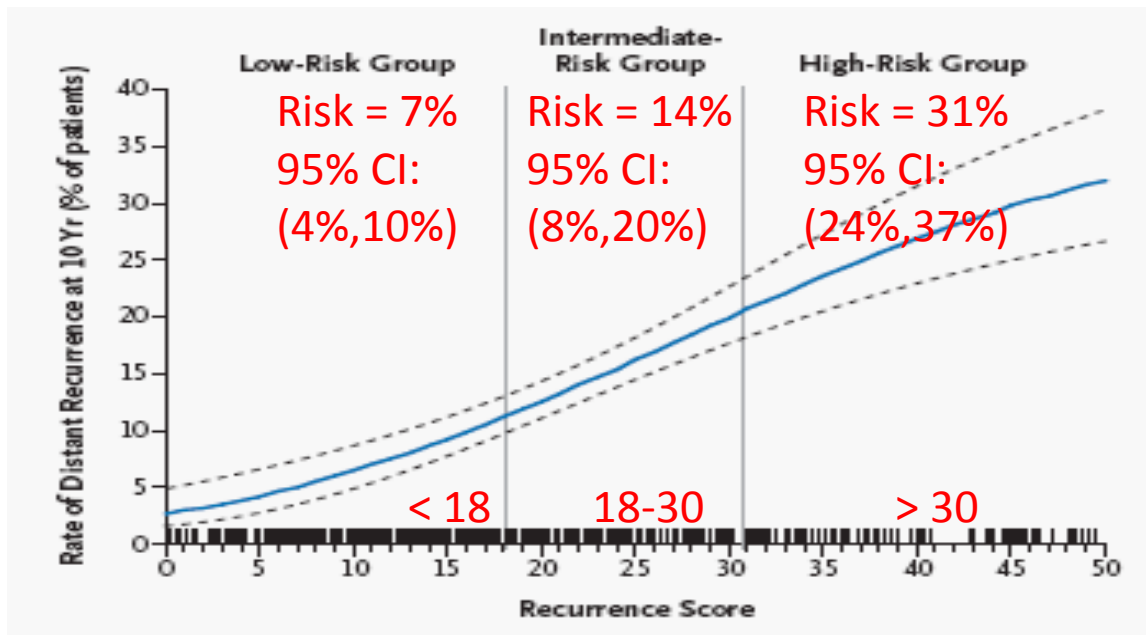
(cont →)

Domain 3: Requirements for rigorous validation of a predictor (cont.)

- Individuals who developed the predictor must remain completely **BLINDED** to the validation data.
- The validation **DATA SHOULD NOT BE CHANGED** and **DATA VALUES SHOULD NOT BE SELECTIVELY ELIMINATED** after observing the performance of the predictor.
- **PREDICTOR SHOULD NOT BE ADJUSTED** (including cut-points) after its performance has been observed on any part of the validation data. Otherwise, the validation is compromised and a new validation may be required.

SUCCESS STORY (part 1): Rigorous validation of Oncotype DX recurrence score (RS) prognostic ability

Prospective-Retrospective Validation of RS on NSABP B-14
Tamoxifen Arm (tamoxifen treated ER+ breast cancer)

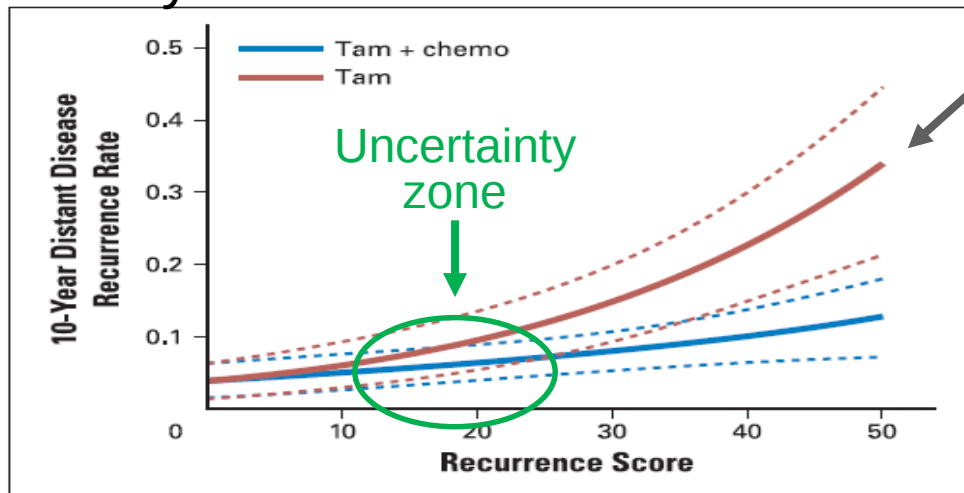


- 21-gene RT-PCR based gene expression assay
- FFPE tissues
- Locked assay and RS algorithm
- RS assignments blinded to outcome data
- “Honest broker” linked final RS to clinical outcome data

(Figure 4 from Paik et al., N Engl J Med 2004;351:2817-26)

SUCCESS STORY (part 2): Is Oncotype DX predictive for chemotherapy benefit?

Preliminary Evaluation of Predictive Ability of RS on NSABP B-20

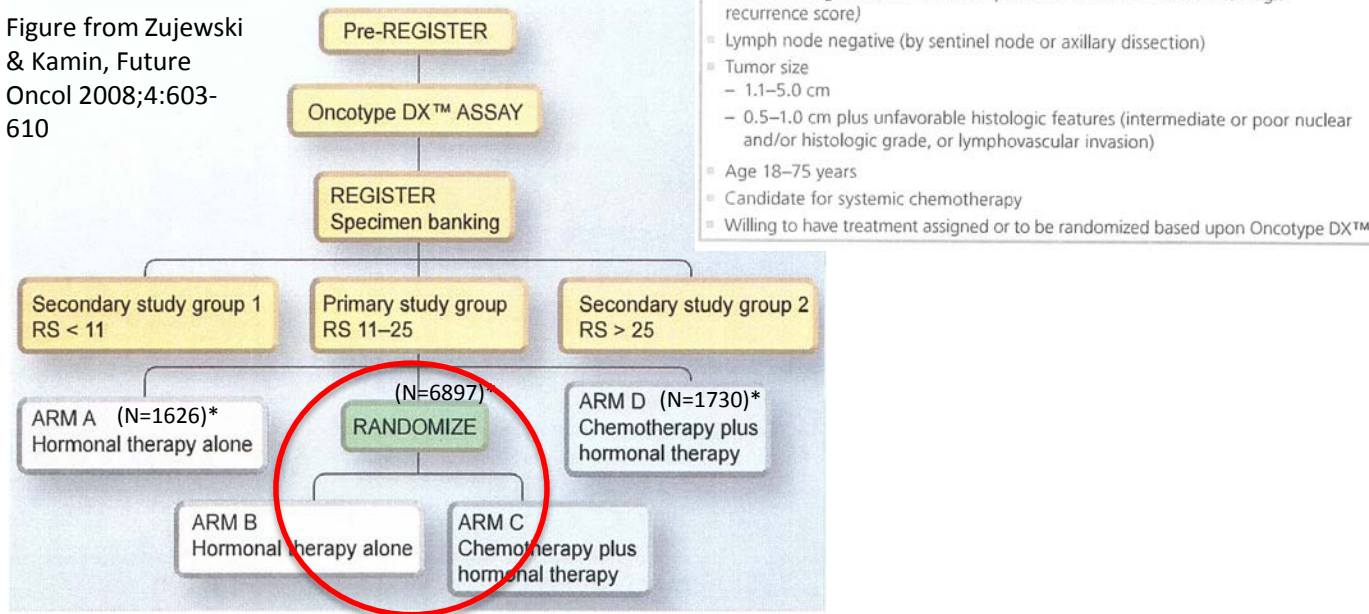


- Possible bias due to use of Tam arm in RS development?
- Few events in the intermediate zone (imprecise estimate of chemo benefit)
- Impact of newer endocrine therapies (e.g., AIs)

This study alone was considered insufficient to establish clinical utility of Oncotype DX for **therapy selection (predictive ability)**.

TAILORx trial design

Figure from Zujewski
& Kamin, Future
Oncol 2008;4:603-
610

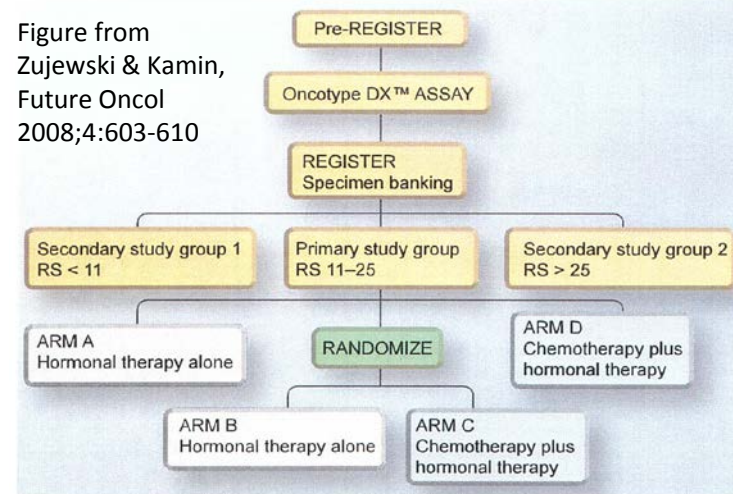


The TAILORx trial was designed to establish whether Oncotype DX RS has clinical utility for selection of which patients with **node negative hormone receptor-positive breast cancer** benefit from receiving chemotherapy in addition to endocrine therapy (predictive ability).

TAILORx: Validation of predictive ability of Oncotype DX RS

9719 eligible patients with follow-up information, 6711 (69%) had a midrange recurrence score of 11 to 25 and were randomly assigned to receive either chemoendocrine therapy or endocrine therapy alone.

Figure from
Zujewski & Kamin,
Future Oncol
2008;4:603-610



RS Group, therapy	5-yr Invas. DFS (%)	9-yr Invas. DFS (%)
RS ≤ 10, Endocrine	94.0 ± 0.6	84.0 ± 1.3
RS 11-25, Endocrine	92.8 ± 0.5	83.3 ± 0.9
RS 11-25, Chemoendocrine	93.1 ± 0.5	84.3 ± 0.8
RS ≥ 26, Chemoendocrine	87.6 ± 1.0	75.7 ± 2.2

Sparano et al, *N Engl J Med* 2015;373:2005-2014

Sparano et al, *N Engl J Med* 2018;379:111-21.

Summary remarks

- We now have several examples of successful omics-based tests.
- Still a need for education about best practices.
- Successful development and clinical translation takes the right expertise, time, and resources.
- For discussion sessions: How can we translate omics to clinically useful tests more effectively (highest positive impact for patients) and more efficiently?

THANK YOU!



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