

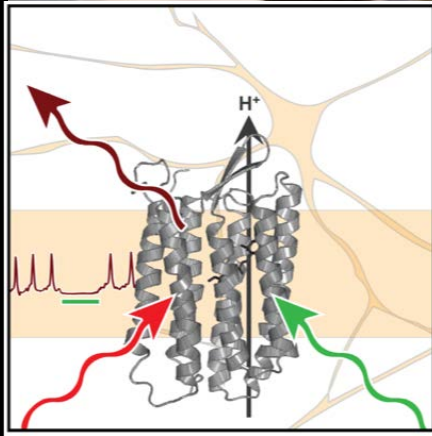
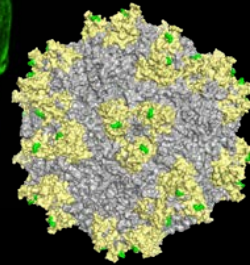
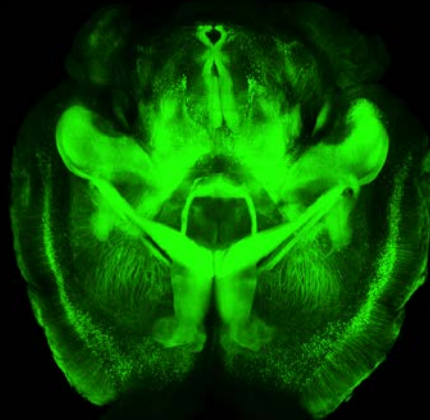
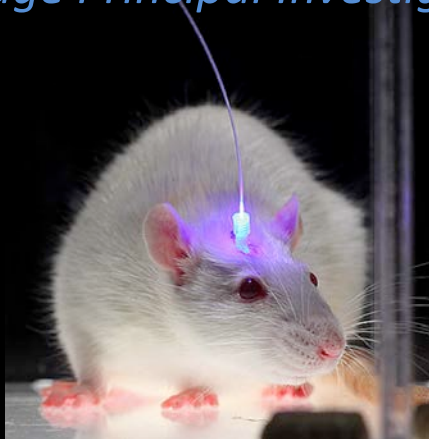
Crossing the Blood-Brain-Barrier for Brain-Wide Anatomical and Functional Mapping with novel AAVs and CLARITY

Viviana Gradinaru, CALTECH

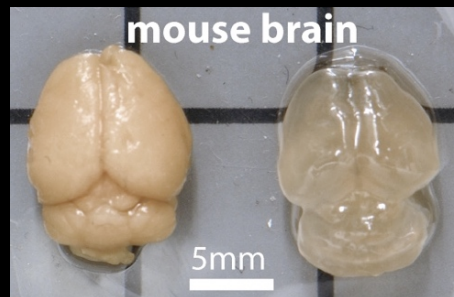
Director of the Caltech's Center for Molecular and Cellular Neuroscience

Assistant Professor of Biology and Biological Engineering

Heritage Principal Investigator



Optogenetics

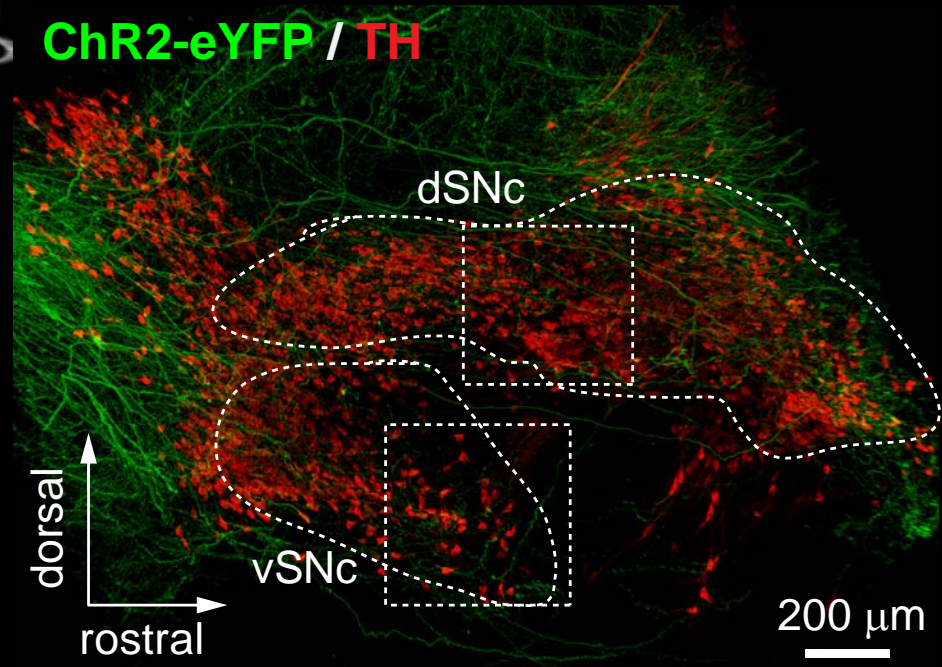
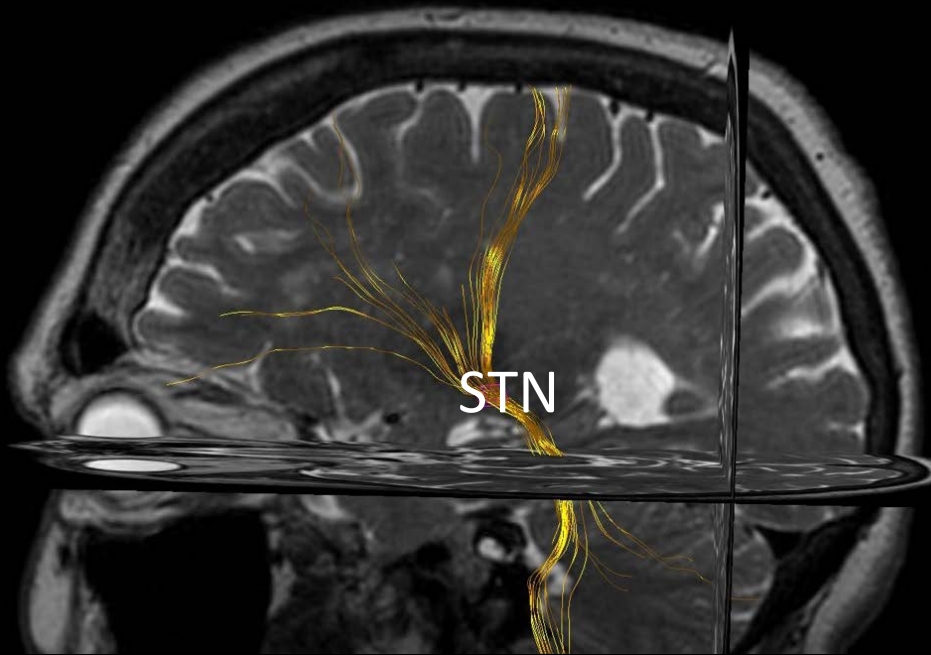


**CLARITY &
delivery vectors**

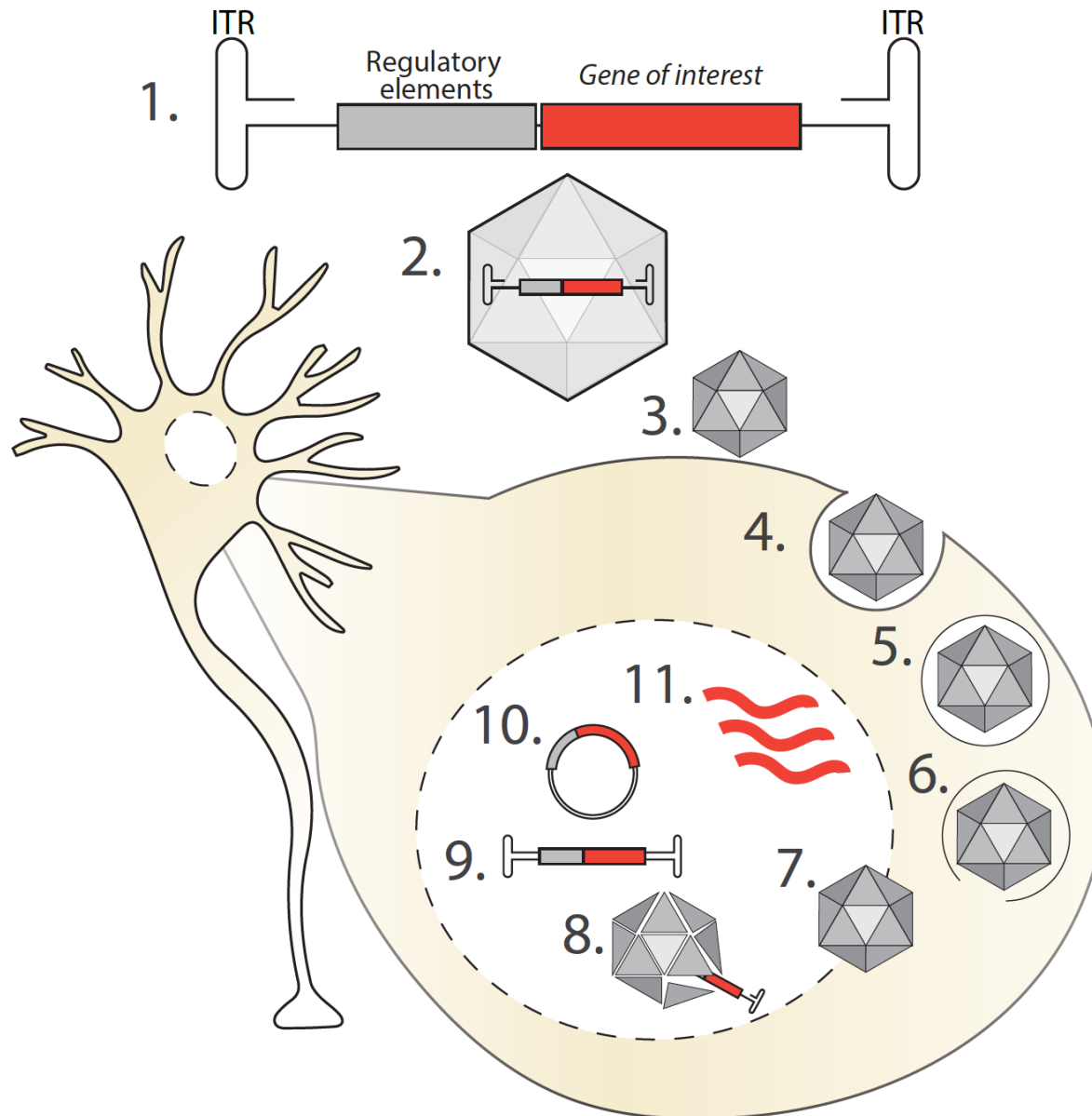


Deep Brain Stimulation

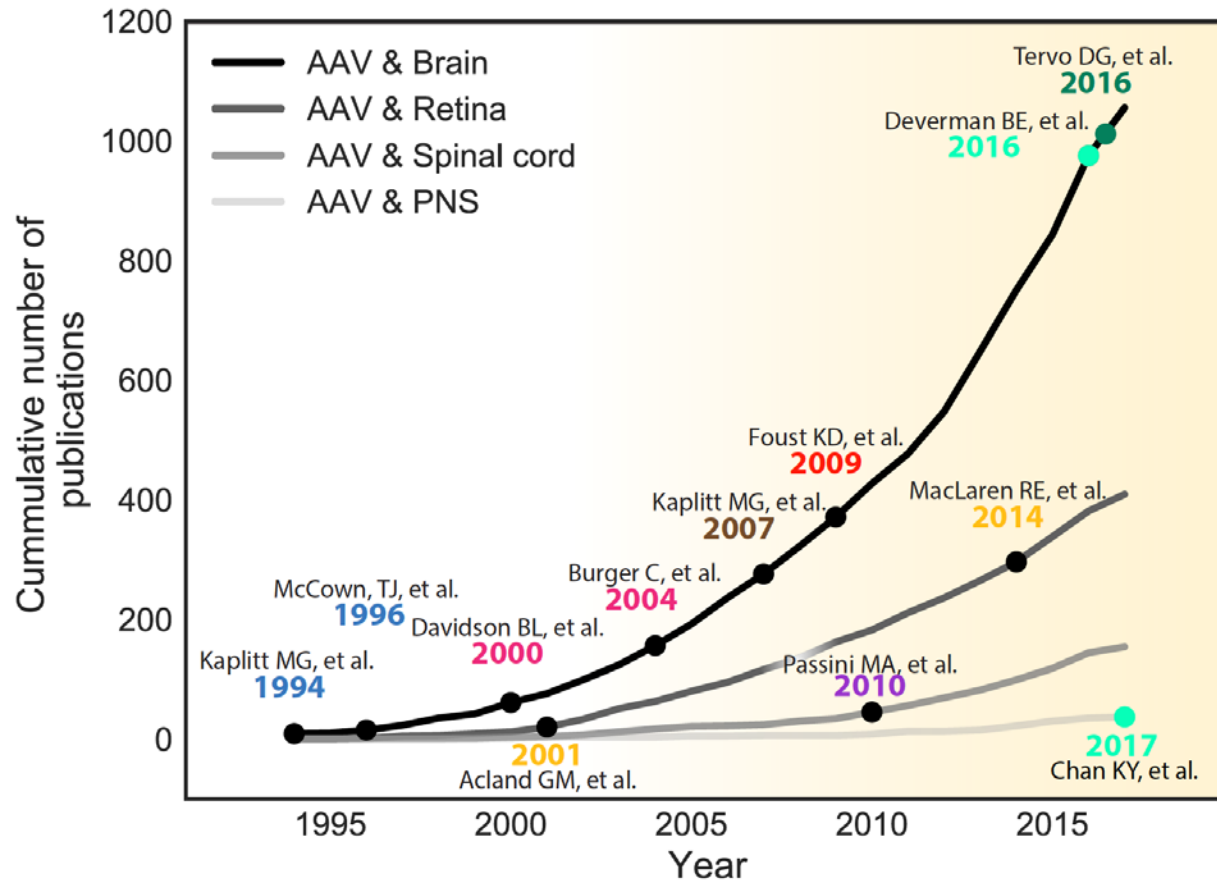
Delivering Genes across the Blood-Brain-Barrier and Mapping Projections Brain-Wide: AAV engineering and Tissue Clearing



rAAV transduction of a neuron



Overview of AAV use in the nervous system



1. First demonstrations of rAAVs in the brain

2. Comparison of rAAV serotypes' gene transfer, tropism, & distribution in the CNS

3. Phase I trial: AAVs for Parkinson's disease

4. CNS-wide transduction via systemic injection of AAV in neonatal mice

5. Retinal gene therapy

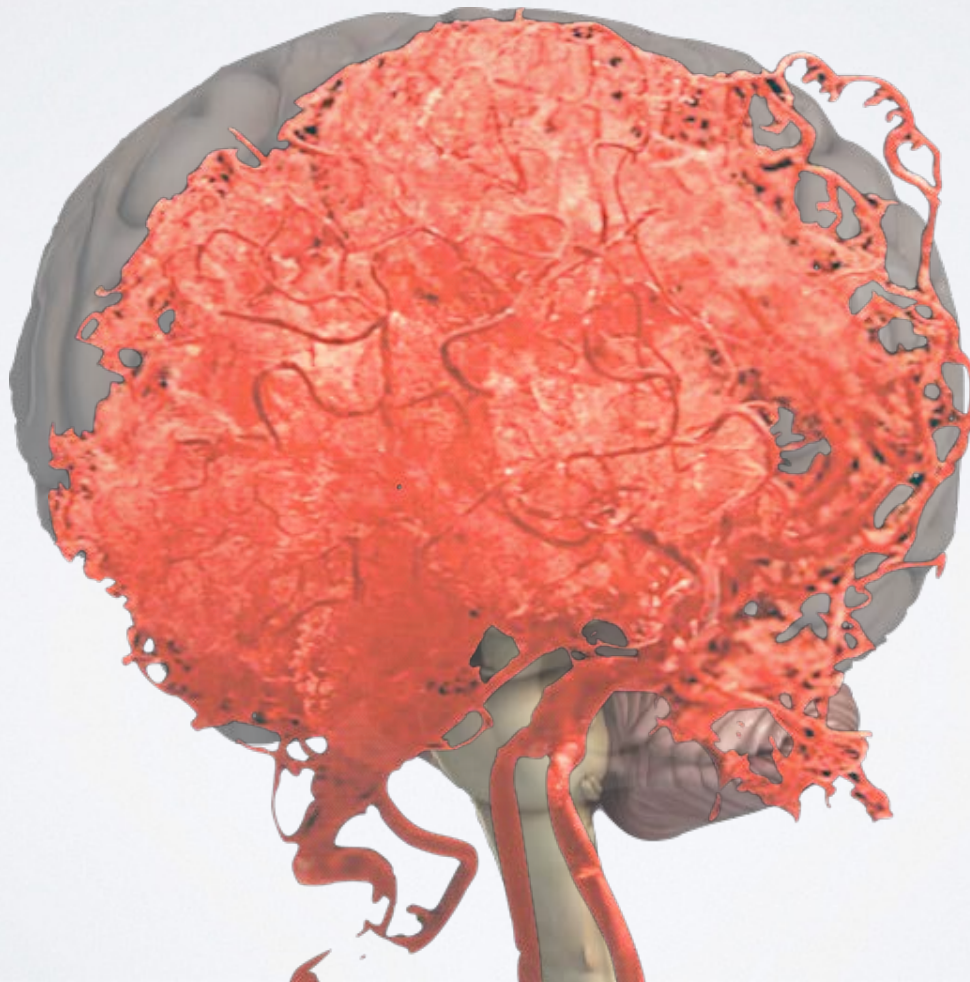
6. Spinal cord gene therapy

7. Designer rAAVs for systemic delivery to the CNS and PNS: rAAV-PHP.B, PHP.eB, & PHP.S

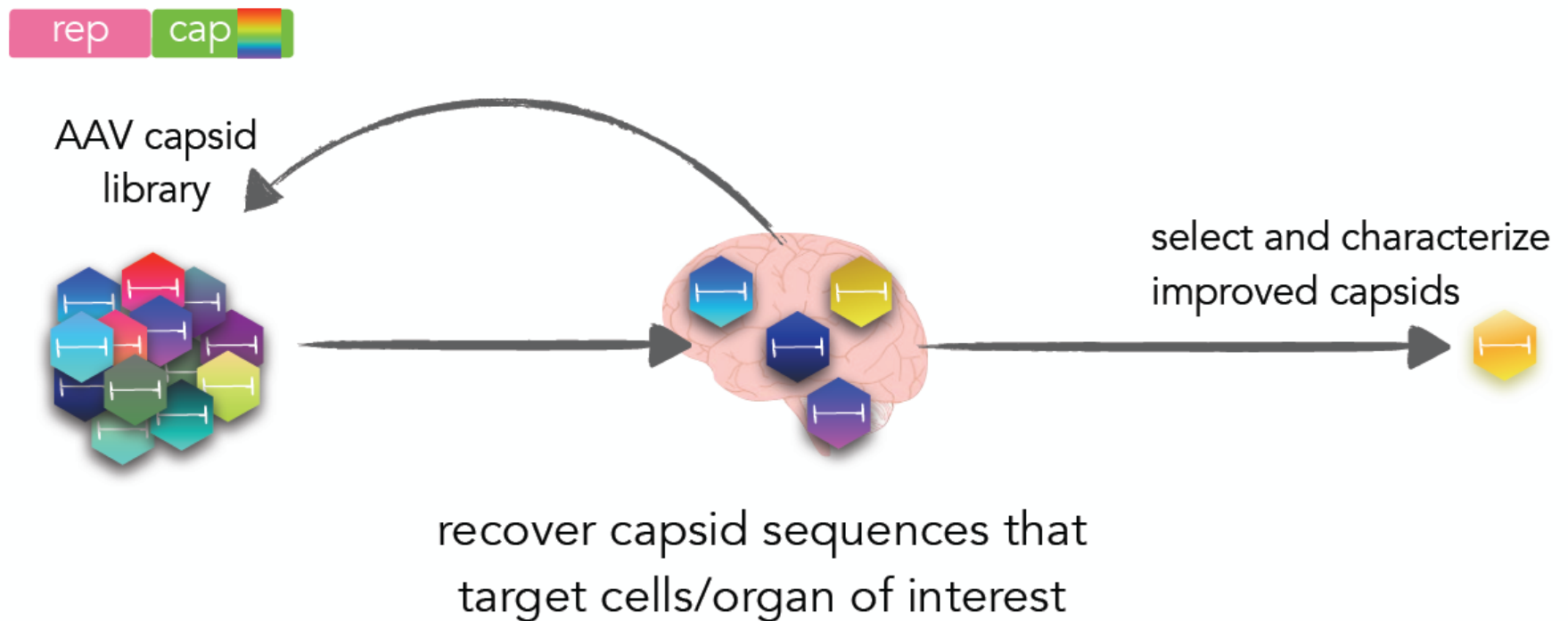
8. Designer rAAV for efficient retrograde trafficking: rAAV2-retro

Engineering Vectors for Efficient Delivery of Therapeutics to the Whole Brain

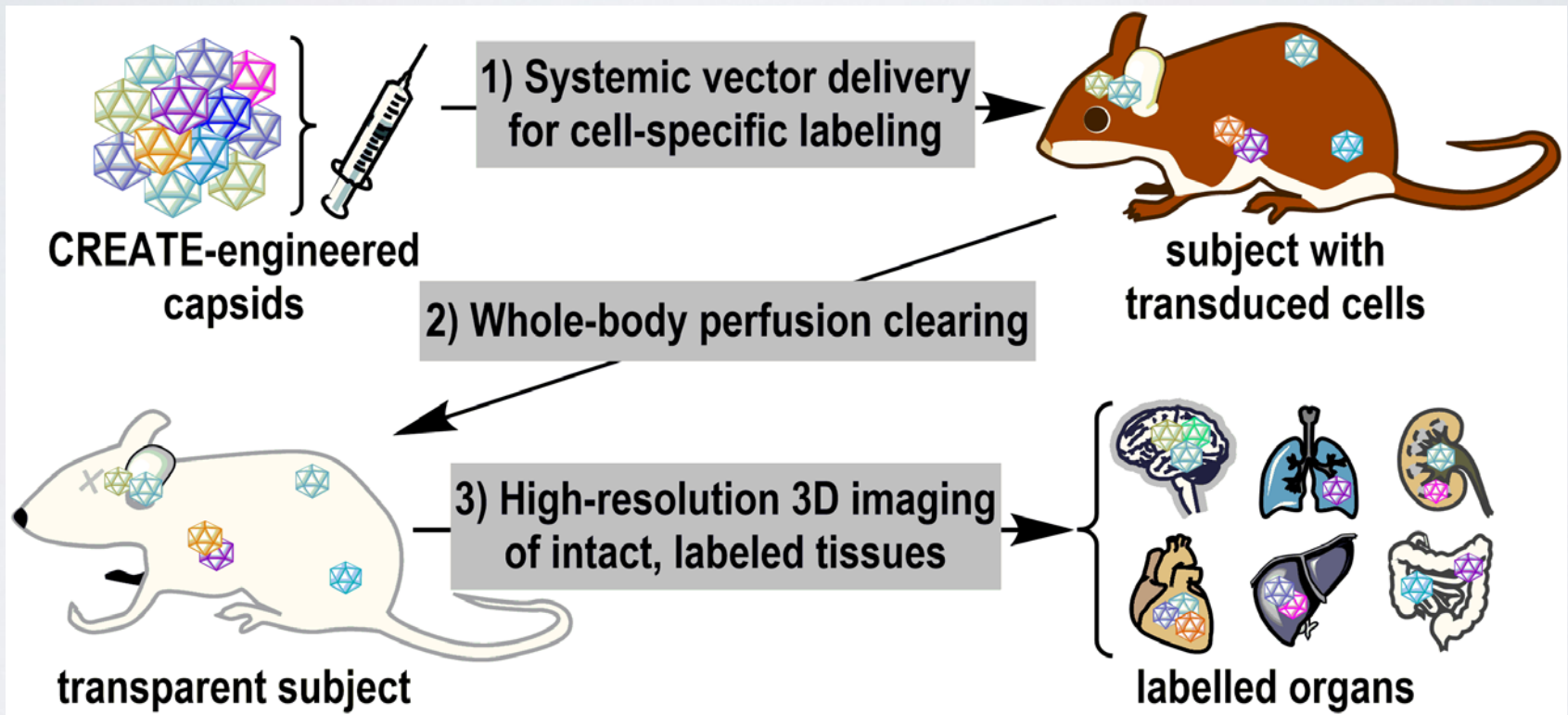
Agents of interest: neuroprotective growth factors, labels for circuit anatomy to inform DBS



In vivo selection for AAV capsids with more desirable properties

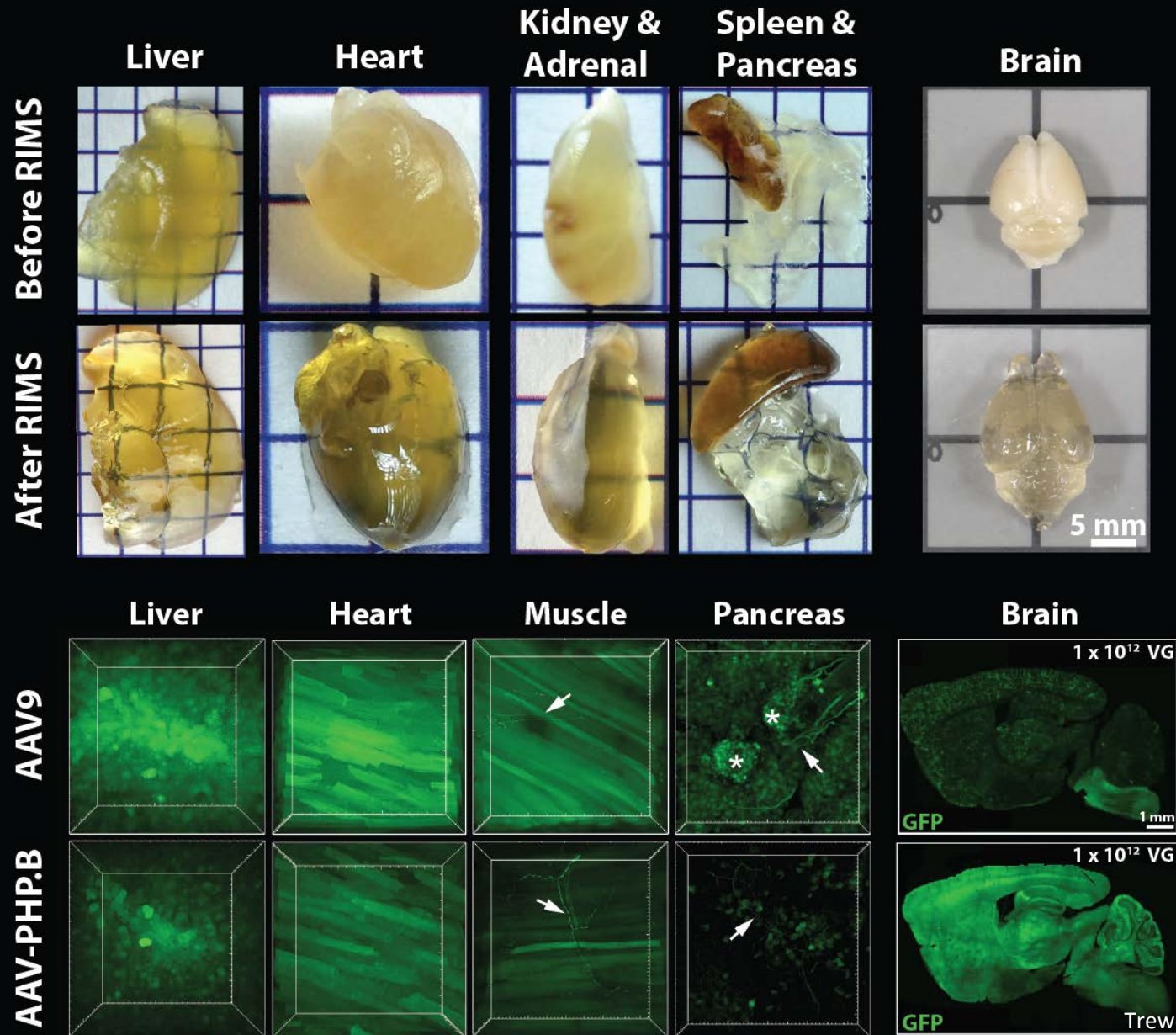


Whole-body PARS for viral biodistribution



PARS = Perfusion-Assisted Agent Release in Situ

Whole-body tissue clearing by PARS for viral biodistribution

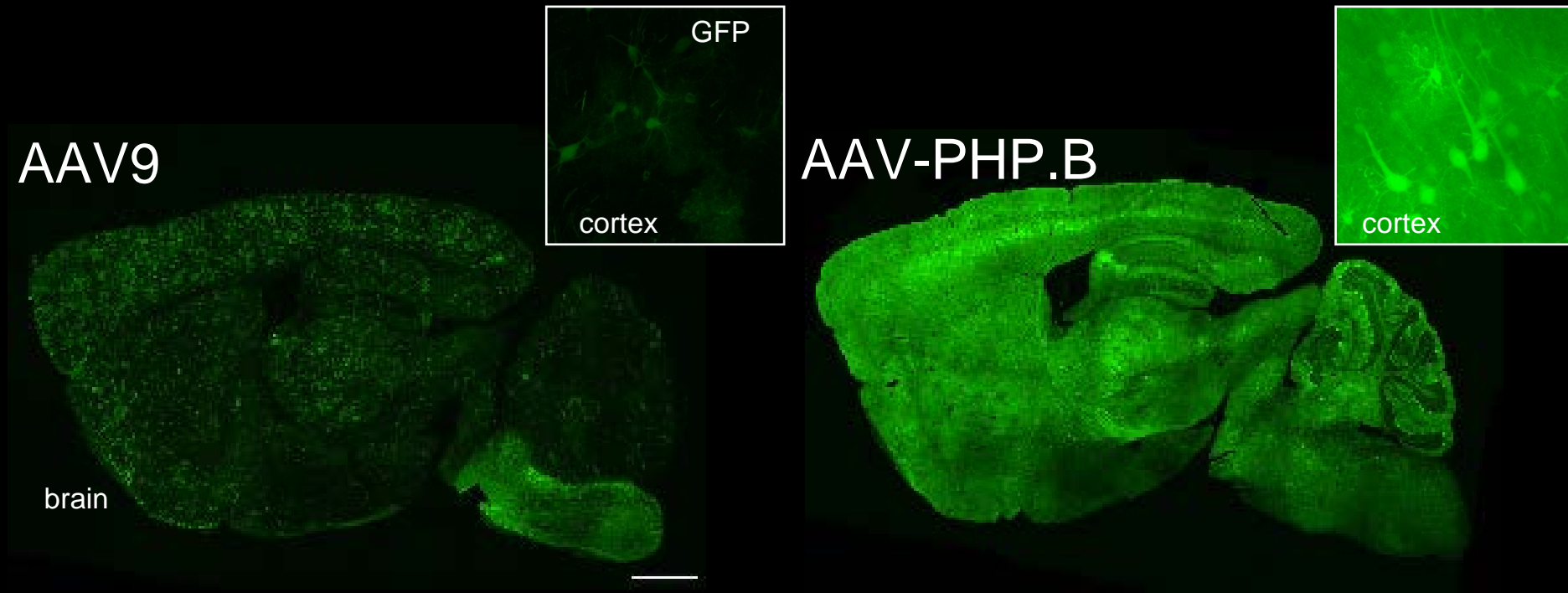


Yang *et al.* (Cell, 2014)

Treweek *et al.* (Nat.Prot, 2015)

Deverman *et al.* (Nat.Biotech, 2016)

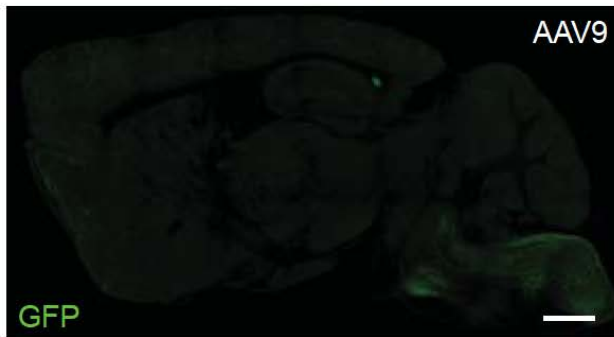
Brain-wide gene transfer via systemic delivery in adult rodent



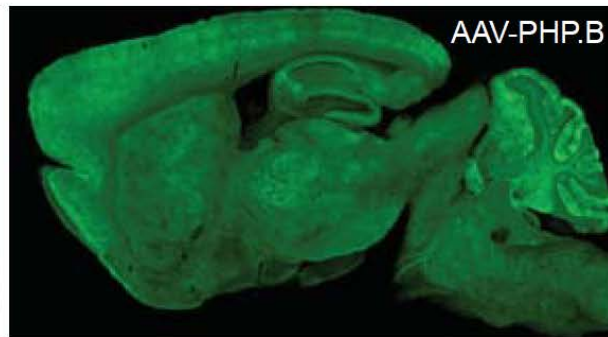
Long Lasting Expression

377 days post-injection

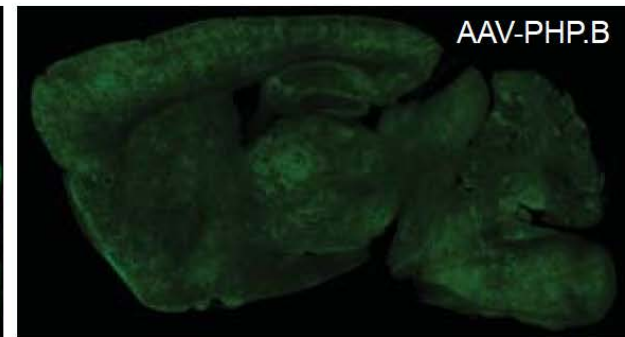
1×10^{12} vg



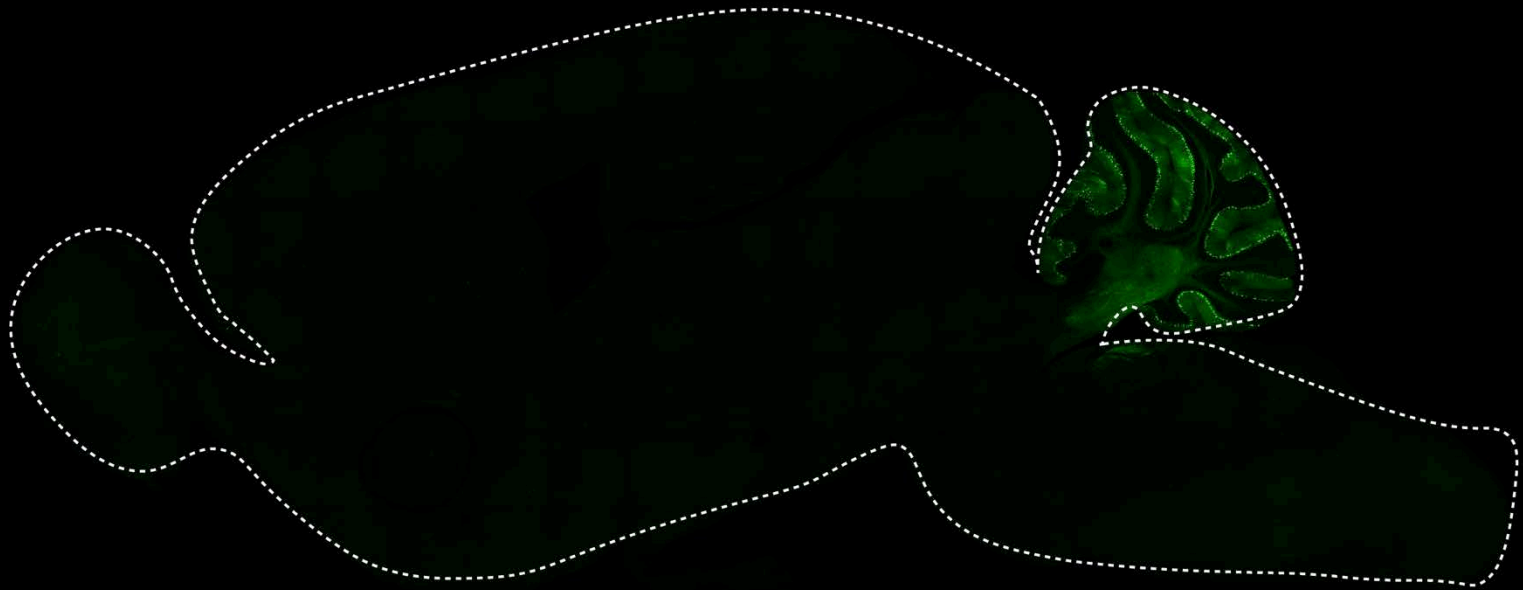
1×10^{12} vg



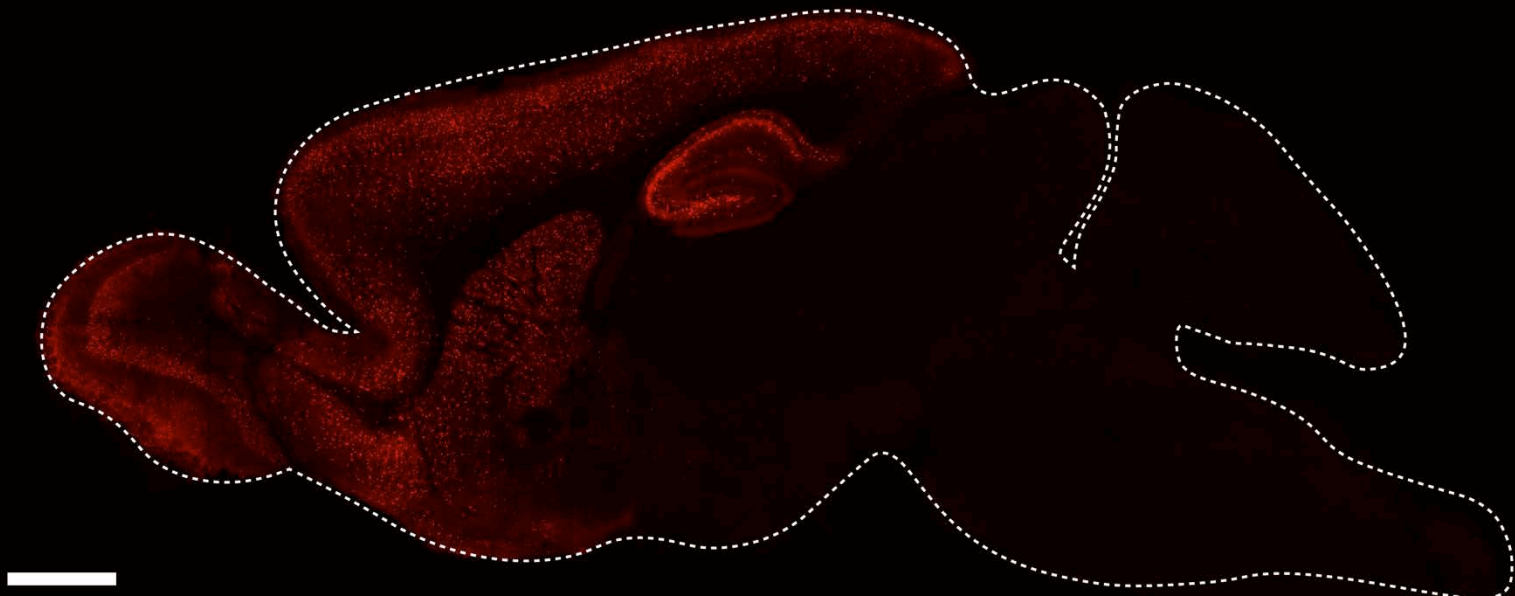
1×10^{11} vg



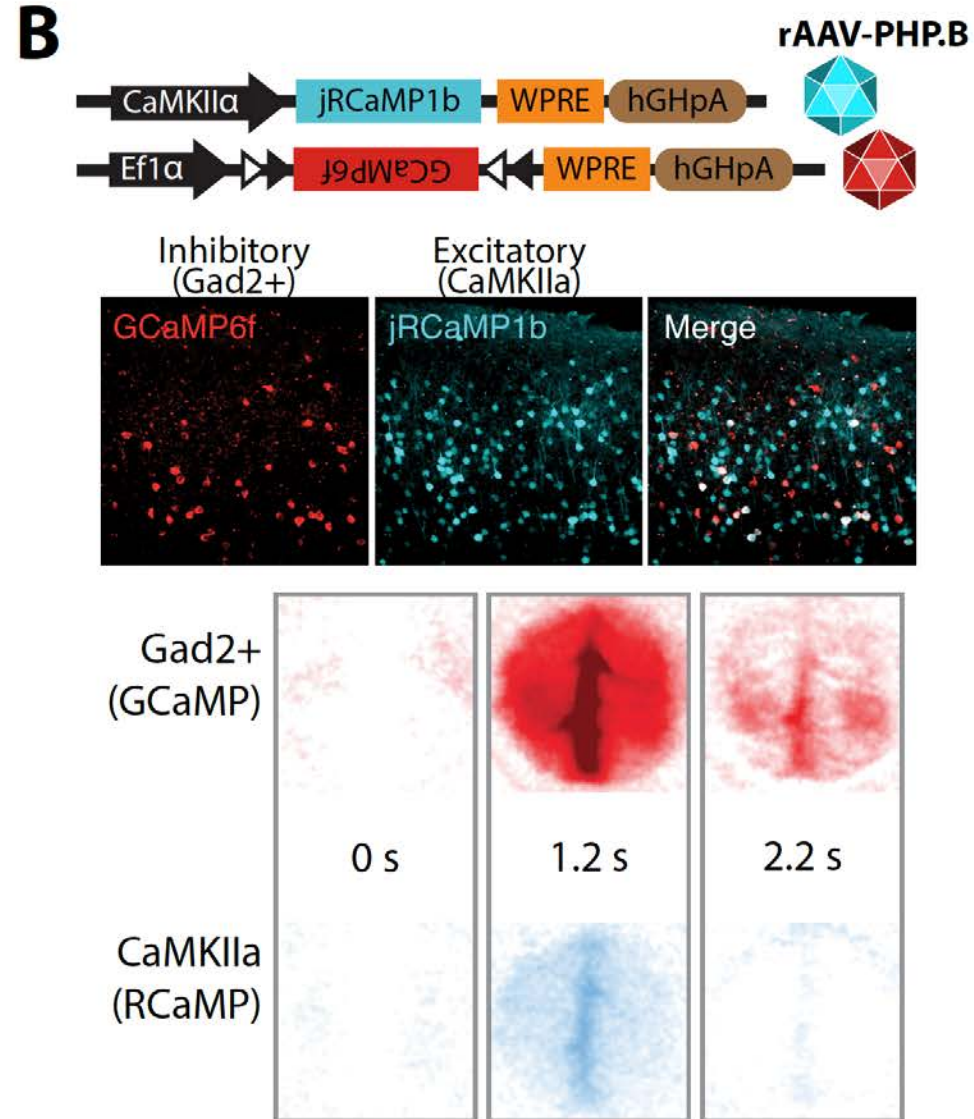
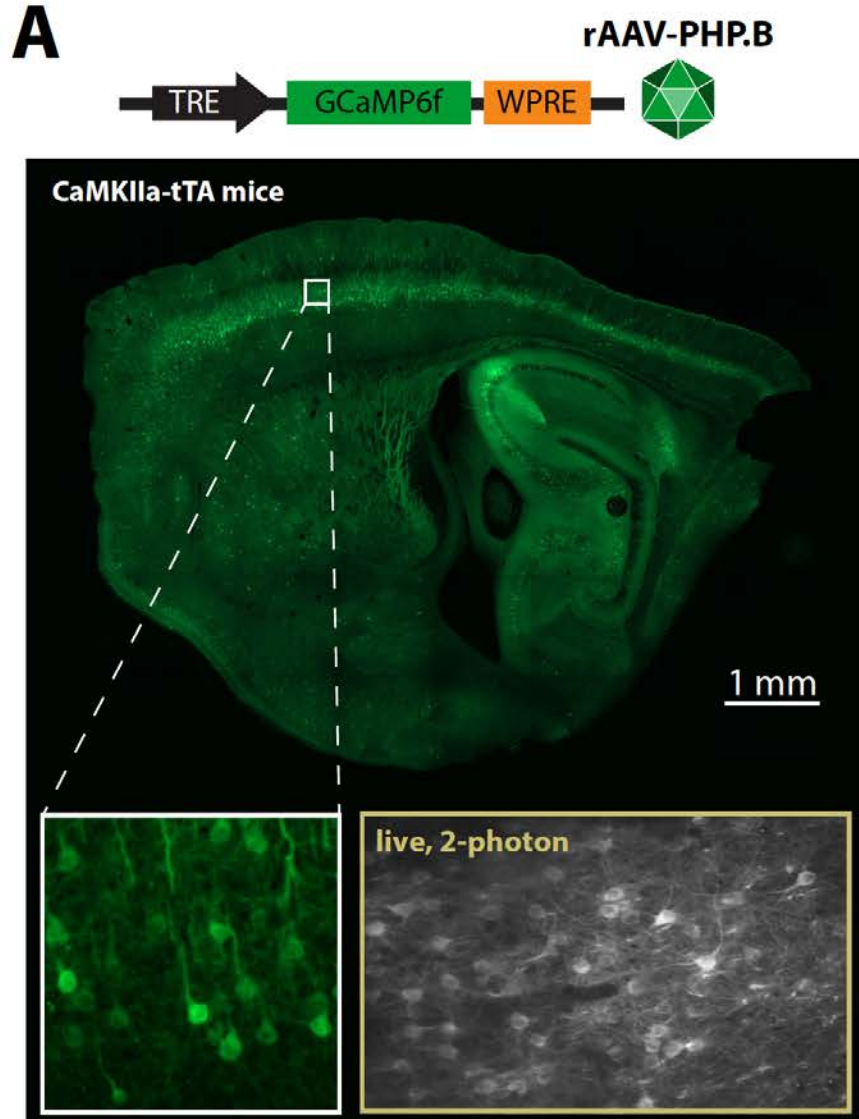
AAV-PHP.eB delivering a vector with a Purkinje cell type-specific promoter driving GFP



AAV-PHP.eB delivering a vector with an inhibitory enhancer specific to the forebrain driving mRuby2



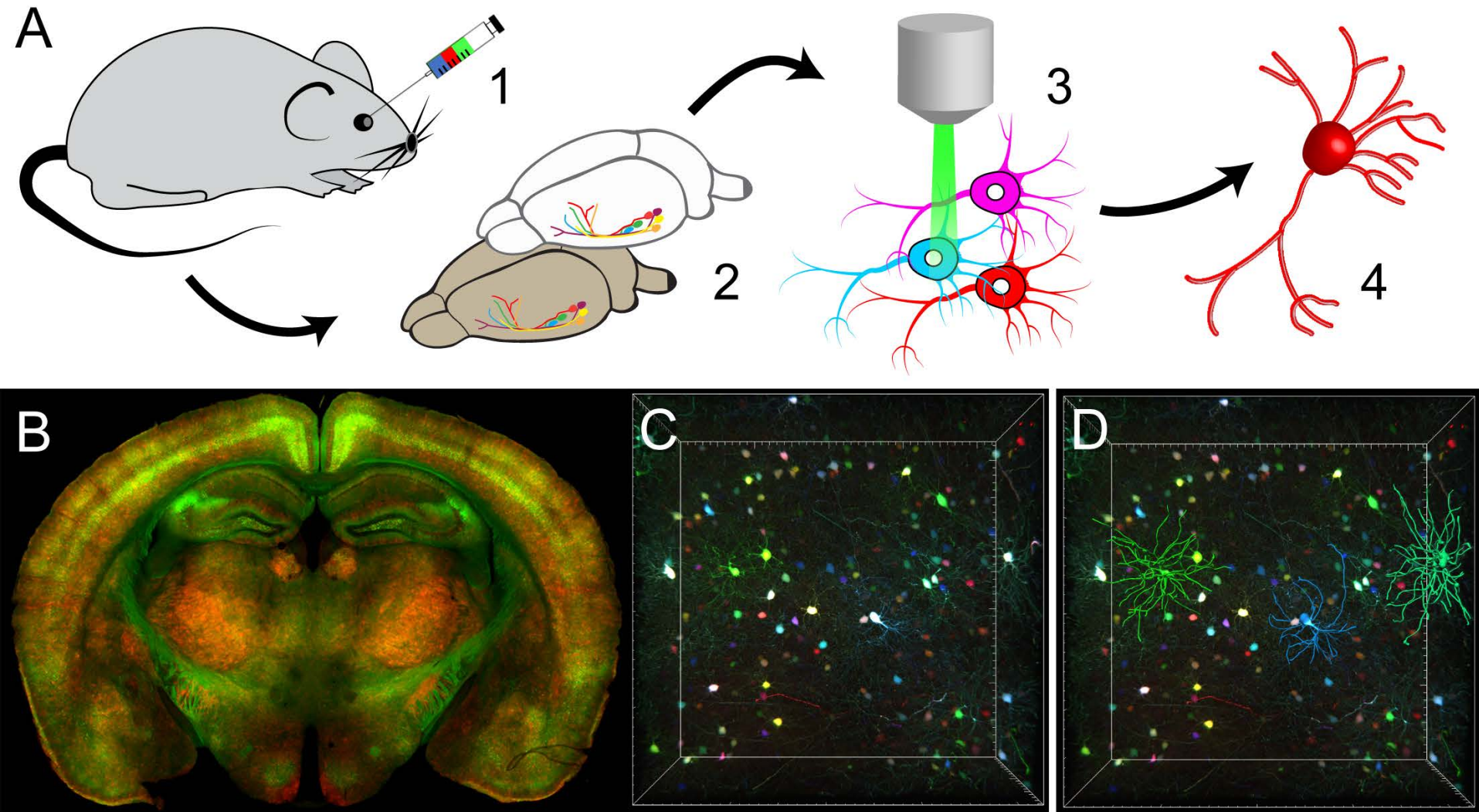
Widespread AAV-mediated delivery for neuronal activity dynamics recording during behavior



A: Gradinaru/Tsao Labs at Caltech

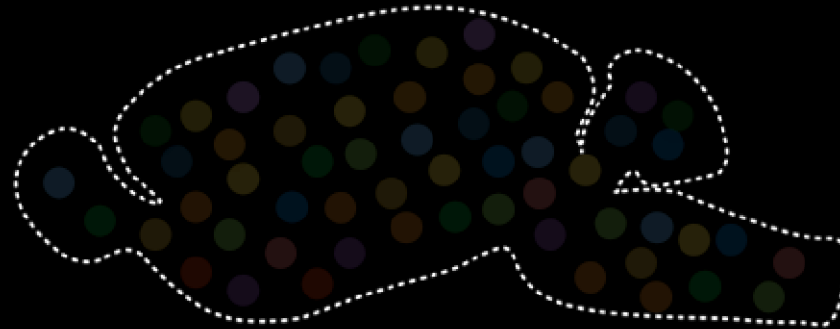
B: Allen et al, Neuron 2017

Broadly transducing AAVs permit brainwide transgene expression and facilitate neurite tracing.

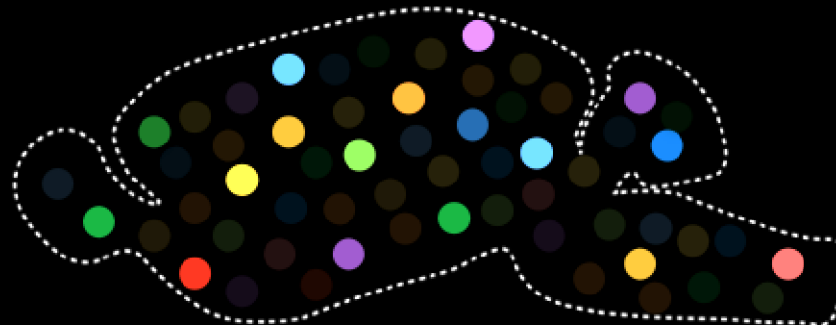
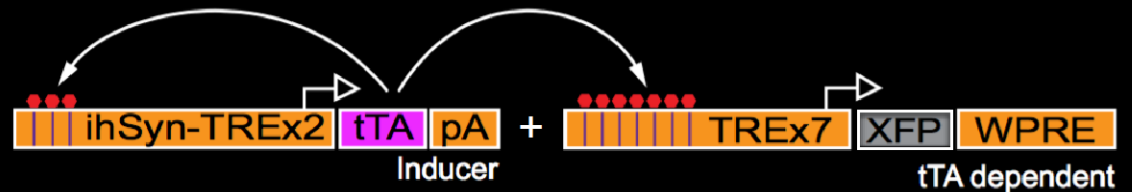


A two-component approach decouples color diversity from labeling density

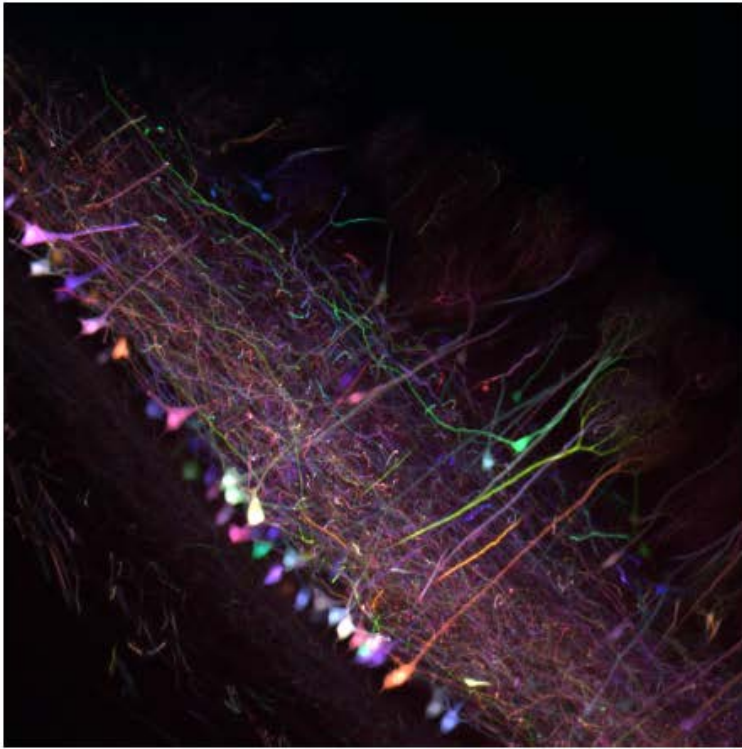
Deliver XFPs at a high dose to with none of them expressing to provide high color diversity



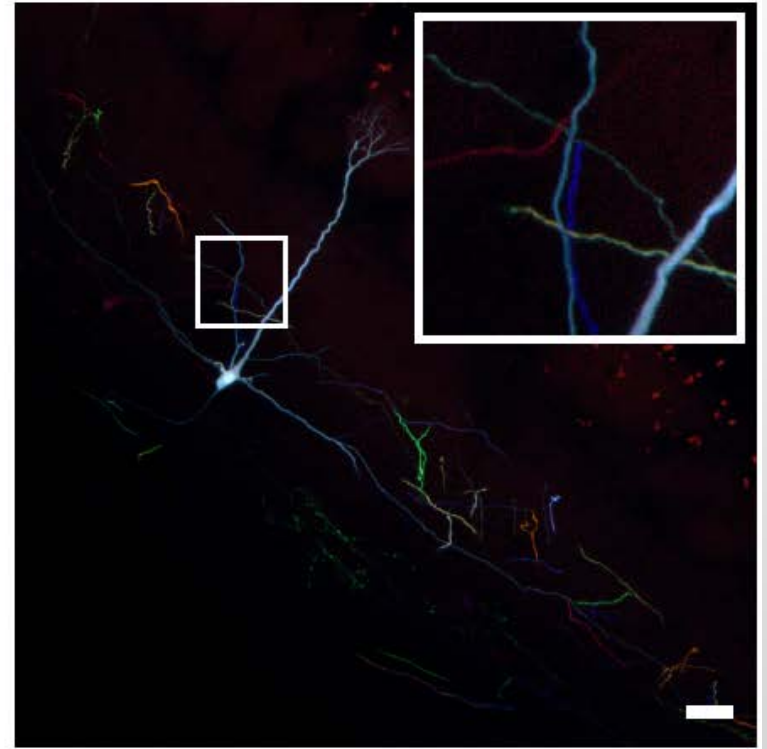
Delivered at a low dose to to sparsely turn on expression



Sparse Stochastic Multicolor Labelling for *Genetic* Tracing



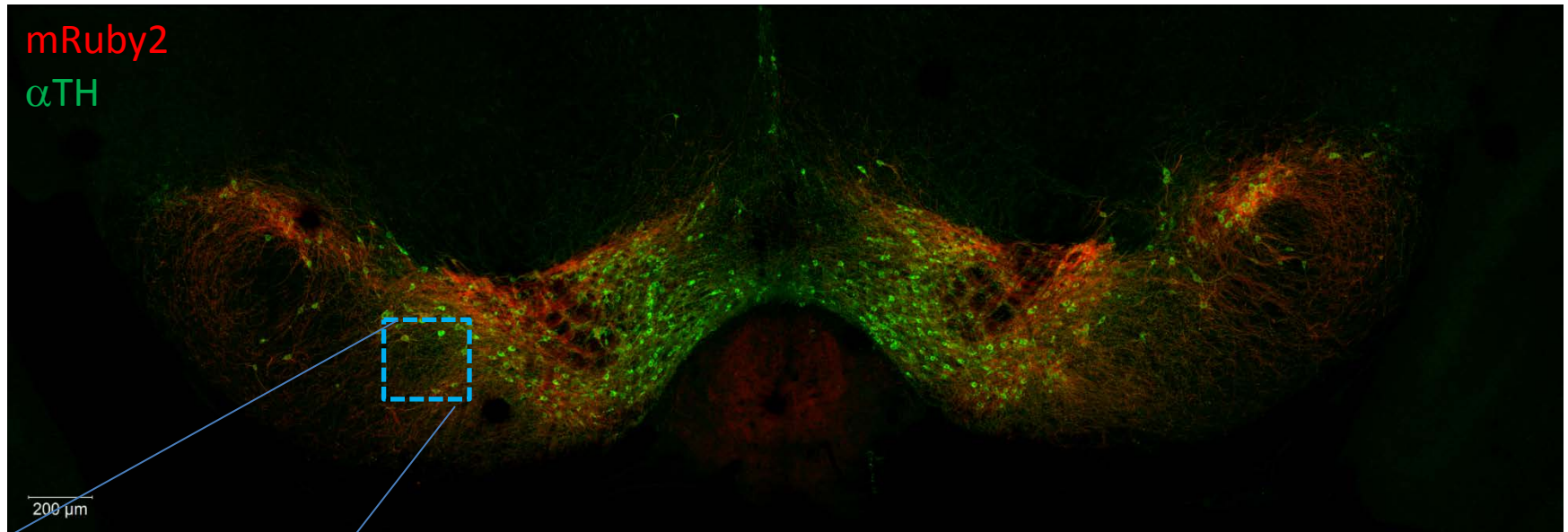
High dose inducer



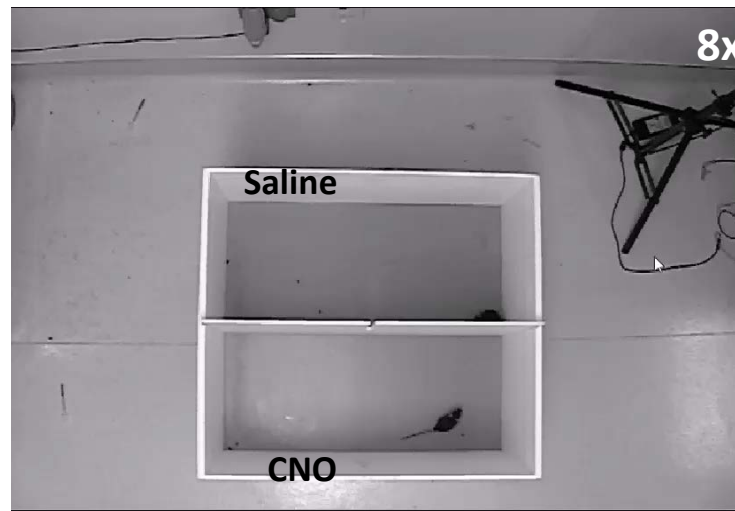
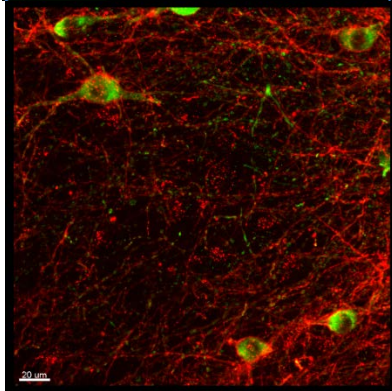
Low dose inducer

Noninvasive behavioral control:

modulation of dopamine neurons with systemic AAVS
and DREADDS changes locomotion



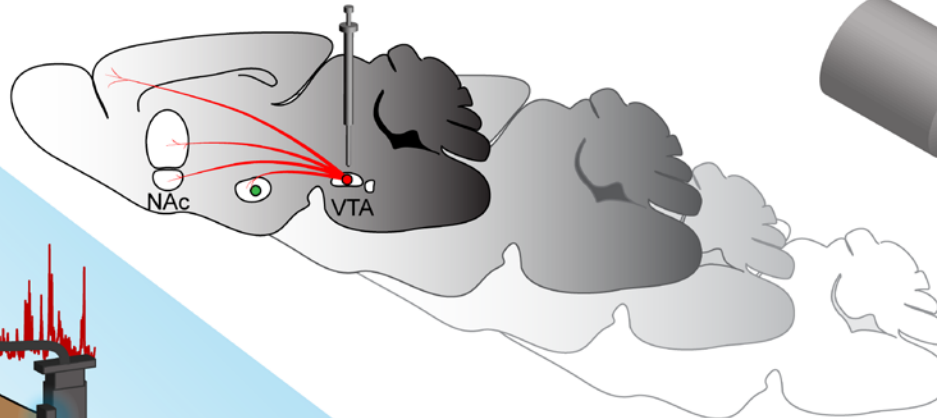
hM3Dq - Open Field



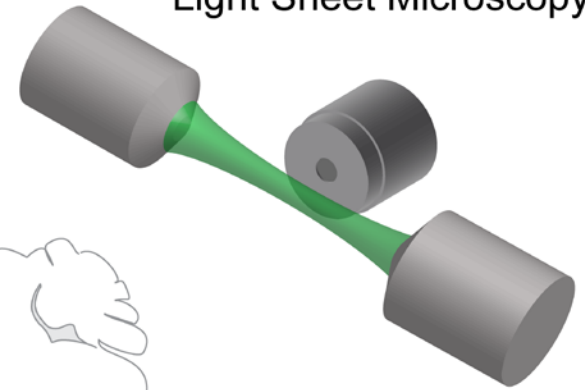
Systemic PHP.eB in
TH-Cre+ animals
delivers
hSyn-DIO-hM3Dq-mRuby2
(5×10^{11} vg, IHC at 3.5 weeks)

Anatomical Analysis and Tracing

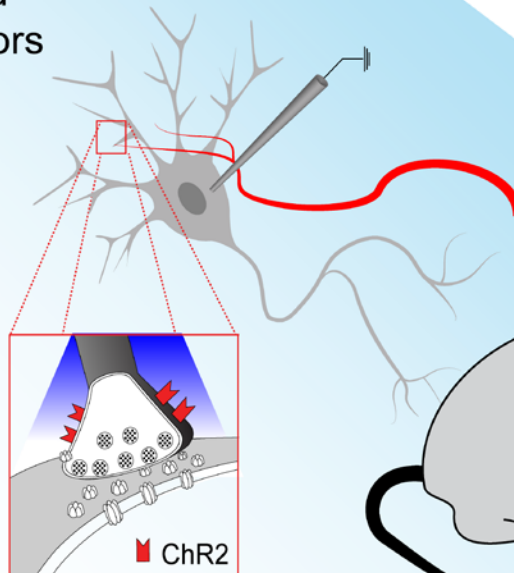
Viral Vector-Based Labelling Methods



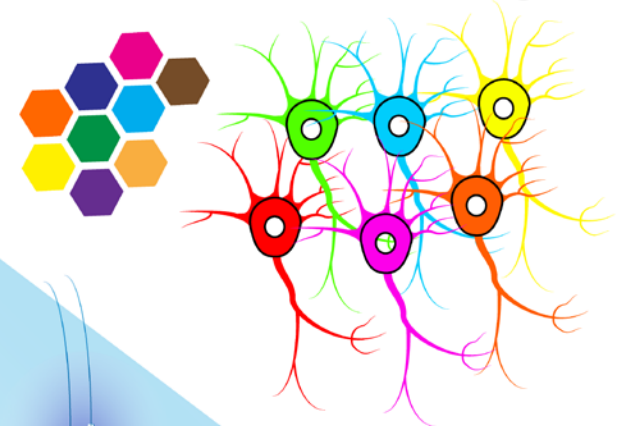
Light Sheet Microscopy



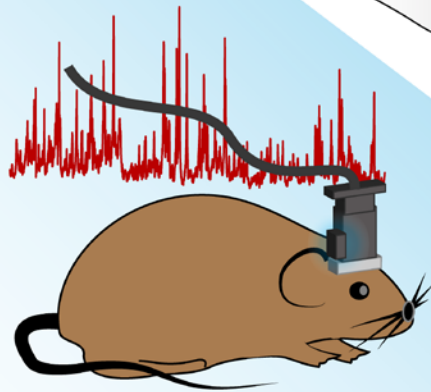
Tissue Clearing



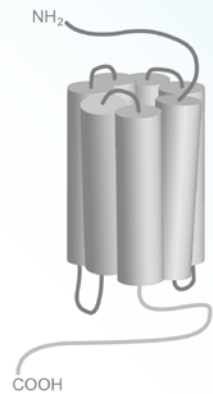
Multicolor Labelling



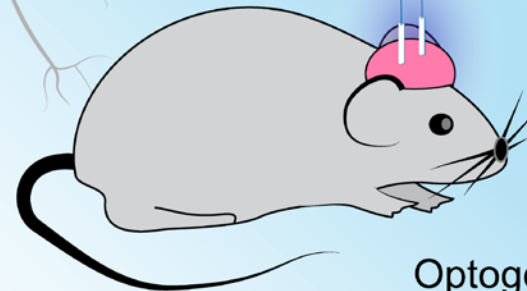
Activity Monitoring and Control



Genetically Encoded Neural Activity Indicators

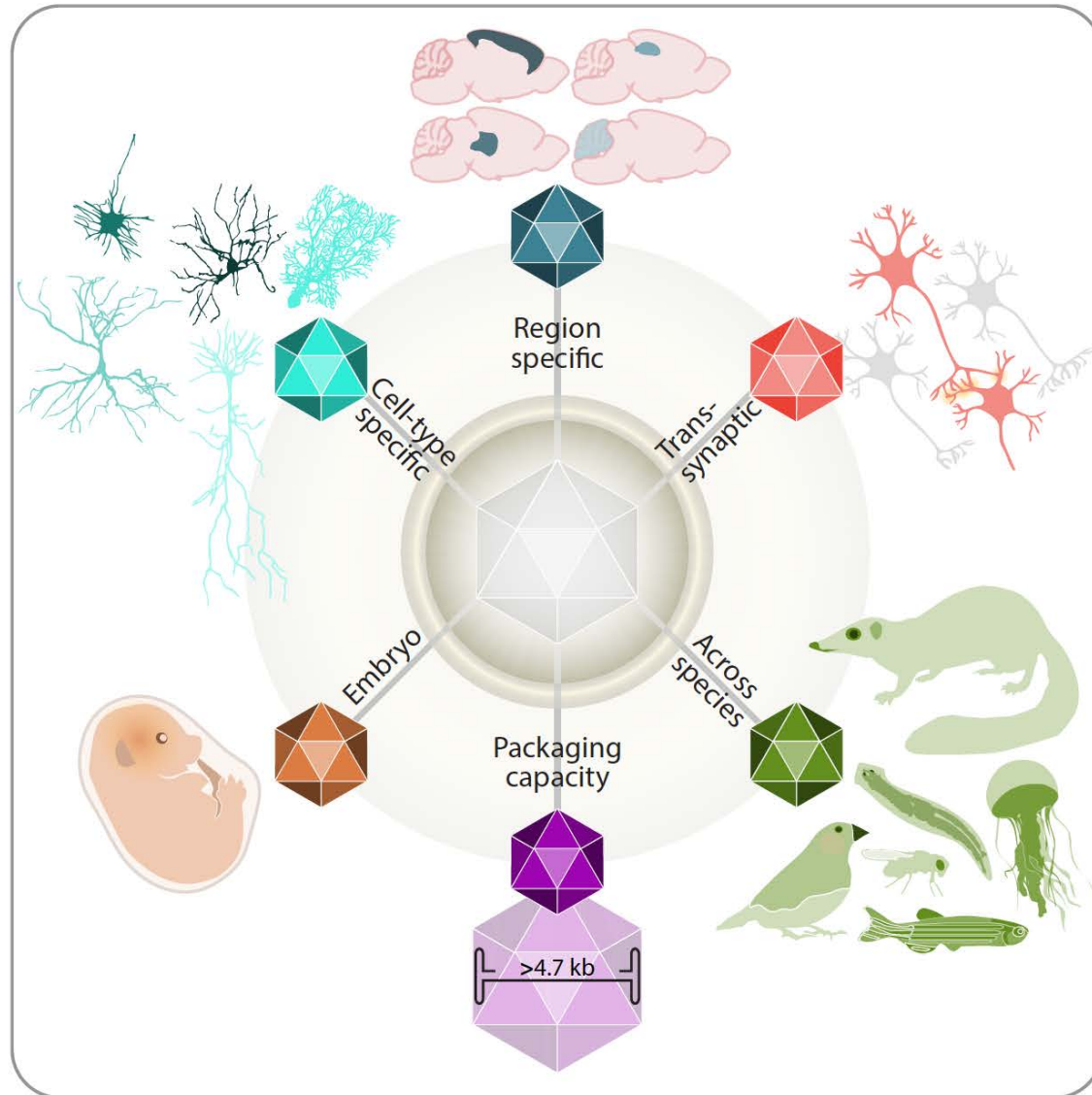


Chemogenetics



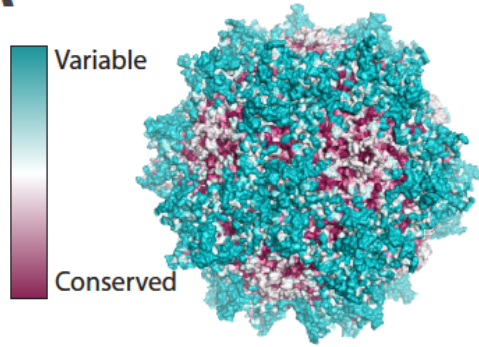
Optogenetics

AAVs – potential developments

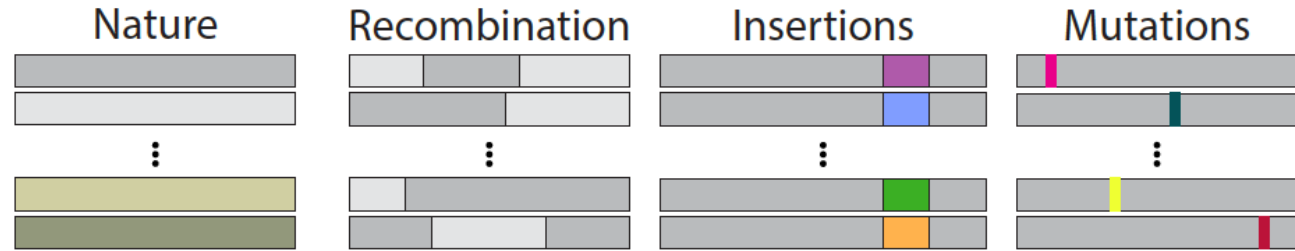


Engineering designer AAVs for neuroscience

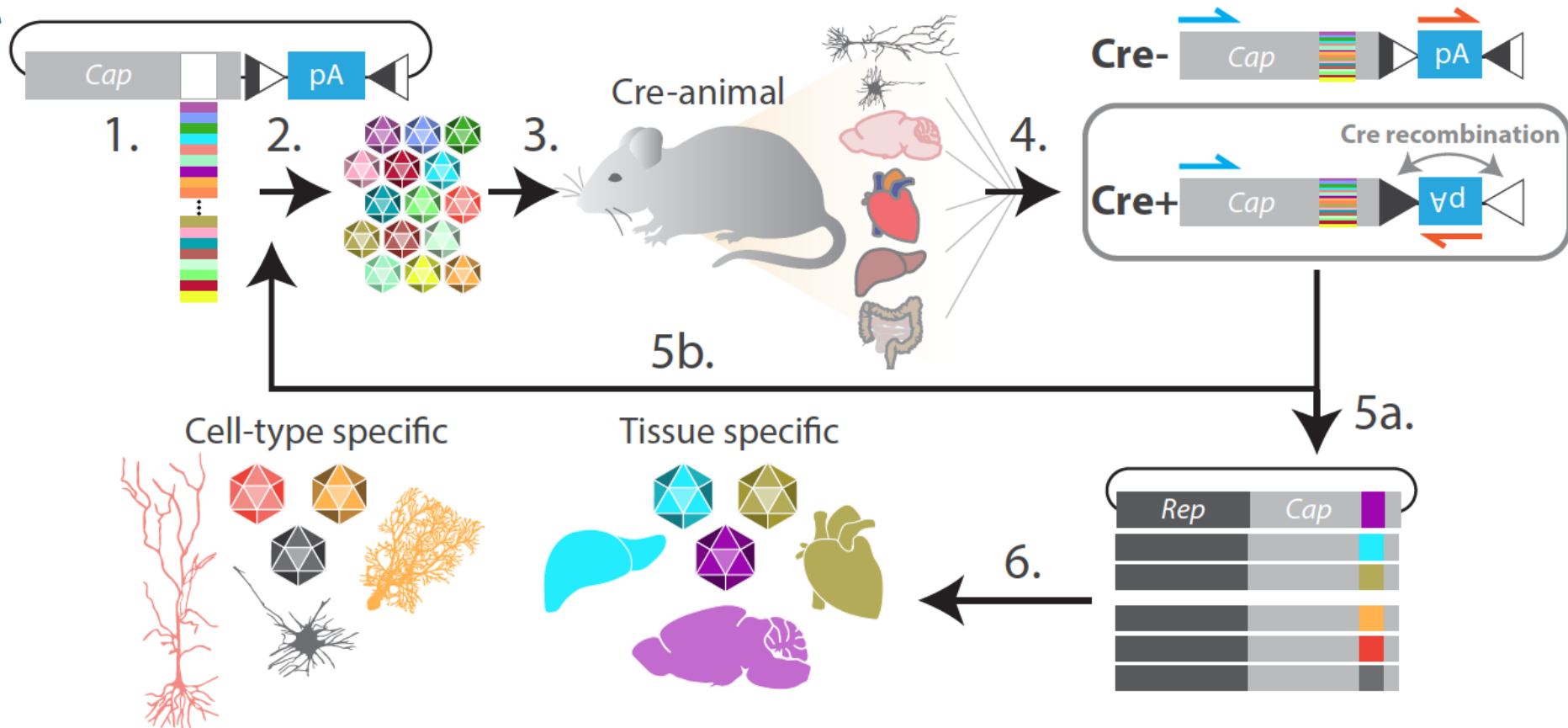
A



B

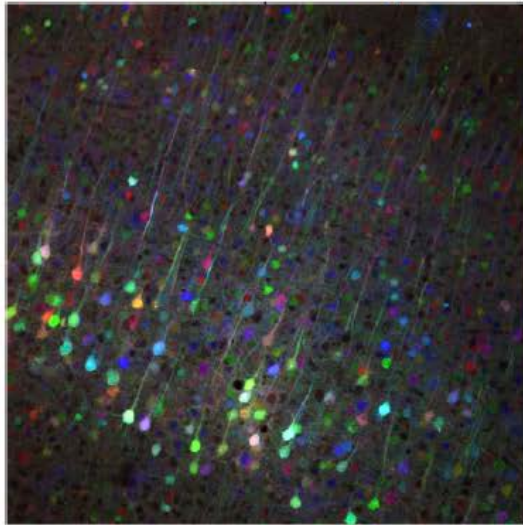


C

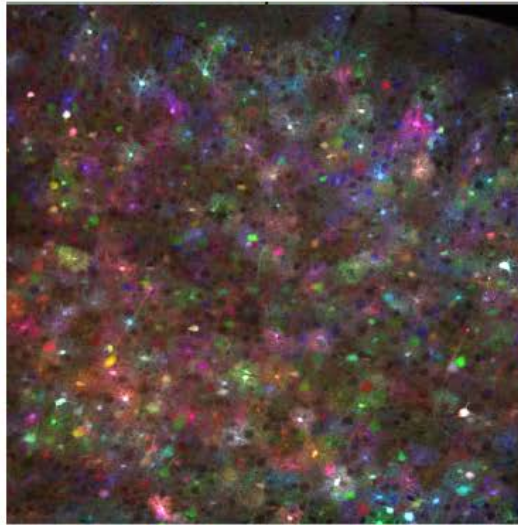


Systemic AAVs

D neuron biased tropism

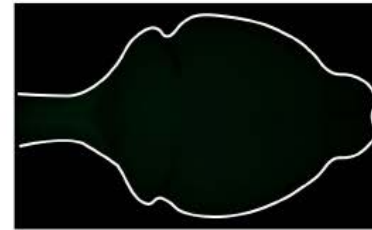


astrocyte biased tropism

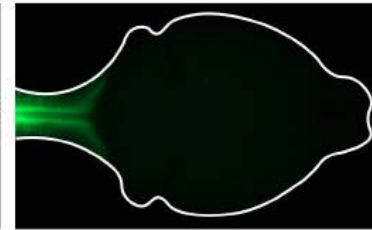


E

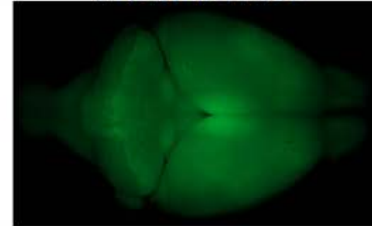
rAAV9



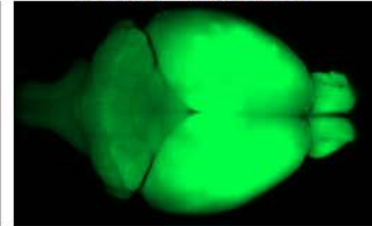
rAAV-PHP.S



rAAV-PHP.B



rAAV-PHP.eB



Visualizing the Activity and Anatomy of Brain Circuits: Optogenetics, Tissue Clearing, Viral Vectors

Gradinaru Lab, Caltech

Claire Bedbrook, Nick Flytzanis

Ken Chan, Priya Kumar, Ryan Cho

Michael Altermatt, Xiaozhe Ding

Jenny Treweek, Alon Greenbaum

Collin Challis, Min Jee Jang

Rosemary Challis, Ben Deverman

Elliott Robinson, Anat Kahan,

Nick Goeden

FORMER MEMBERS:

— Cheng Xiao, Chunyi Zhou

— Lindsay Bremner



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Financial support from:

NIH: R01 NIA, NINDS BRAIN, R24&UC4 NIDDK,

NIH Director's New Innovator, PECASE, SPARC

Foundations: Moore, Beckman, Curci, Sloan, Pew,

Heritage Medical Foundation

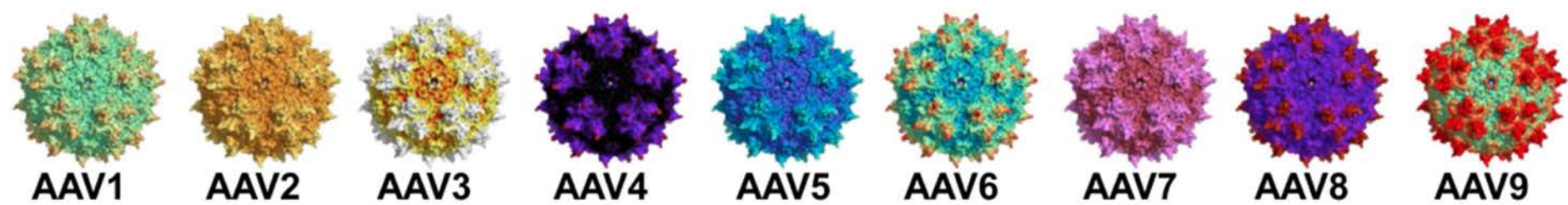
**CLOVER: CLARITY, OPTOGENETICS AND VECTOR ENGINEERING RESEARCH
CENTER @ THE BECKMAN INSTITUTE CALTECH**

PI: Viviana Gradinaru

Director: Ben Deverman (bd@caltech.edu)

<http://www.beckmaninstitute.caltech.edu/clover.shtml>

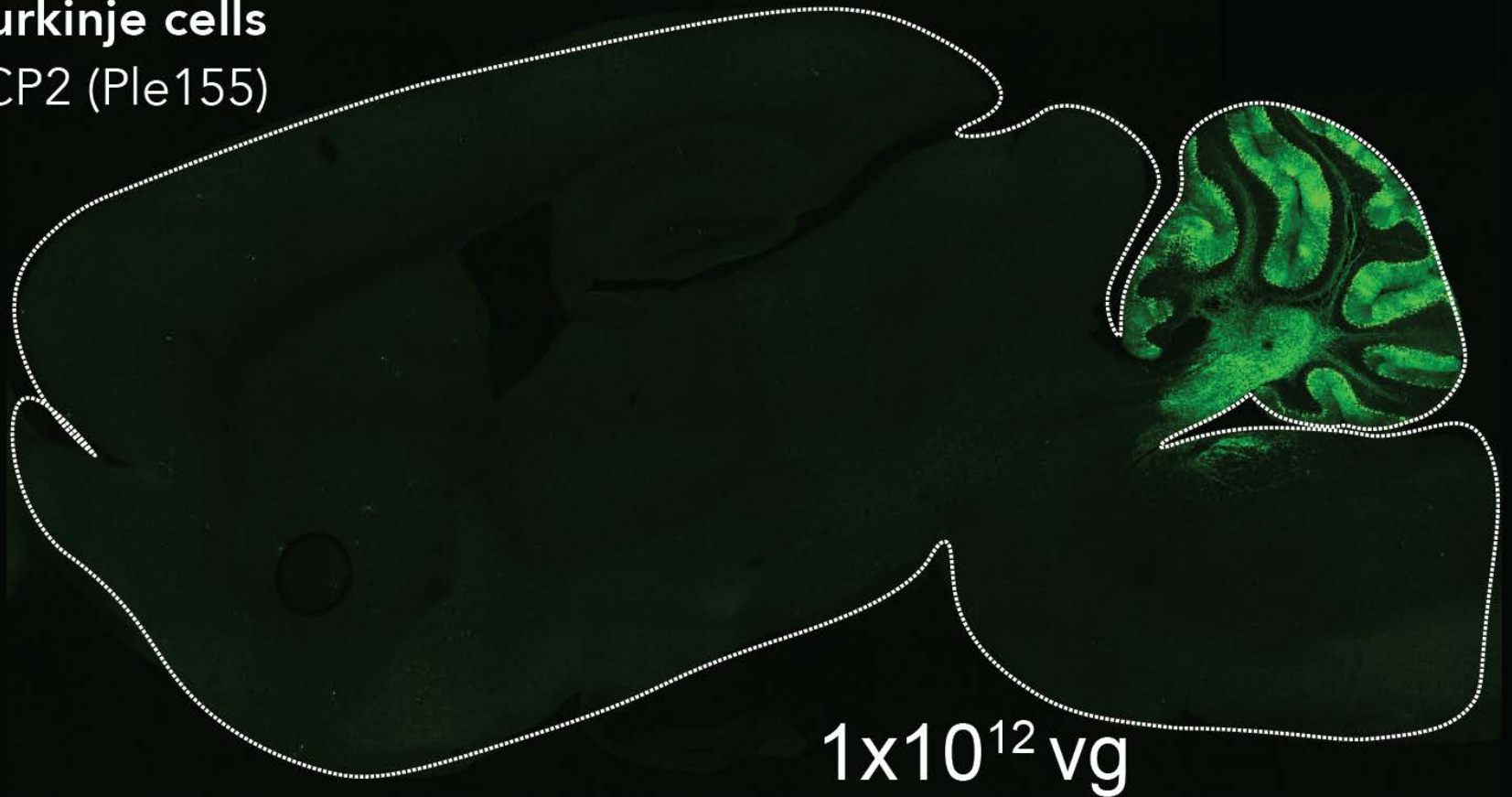
Current applications of AAVs: Pre-clinical (1st row) and Clinical (2nd row)



Heart disease	Various animal model studies	Liver cancer	Mucopolysaccharidoses type VII	Globoid cell leukodystrophy	Duchenne muscular dystrophy		Hemophilia A	Heart failure
Muscular dystrophy			Familial ALS	Human immunodeficiency			Familial hypercholesterolemia	CNS disorders
			RPE65 deficient vision loss				Glycogen storage disease type II	
Lipoprotein lipase deficiency	Cystic fibrosis							
	Hemophilia B							
	Leber's congenital amaurosis		RPE65 deficient disease	Hemophilia A			Hemophilia B	Spinal muscular atrophy
Heart disease	Parkinson's disease							
	Canavan disease							

Purkinje cell restricted expression with IV AAV-PHP.eB and the PCP2 promoter

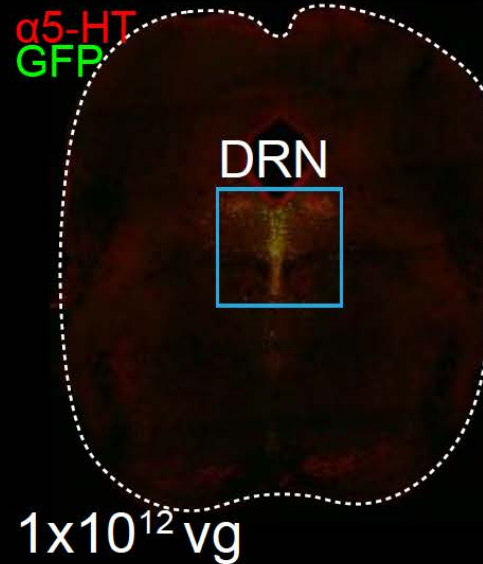
Purkinje cells
PCP2 (Ple155)



Achieving cell type-restricted expression with systemically delivered AAVs

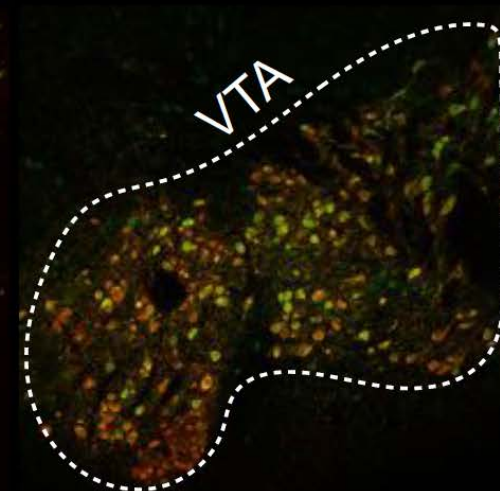
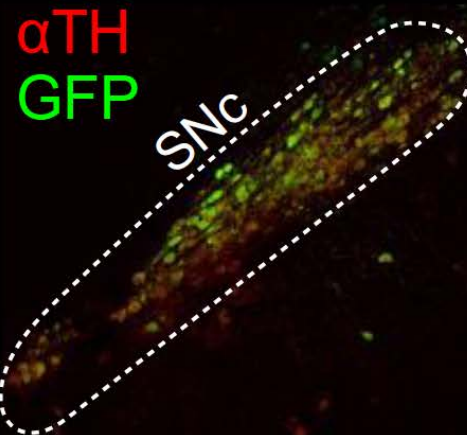
serotonergic
neurons

FEV
(Ple67)



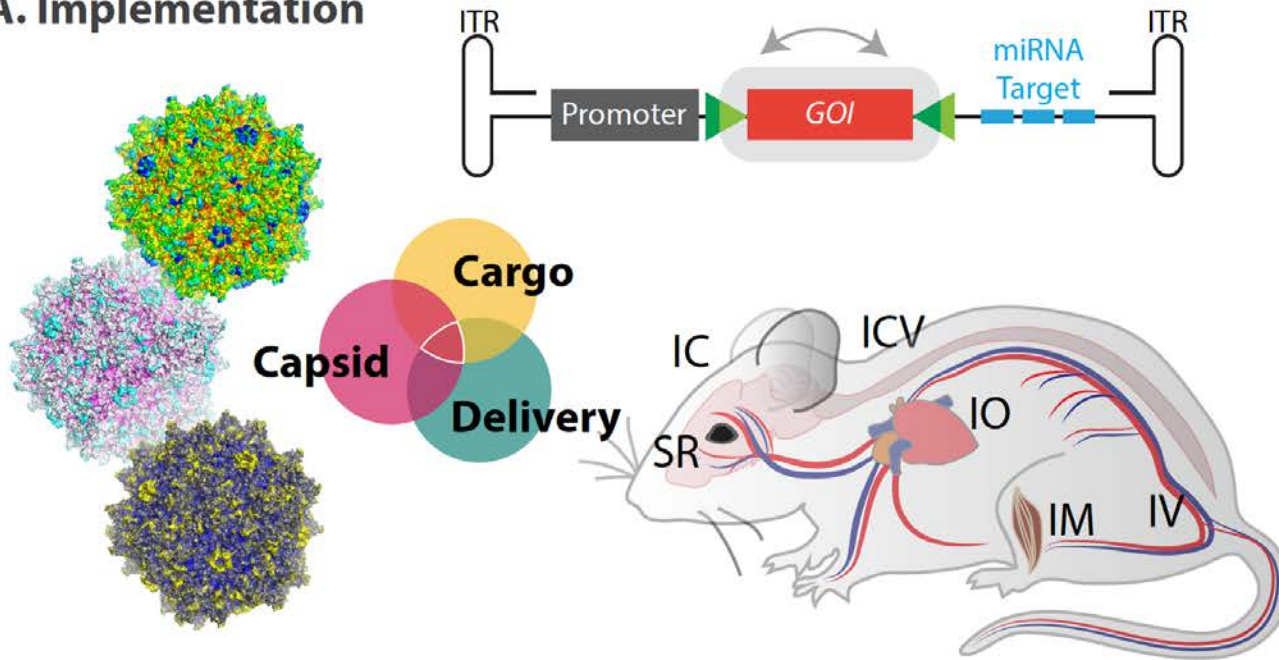
dopaminergic
neurons

mTH



Methods for cell type-restricted expression in the CNS

A. Implementation



B. Delivery: Direct intracranial

Cargo: DIO-FP / Vip-Cre transgenic
Capsid: rAAV5

