

Non-Viral Delivery Nanoplatfoms for Brain-Targeted Genome Editing

Shaoqin Sarah Gong
Vilas Distinguished Professor

Department of Biomedical Engineering
Wisconsin Institute for Discovery
Department of Chemistry
Department of Materials Science & Engineering
University of Wisconsin-Madison

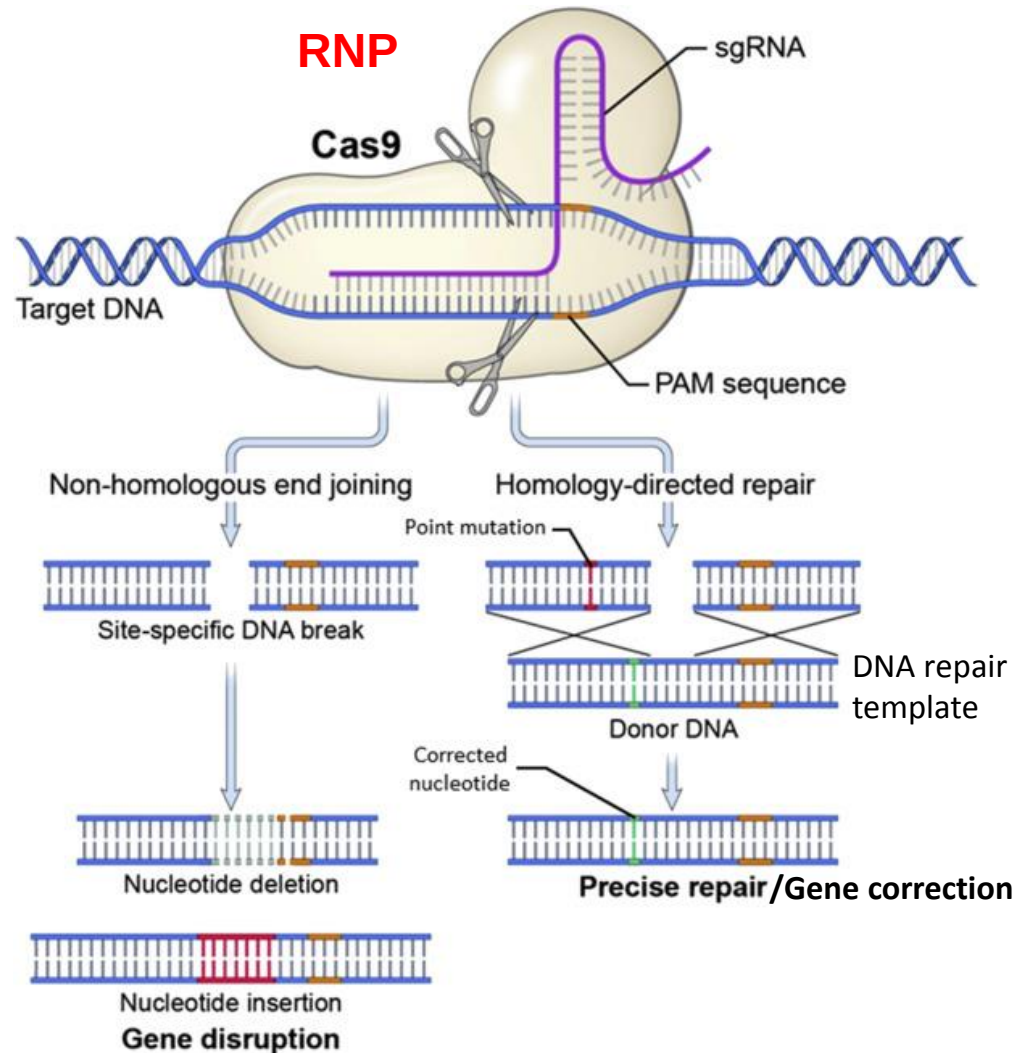
shaoqingong@wisc.edu

CRISPR-Cas9 Genome Editing

Gene deletions, insertions, or alterations

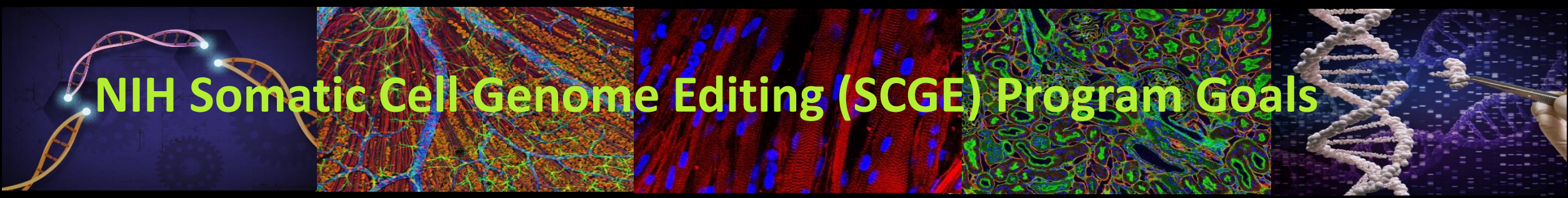
Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR)

CRISPR Associated Proteins 9 (Cas9)



- Cas9/sgRNA **RiboNucleoProtein (RNP)**
- The RNP homes in on the target DNA site encoded in the gRNA.
- Once RNP is activated, it causes a double-strand breakage (DSB) at the target site.
- The cell attempts to repair the DSB, causing either imprecise or precise editing.

Challenge: Safe and efficient delivery of the CRISPR genome editing machinery.



NIH Somatic Cell Genome Editing (SCGE) Program Goals

Lower the Barriers for New Genome Editing Therapies by:

- Testing genome editing reagents and delivery systems in better animal models
- Assessing unintended biological effects
- Improving *in vivo* delivery of genome editing machinery
- Expanding the human genome engineering toolkit
- Coordinating partnerships and disseminating information

Our Collaborative Team



Dr. Shaoqin Sarah Gong
Vilas Distinguished Professor
Dept. of Biomedical Engineering,
UW-Madison
Focus: Nanomedicine; targeted delivery



Dr. Marina E. Emborg
Professor, Dept. of Medical Physics,
UW-Madison
Focus: Neurodegenerative disorders;
translational neuroscience



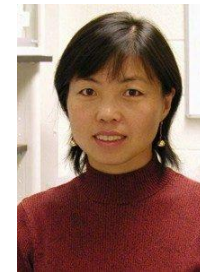
Dr. Jon Levine
Professor, Wisconsin National Primate
Research Center, UW-Madison
Focus: Gene silencing in neural
populations; mouse and monkey models



Dr. Subhojit Roy
Professor, Dept. of Neuroscience,
UW-Madison
Focus: Neuron cell and brain models;
CRISPR tools editing APP



Dr. Krishanu Saha
Assistant Professor, Dept. of
Biomedical Engineering,
UW-Madison
Focus: Novel CRISPR technologies

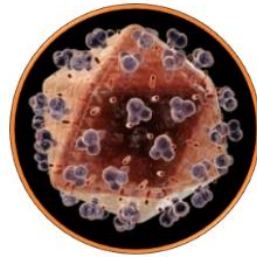


Dr. Xinyu Zhao
Professor, Dept. of Neuroscience,
UW-Madison
Focus: Neural stem cell and brain
development

Non-Viral vs. Viral Vectors

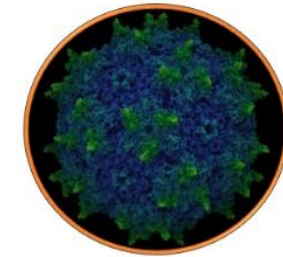
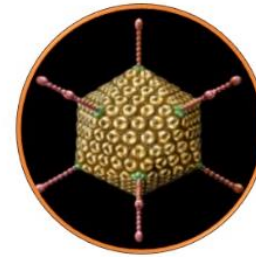
Viral Vectors

- ✓ High efficiency
- ✗ Harder to scale up
- ✗ Immunogenicity
- ✗ Mutagenesis



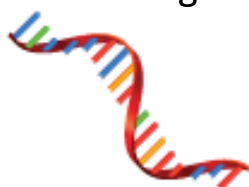
Non-Viral Vectors

- ✓ Better safety profile
- ✓ Versatile chemistry
- ✓ Easier to scale up
- ✗ Low efficiency



Features	Lentivirus	Adenovirus	AAV
Packaging Capacity	8-9 kb	8-9 kb	3-4 kb
Integrating	Yes	No	Rare
Immune Response	**	****	**

Delivery of Genome Editing Machinery

Cas9 Delivery Methods			
	pDNA 	mRNA +sgRNA 	Protein +sgRNA (RNP) 
High Efficiency	++++	++++	++++
Low Cost	++++	++++	++++
Specificity	++++	++++	++++

Mirus Bio

Protein

- Precise control over the Cas9/sgRNA ratio
- Rapid editing (no lag time)
- Lower off-target effect

pDNA or mRNA

- Variable copy number
- Delayed editing

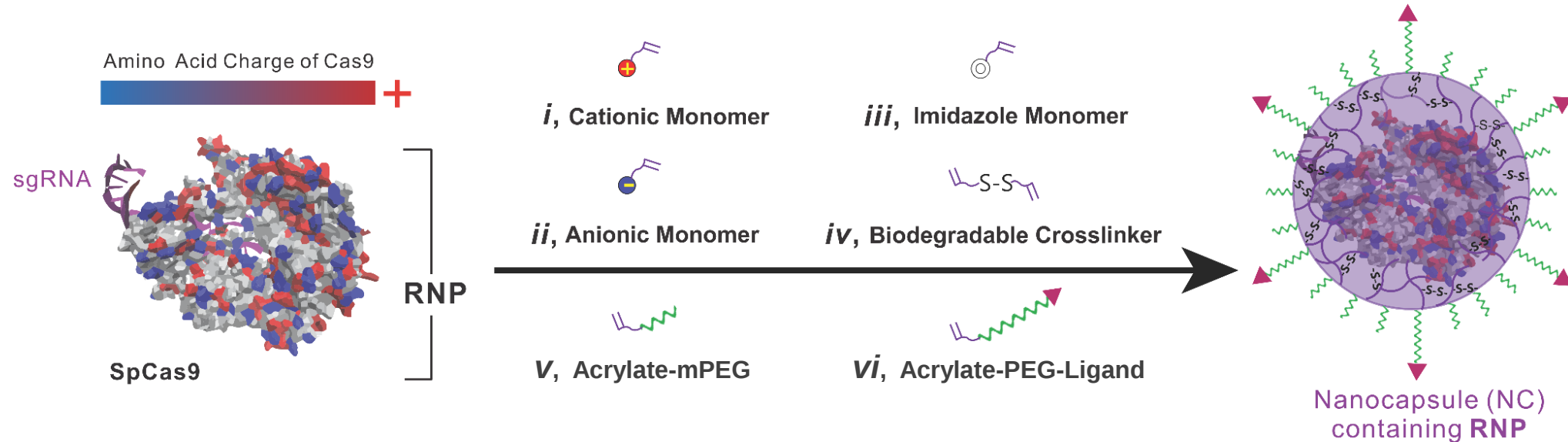
pDNA

- Higher off-target effect

Direct delivery of RNP requires non-viral vectors.

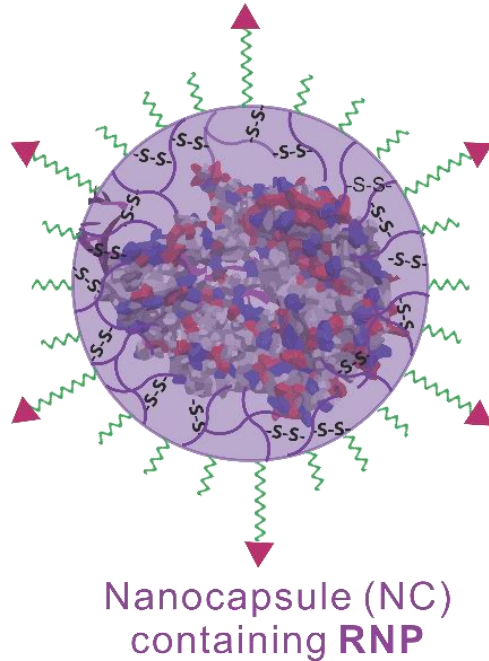
RNP Nanocapsule (NC)

Data presented in this talk have not been published.

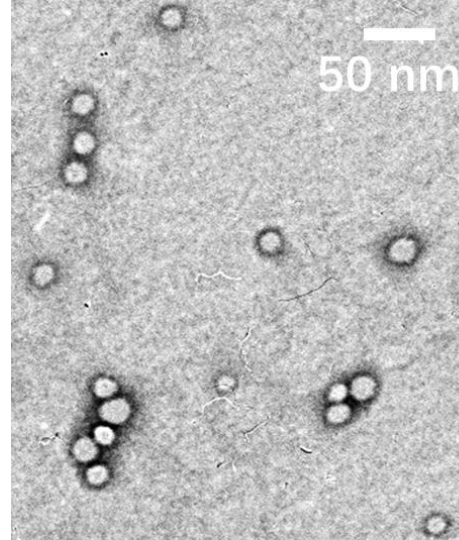


- Formation of the **covalently-crosslinked**, yet **intracellularly biodegradable**, nanocapsule (NC) for the delivery of the Cas9 RNP complex
- The surface of the RNP NCs can be conveniently functionalized with various types of targeting ligands.

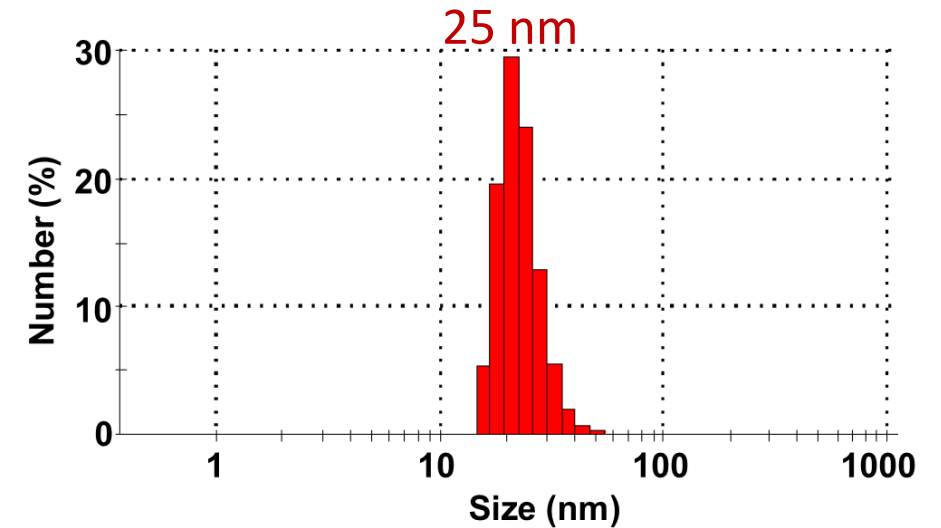
Characterizations of RNP NCs



TEM image



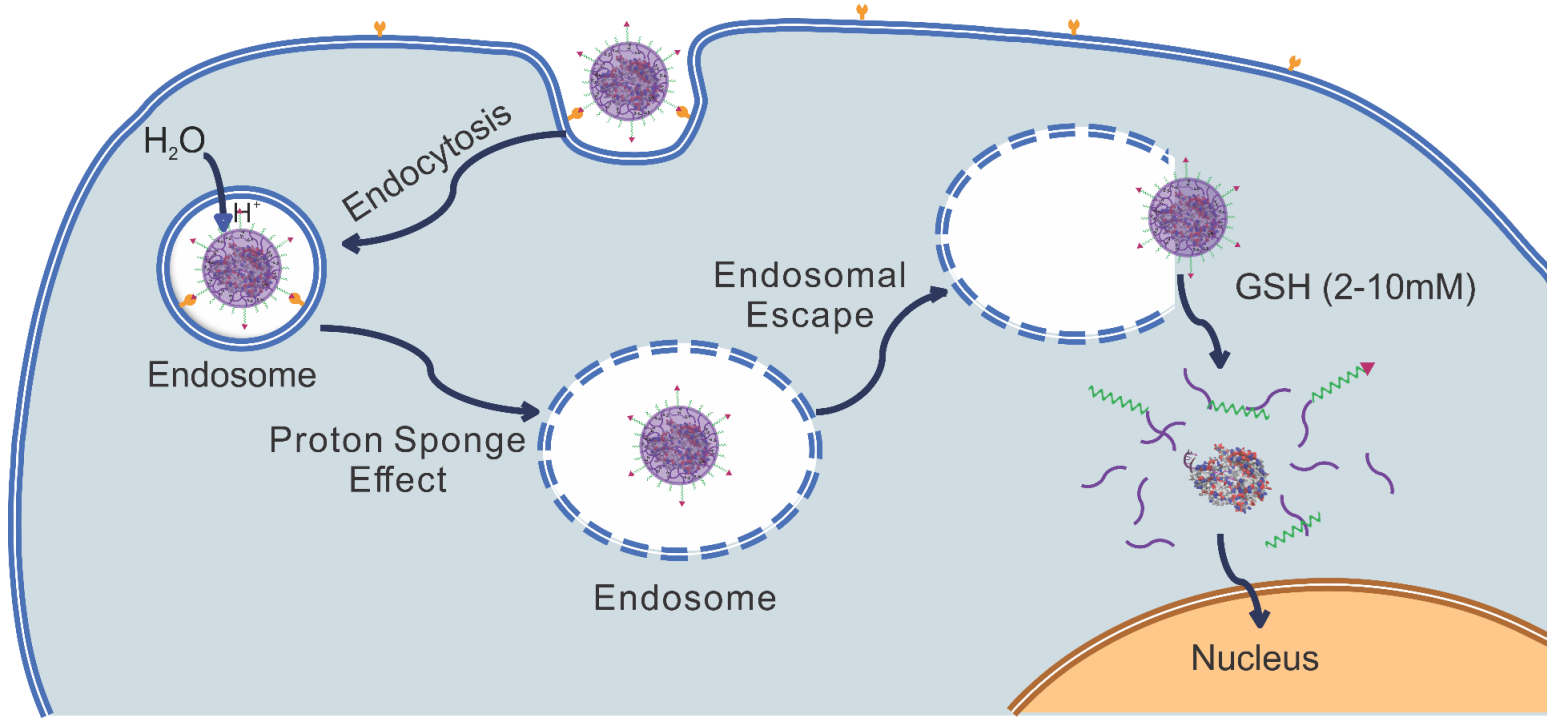
Dynamic light scattering (DLS)



- Hydrodynamic diameter ~ 25 nm
- **Neutral** surface charge (-4 mV)

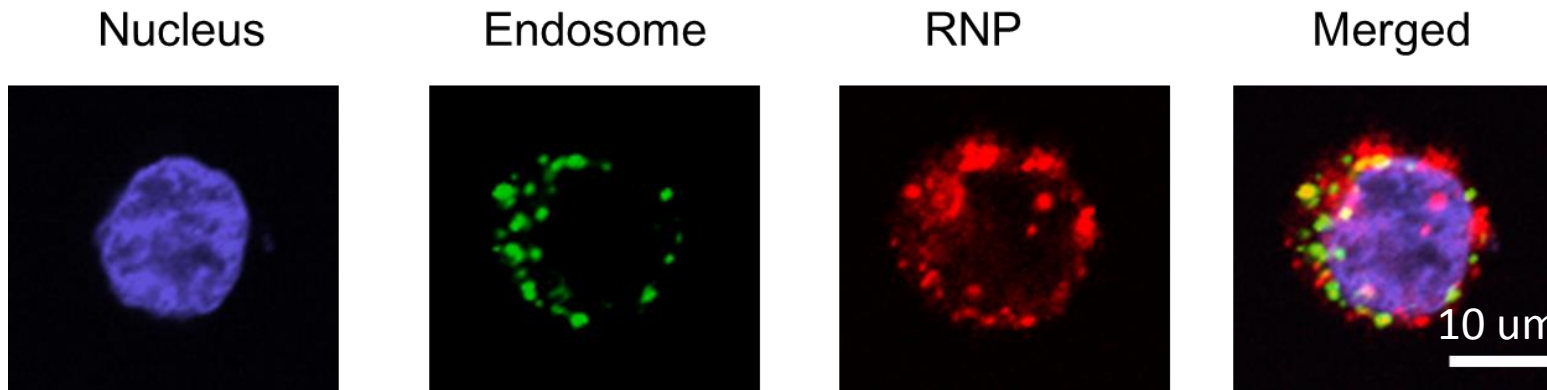
	Cas9 protein	RNP	NC
Size (nm)	7 ± 1	9 ± 1	25 ± 6
Zeta potential (mV)	13 ± 3	-20 ± 5	-4 ± 1

Intracellular Pathway of RNP NCs



Cellular uptake and subcellular trafficking of the RNP NCs:

- High cellular uptake
- Efficient endosomal escape
- Rapid release of the RNP inside of the cytosol
- Efficient nuclear transport

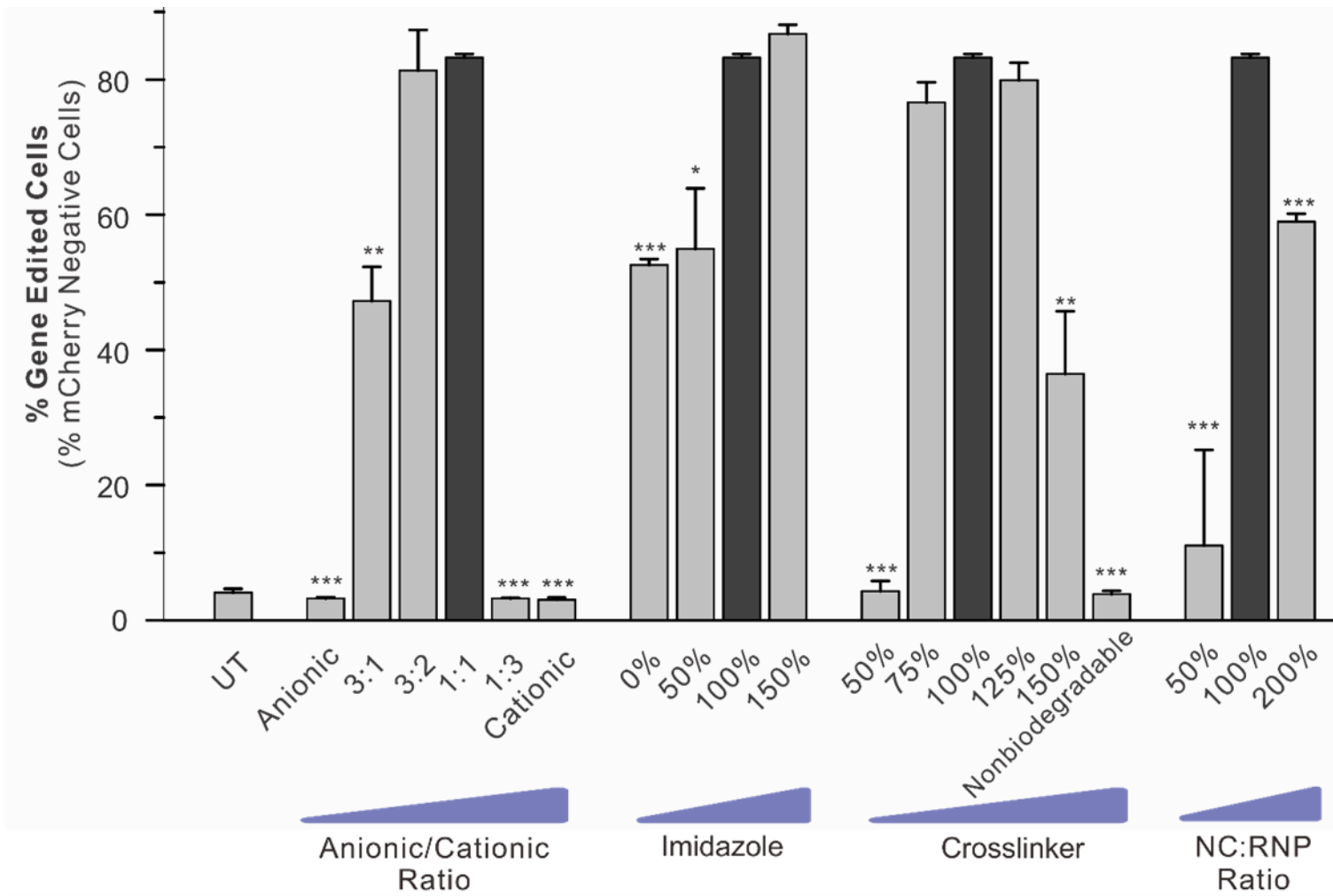


RNP NCs can effectively **escape from endosomes**.

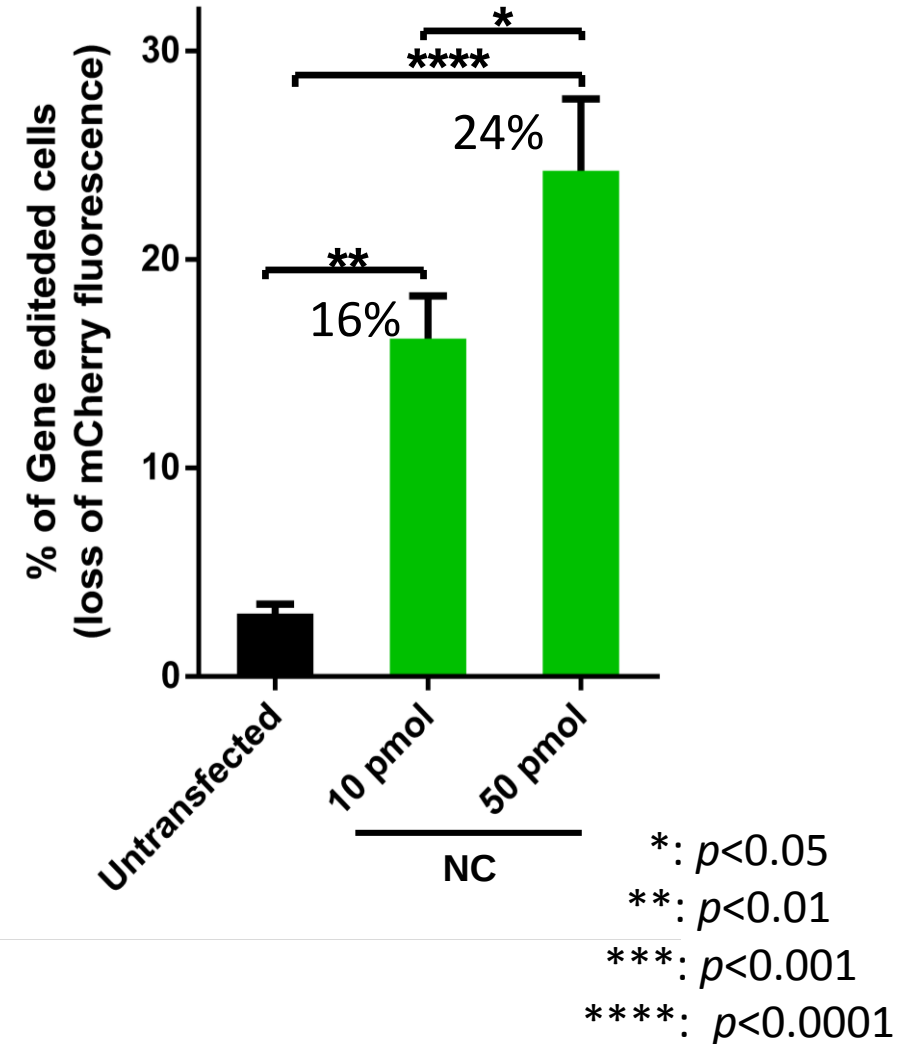
Blue: DAPI Green: Lysotracker Red: ATTO-labeled RNP 6 hr post-incubation

Optimization of RNP NCs

mCherry-HEK 293 cells
gRNA targets the mCherry gene

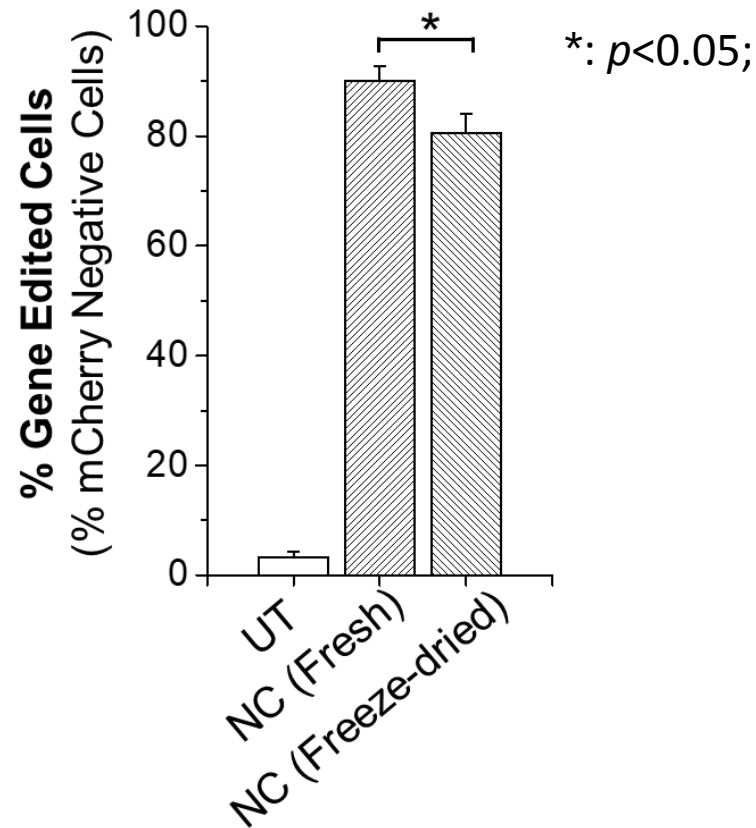


mCherry-human neural progenitor cells (mCherry-NPCs)



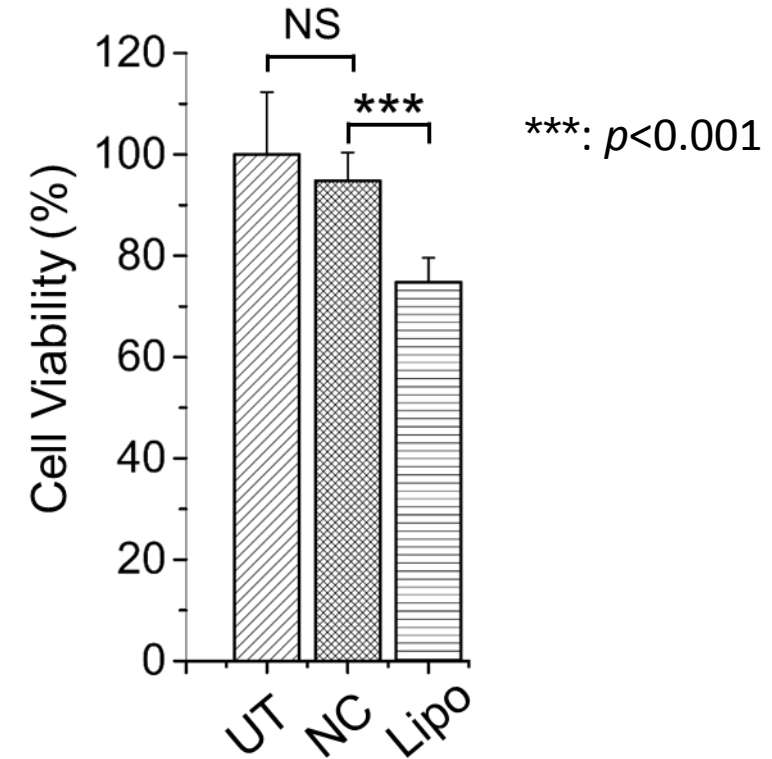
Effect of Lyophilization on Gene Editing and Cytotoxicity of NCs

Gene editing after lyophilization



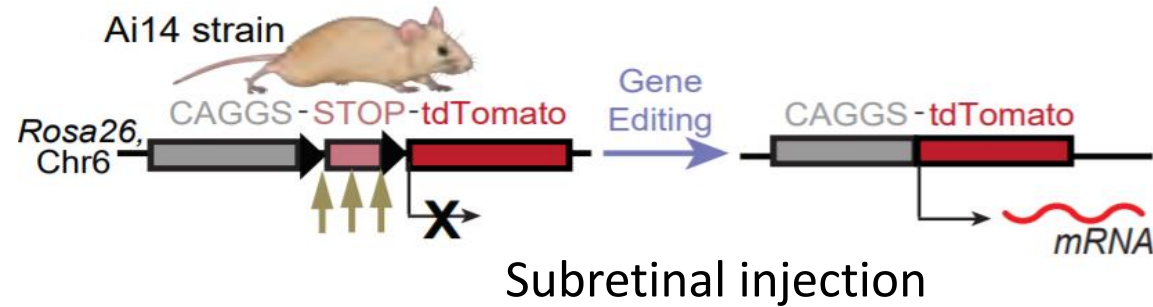
- Over **90%** of gene editing capability was retained after lyophilization of NCs.

Cytotoxicity

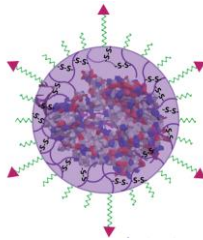
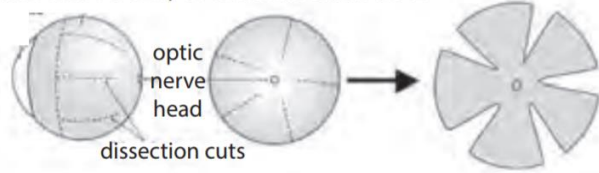


- NCs did **not** induce significant cytotoxicity.
- Lipo2000**, a commercially available agent to deliver RNP, caused significant cell death (**>25%**).

RNP NCs: *In Vivo* Gene Editing With RPE Cells

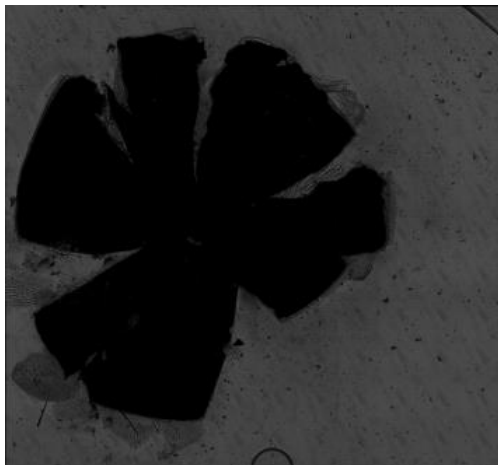


Confocal: Fixed, Whole Mount RPE

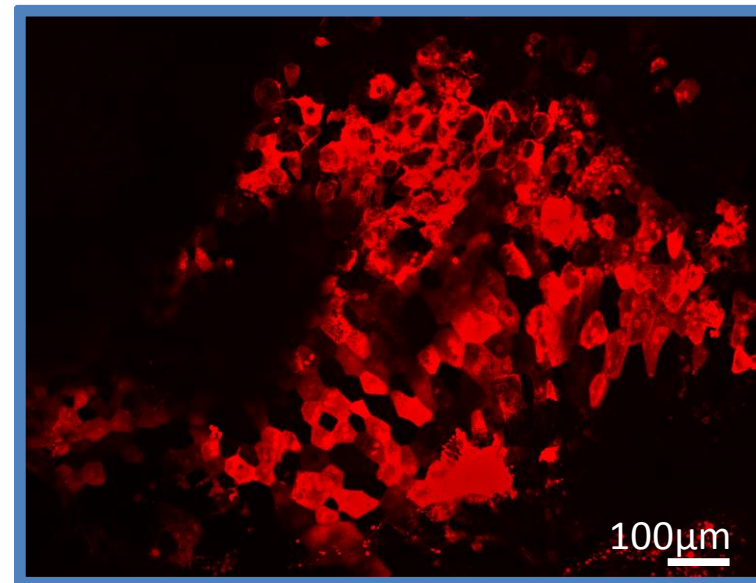
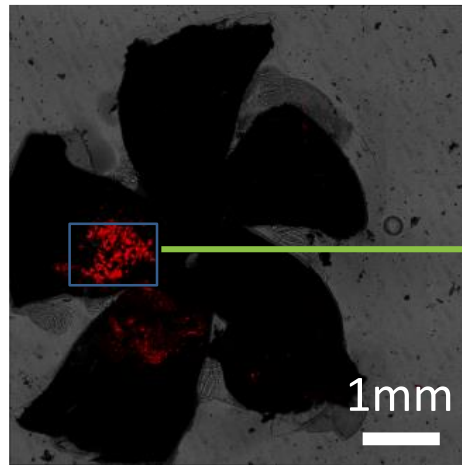


- The Ai14 mice harbor a *LoxP*-flanked stop cassette that prevents expression of the **tdTomato** fluorescent protein.
- RNP-guided excision of the stop cassette leads to tdTomato expression.

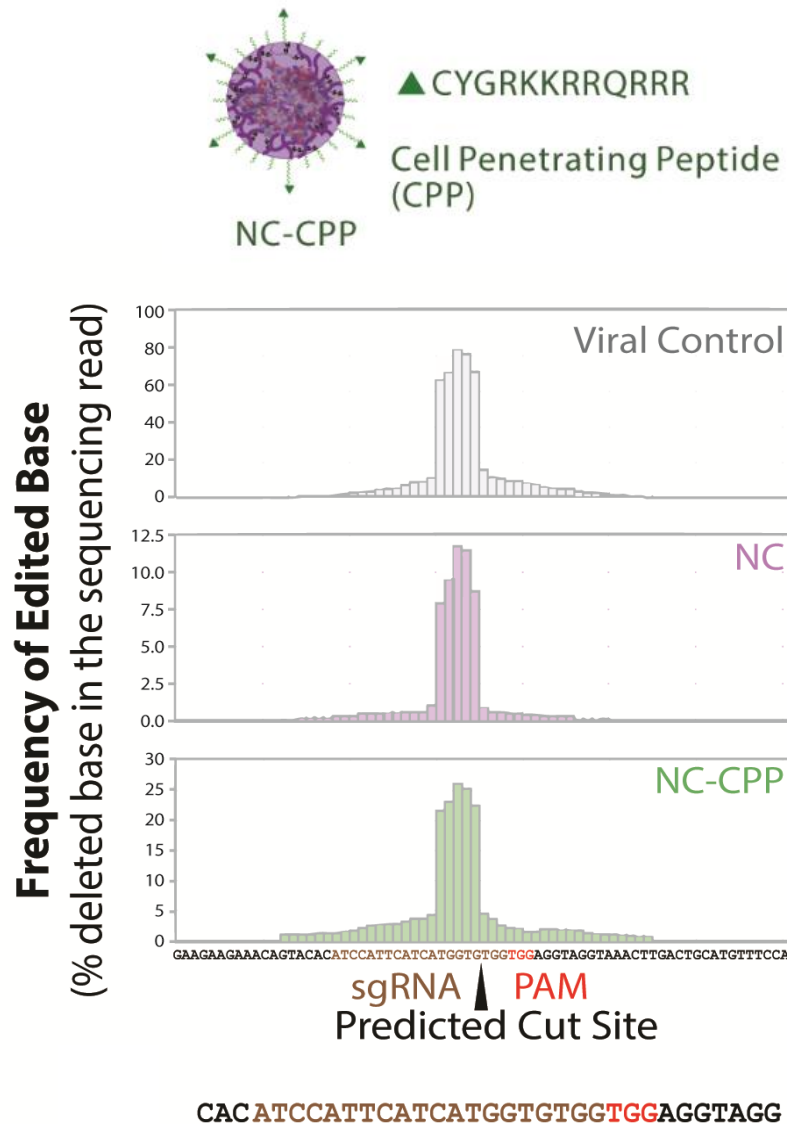
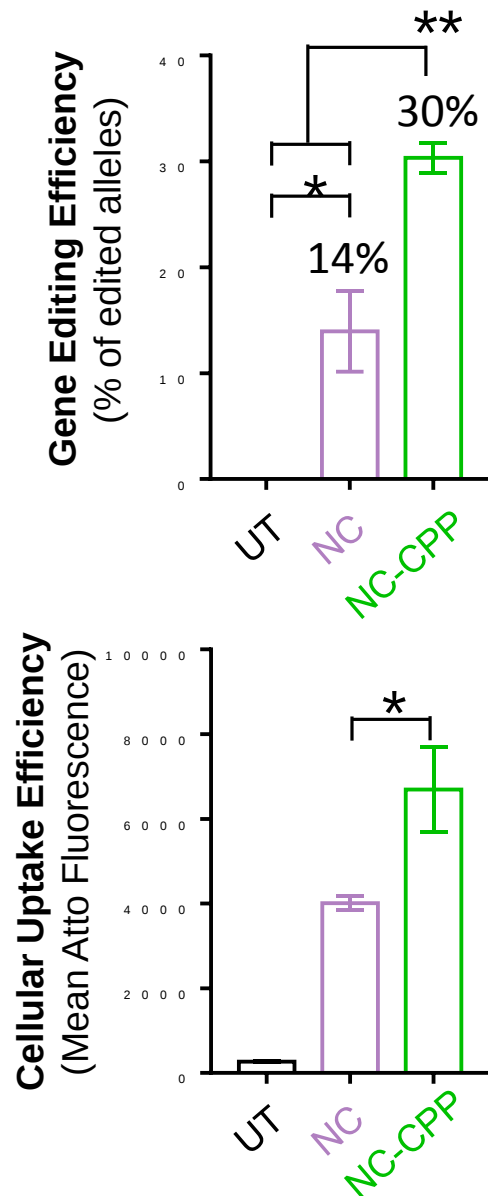
PBS



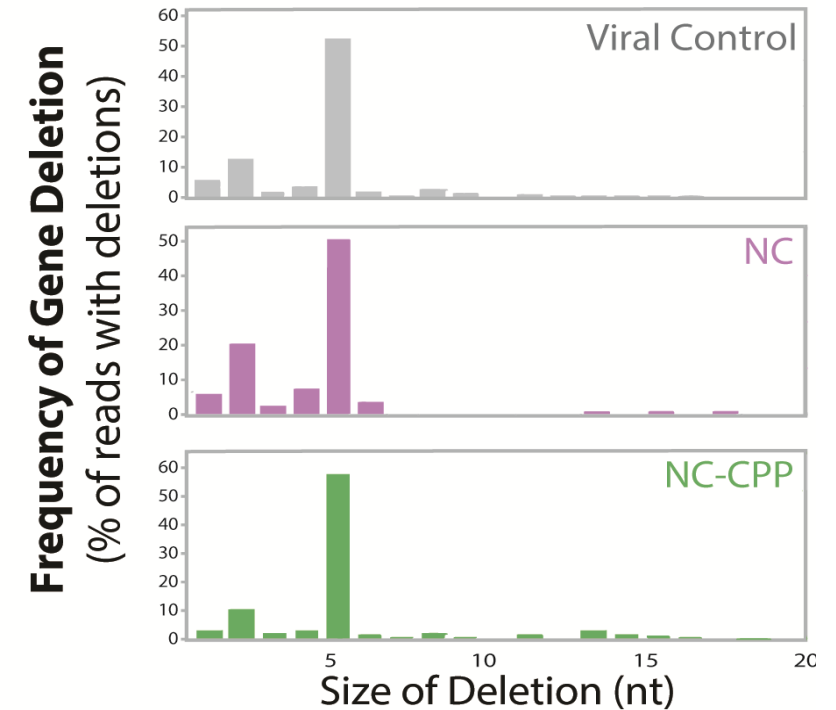
NC-RPE targeting ligand



RNP NCs Targeting Human APP Gene in HEK cells



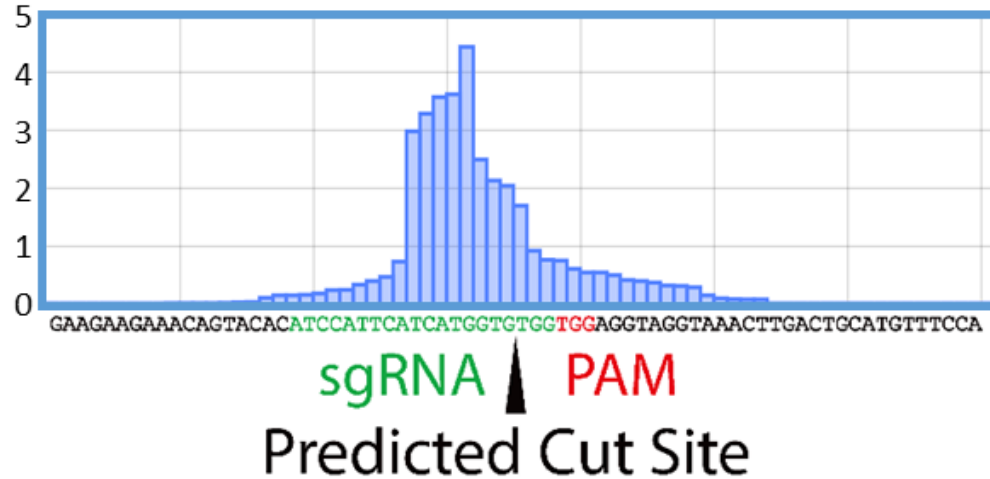
APP: amyloid precursor protein
CPP: cell-penetrating peptide



RNP NC-CPP Targeting Mouse APP Gene in Primary Mouse Neurons

Frequency of edited base
(% of sequencing reads with base deleted)

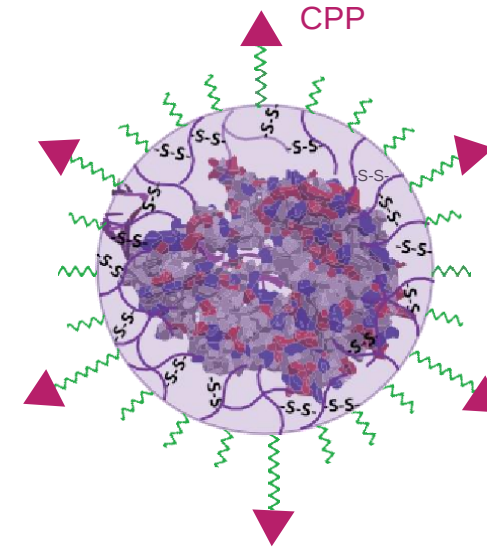
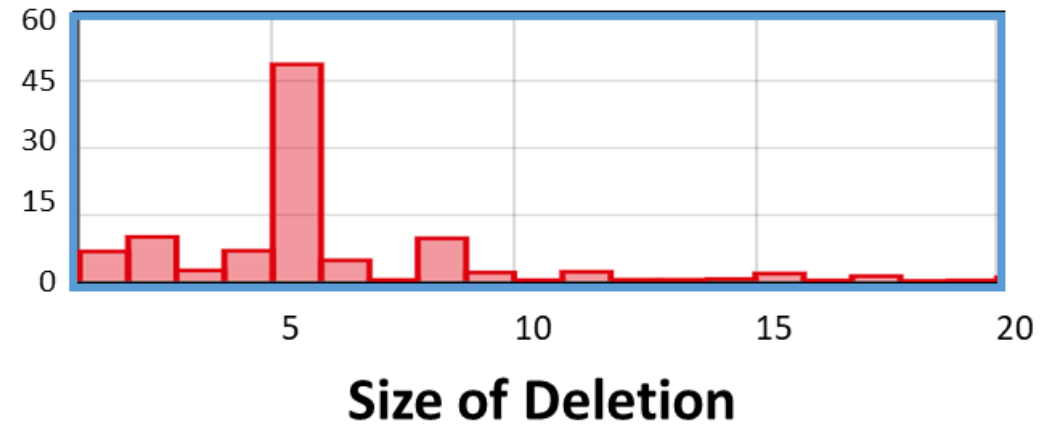
5.9% editing in primary neurons



APP: amyloid precursor protein

Preliminary data

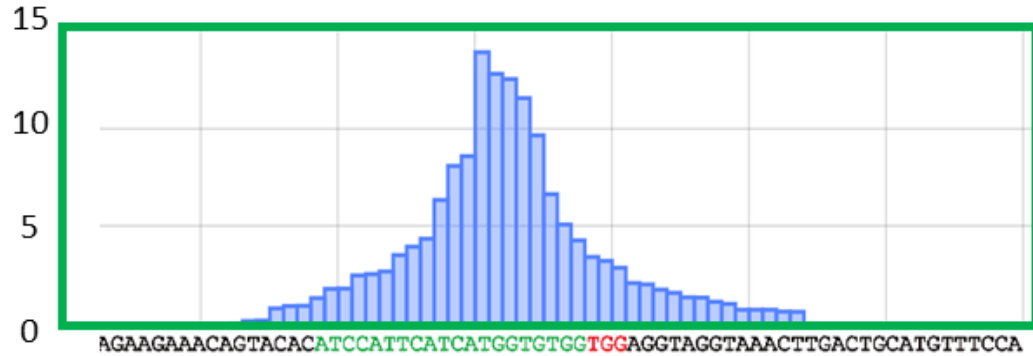
Frequency of gene deletion
(% of sequencing reads with size deletion)



RNP NC-CPP Targeting Mouse TH Gene in Mouse Glial Cells

40.3% editing in glial cells

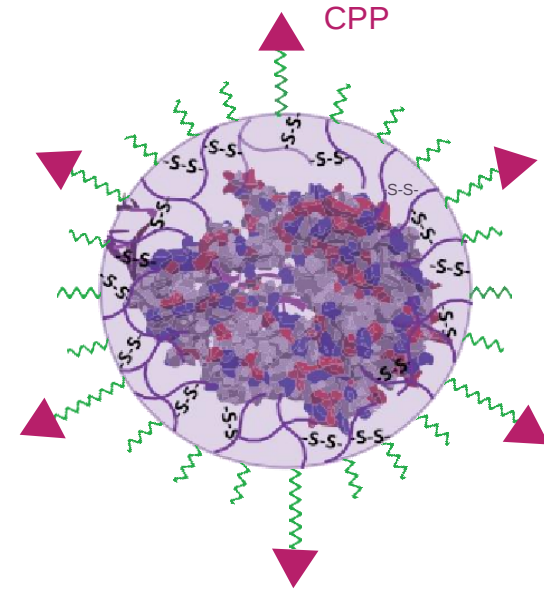
Frequency of edited base
(% of sequencing reads with base deleted)



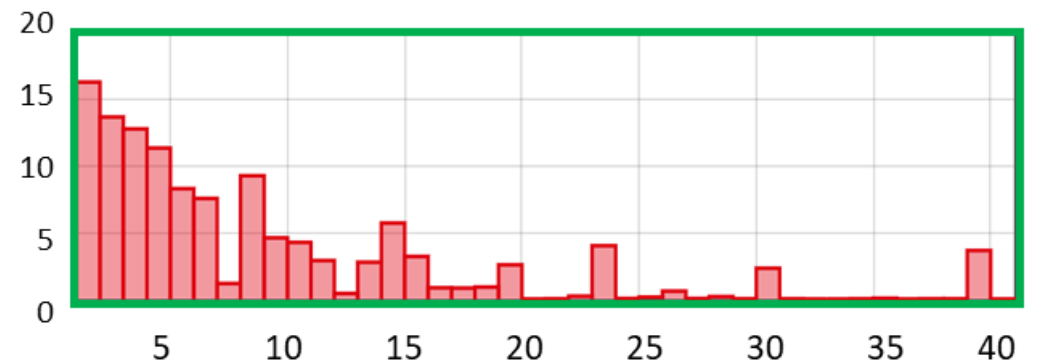
sgRNA ▲ PAM
Predicted Cut Site

TH: tyrosine hydroxylase

Preliminary data

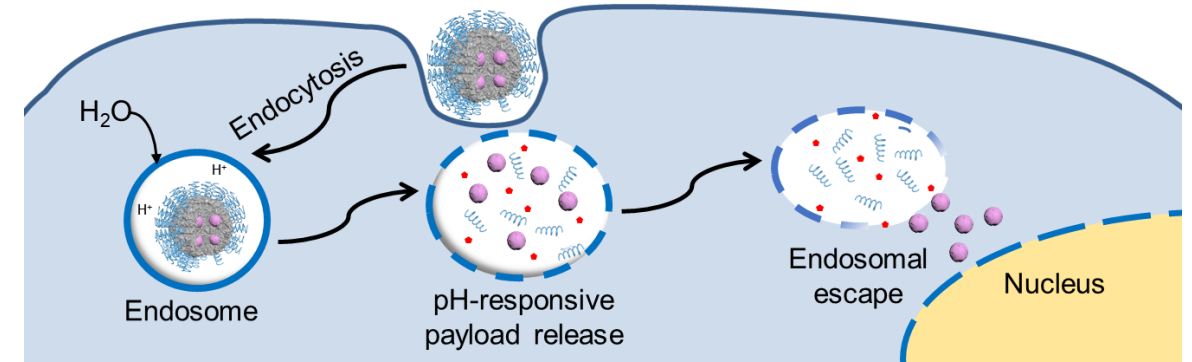
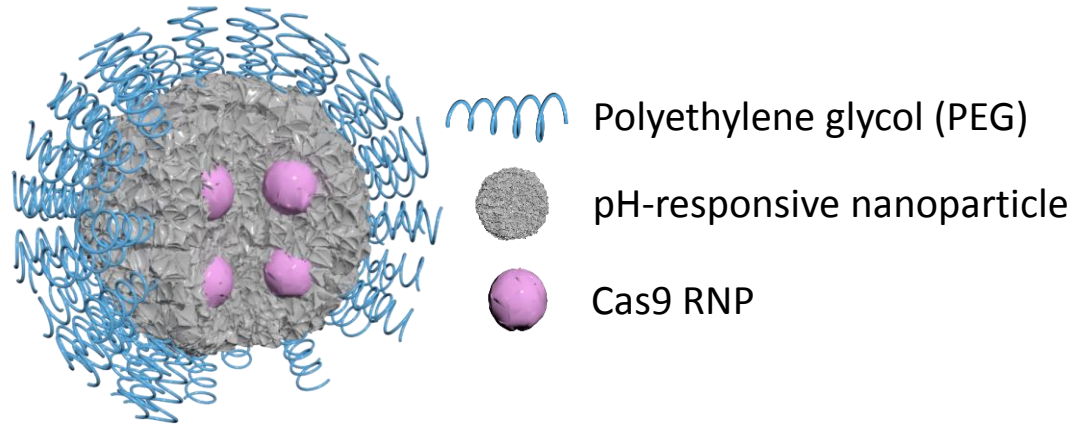


Frequency of gene deletion
(% of sequencing reads with size deletion)



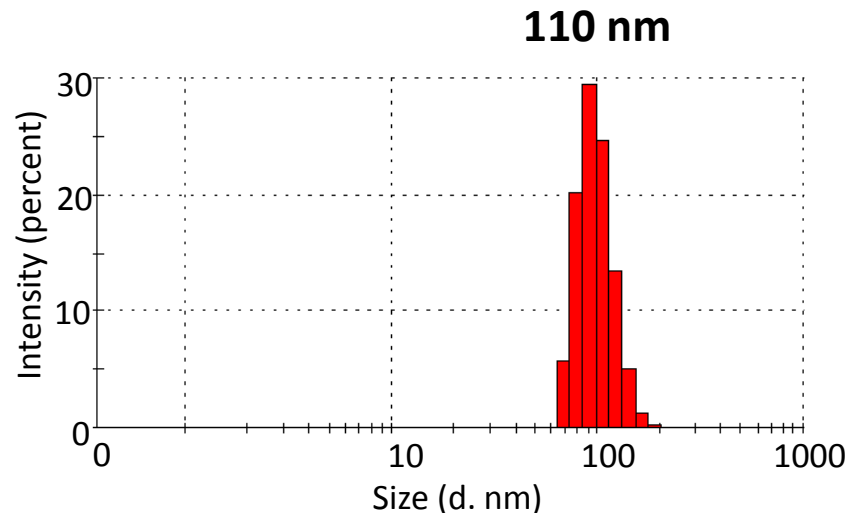
Size of Deletion

pH-Responsive Nanoparticles Enabling Both Gene Disruption and Gene Correction

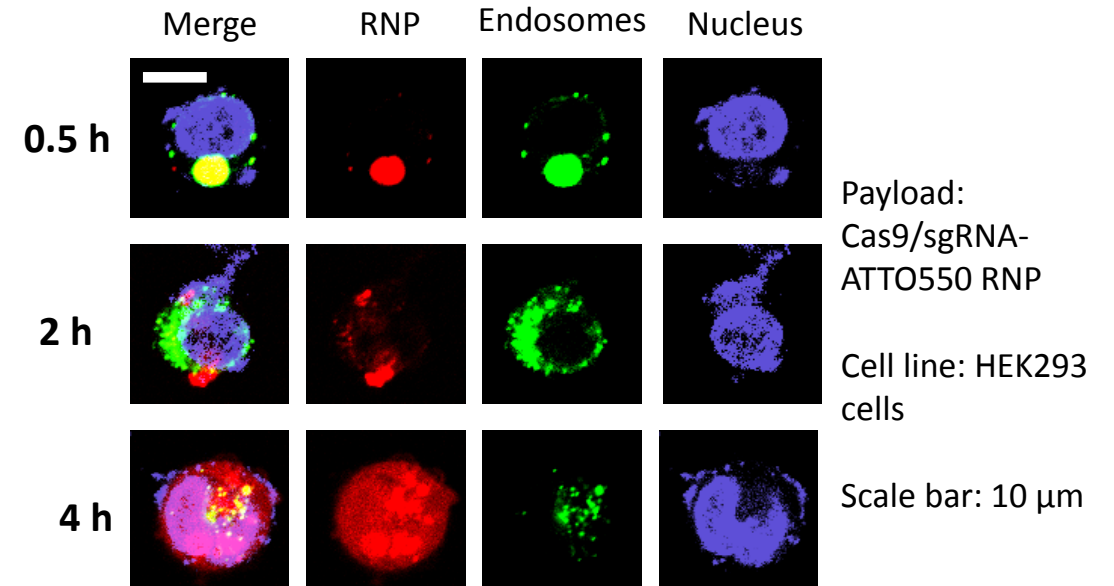


Hydrophilic payloads including DNA, mRNA, proteins, RNP, RNP + ssODN, hydrophilic small molecular drug

DLS



CLSM image

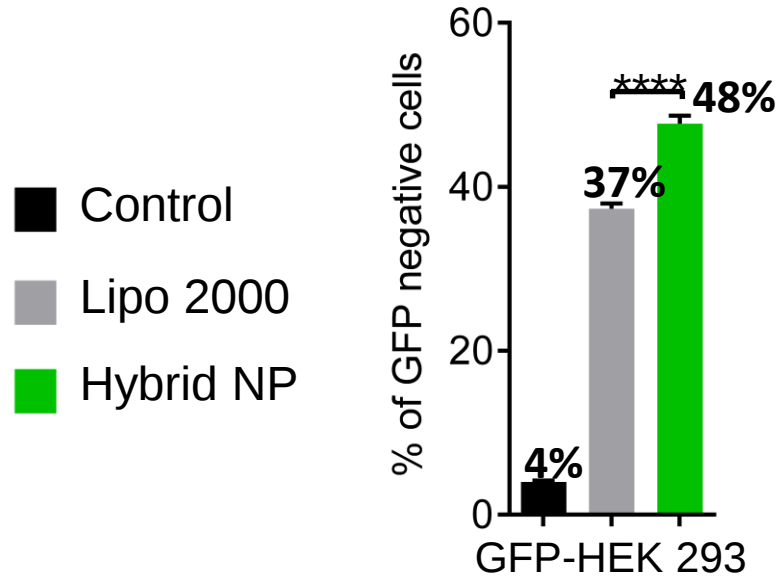


Payload:
Cas9/sgRNA-
ATTO550 RNP

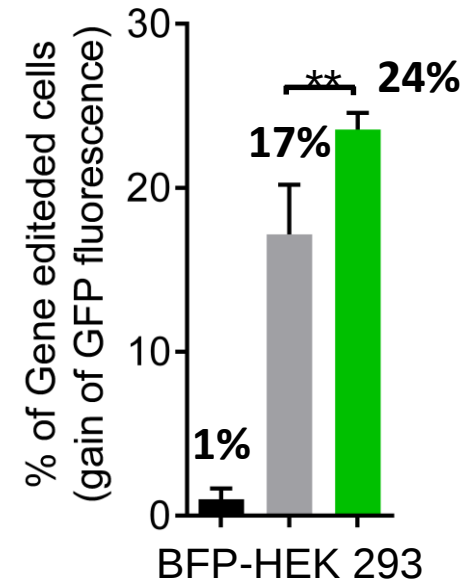
Cell line: HEK293
cells

pH-Responsive Nanoparticles Enabling Both Gene Disruption and Gene Correction

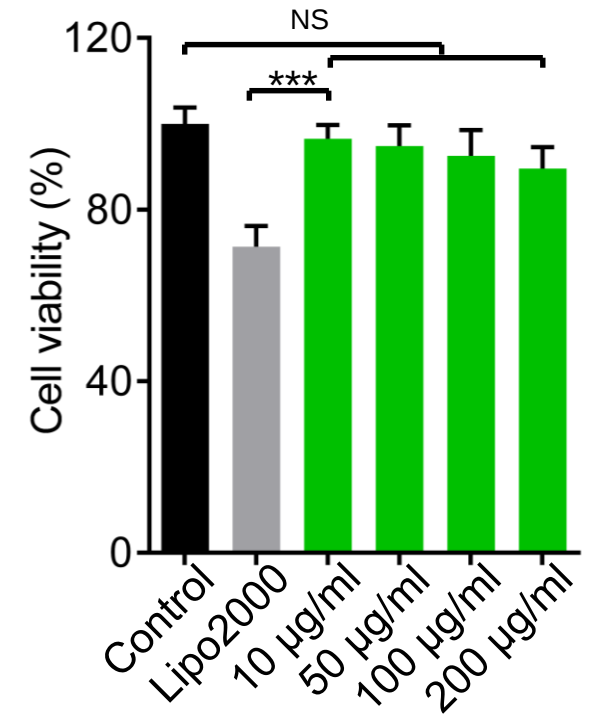
Payload: RNP
Gene Disruption



Payload: RNP + ssODN
Gene Correction/Precise editing



Cytotoxicity

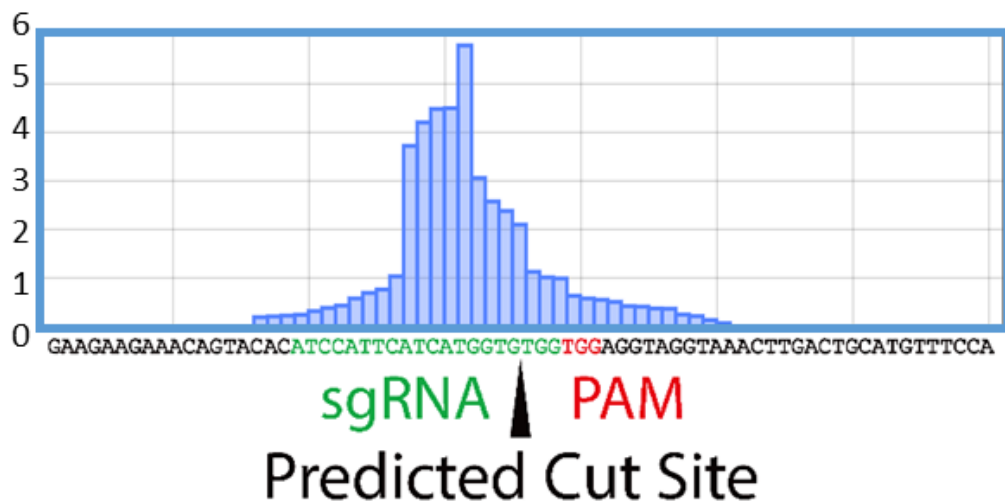


➤ Low cytotoxicity

pH-Responsive Nanoparticles Targeting Mouse APP Gene in Primary Mouse Neurons

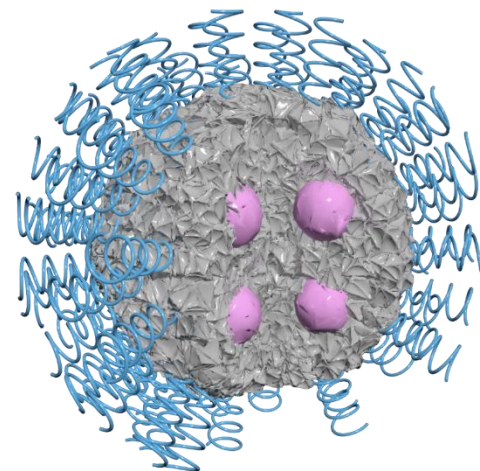
7.2% editing in primary neurons

Frequency of edited base
(% of sequencing reads with base deleted)

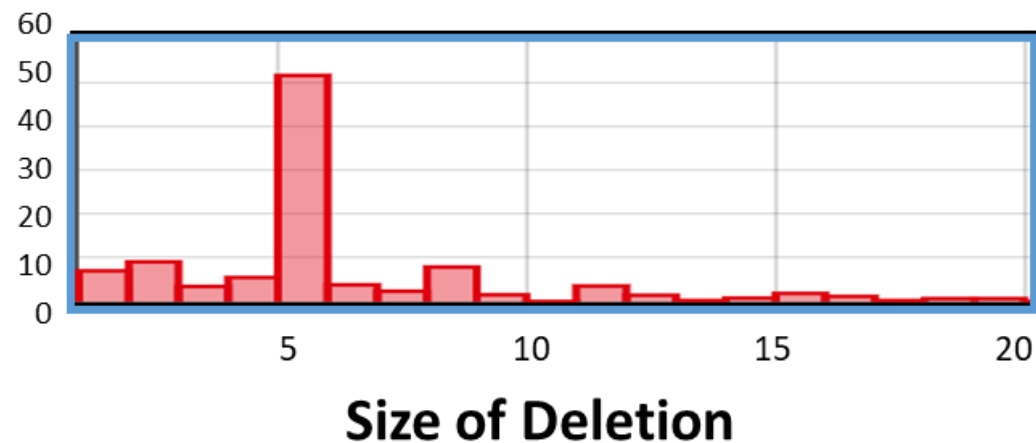


APP: amyloid precursor protein

Preliminary data

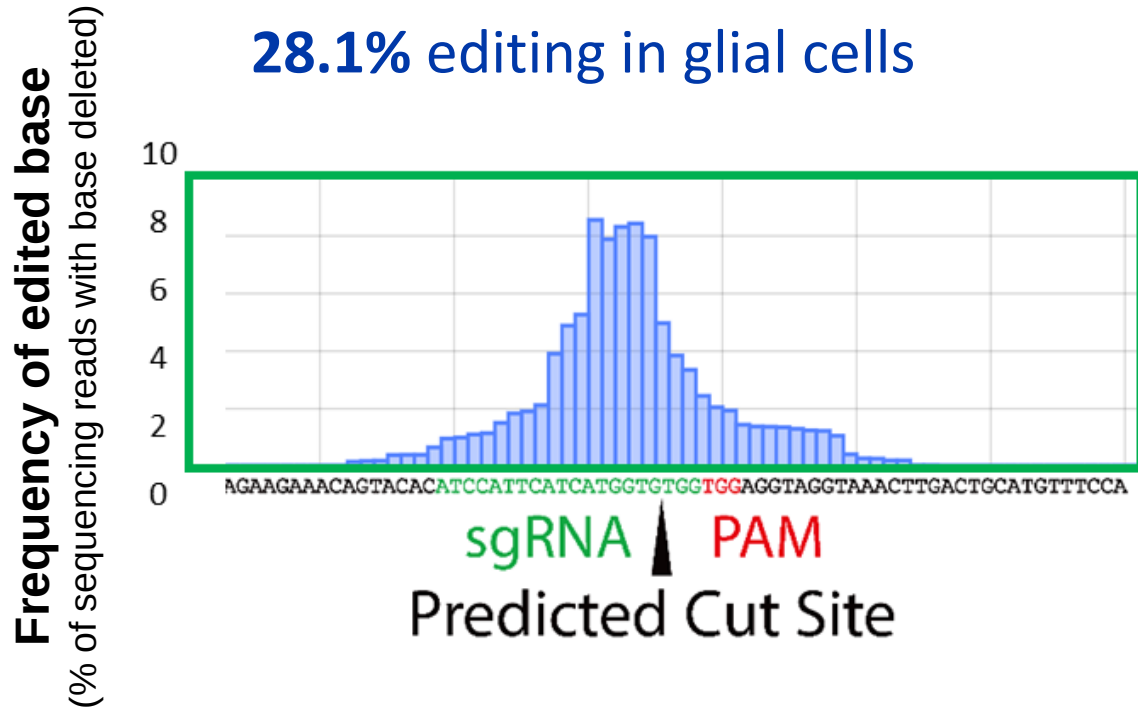


Frequency of gene deletion
(% of sequencing reads with size deletion)

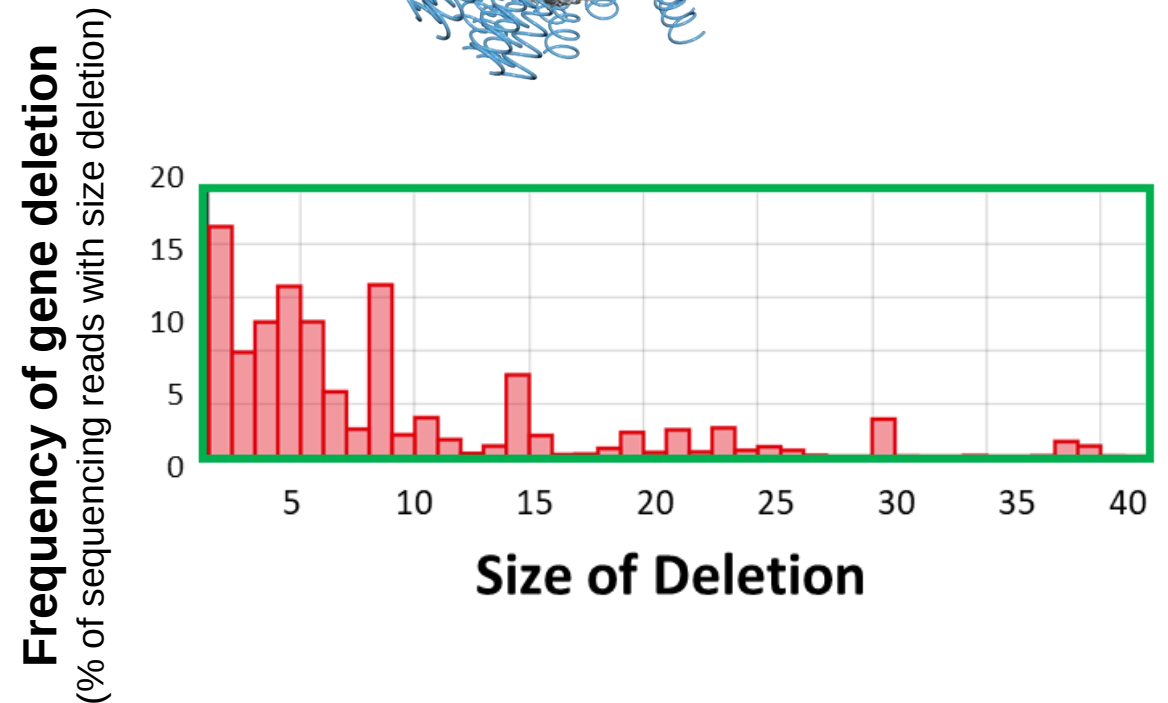
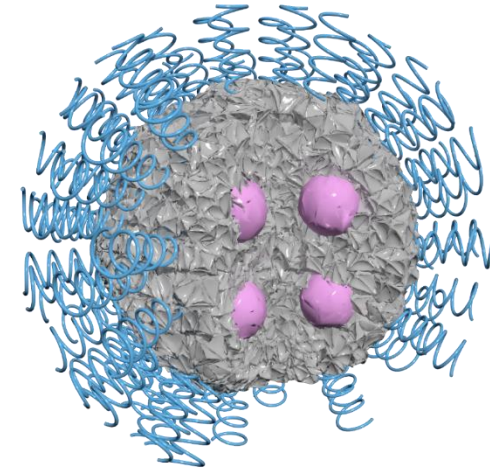


pH-Responsive Nanoparticles Targeting Mouse TH Gene in Mouse Glial Cells

28.1% editing in glial cells

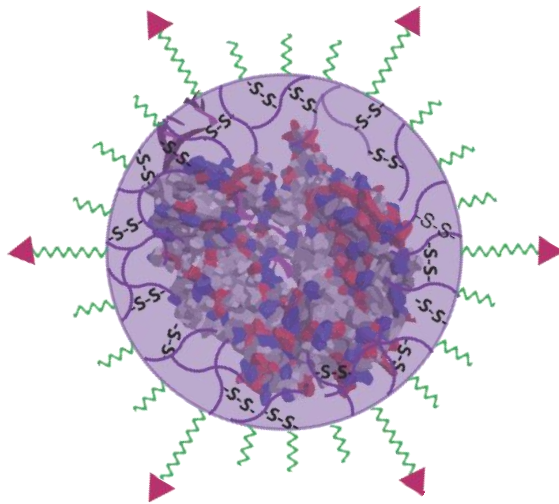


Preliminary data

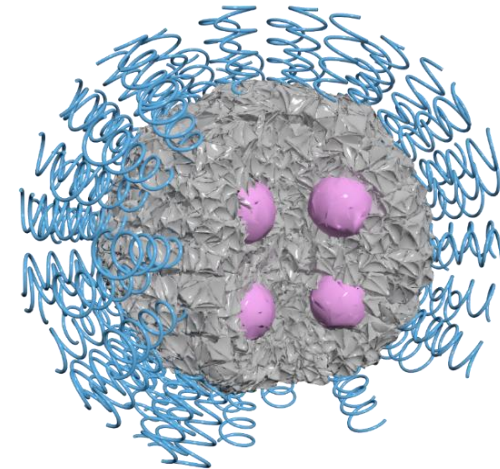


Summary

- CRISPR-Cas9 genome editing offers great promise to the field of neuroscience-both as a research tool and a treatment for various brain-related disorders.



~25 nm
RNP
Nanocapsule



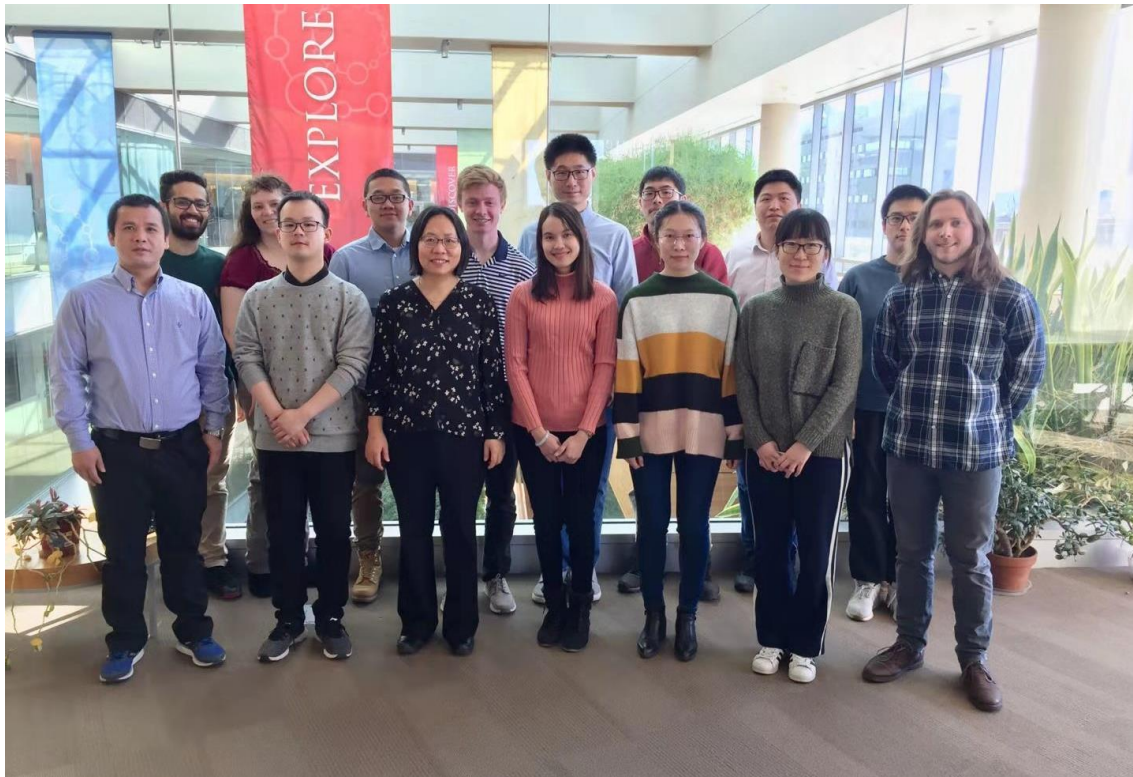
~110 nm
pH-responsive
nanoparticle
RNP, RNP + ssODN

- Capable of gene disruption
 - Low cytotoxicity
- Capable of gene disruption and gene correction
 - Low cytotoxicity
- Future research will focus on further optimizing the performance of these nanoplatforms *in vivo*

Acknowledgments

The Gong Lab

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UW Collaborators

Prof. Krishanu Saha

Dr. Amr Abdeen

Kirstan Gimse

Prof. Subhojit Roy

Dr. Pankaj Dubey

Prof. Jon Levine

Dr. Matthew Flowers

Prof. Marina Emborg

Megan Murphy

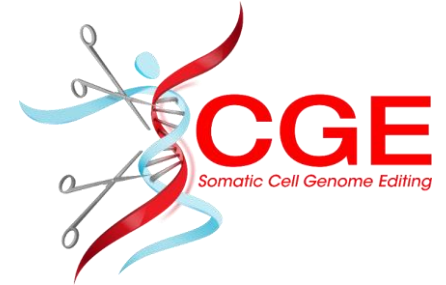
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