



# Developing algorithms for single cell genomics

**Elana J. Fertig, PhD**

Associate Director of the Convergence Institute

Assistant Director of the Research Program in Quantitative Sciences

Associate Professor of Oncology, Biomedical Engineering, and Applied Mathematics and Statistics

Sidney Kimmel Comprehensive Cancer Center - Johns Hopkins University

@FertigLab

# What are single cell technologies?

Bulk RNA-Seq



Single cell RNA-Seq



Spatial transcriptomics



# Single cell RNA-seq data of retinal development

Cell Type

Müller glia

Amacrine

Neurogenic

Cones

PR precursors

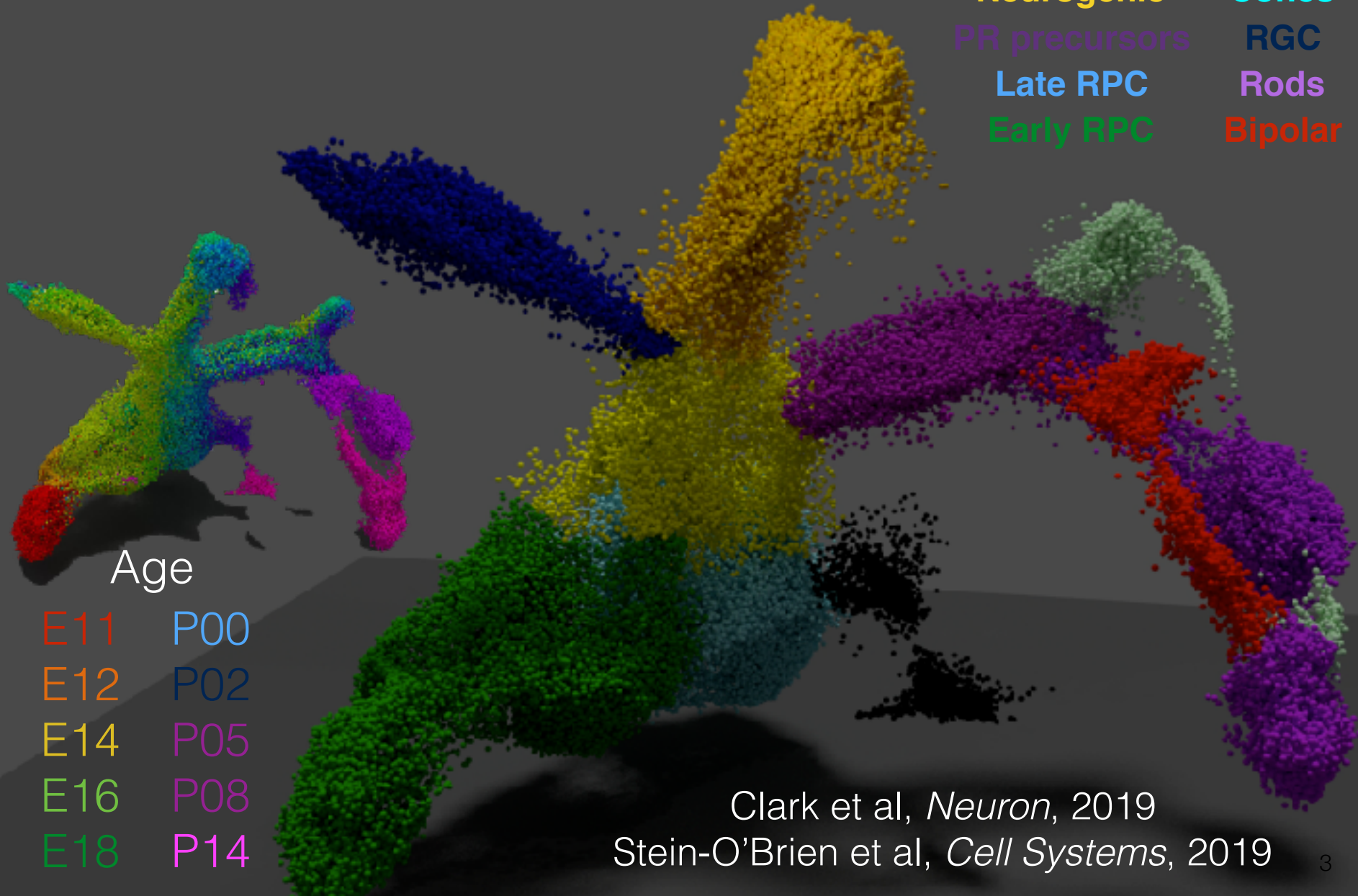
RGC

Late RPC

Rods

Early RPC

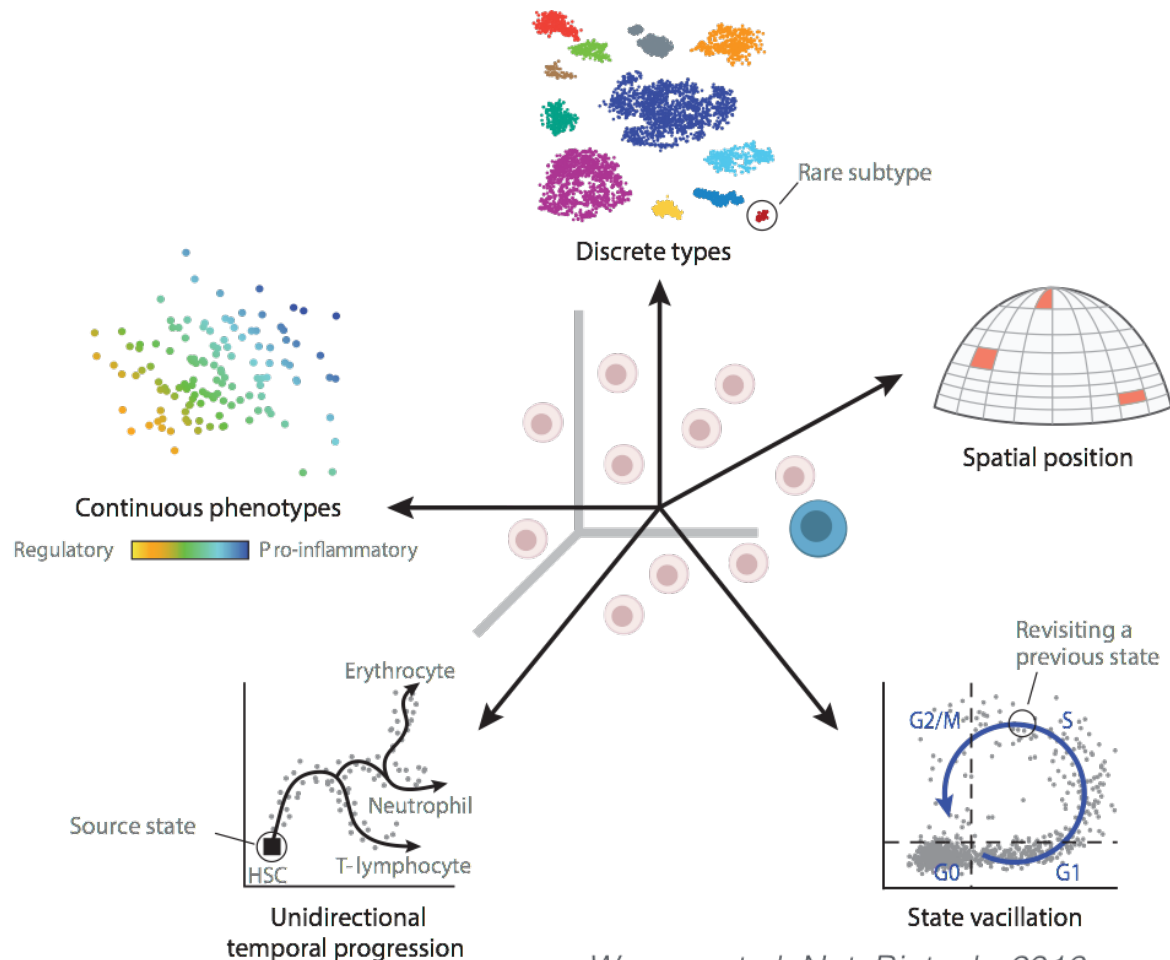
Bipolar



Clark et al, *Neuron*, 2019

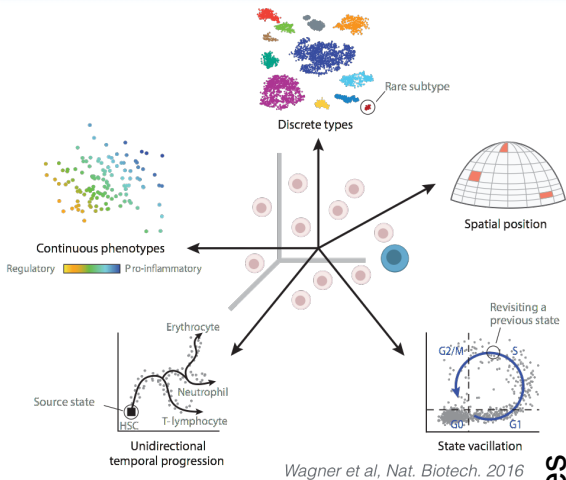
Stein-O'Brien et al, *Cell Systems*, 2019

# In addition to cell types, single cell data also captures axes of cellular activity

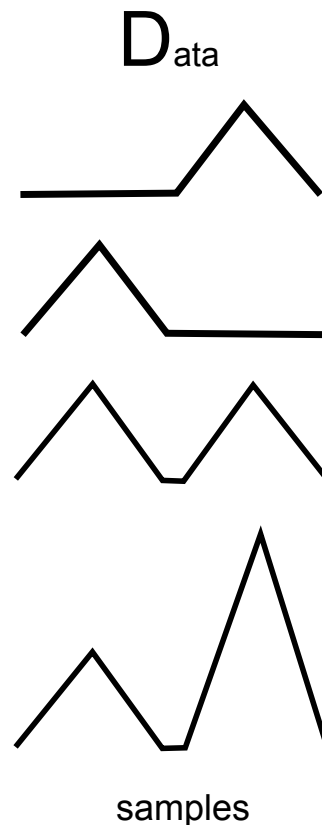


Wagner et al, Nat. Biotech. 2016

# Axes of cellular identity mirrored by CoGAPS matrix factorization



genes



$$= A_{\text{Amplitudes}} \times P_{\text{patterns}}$$

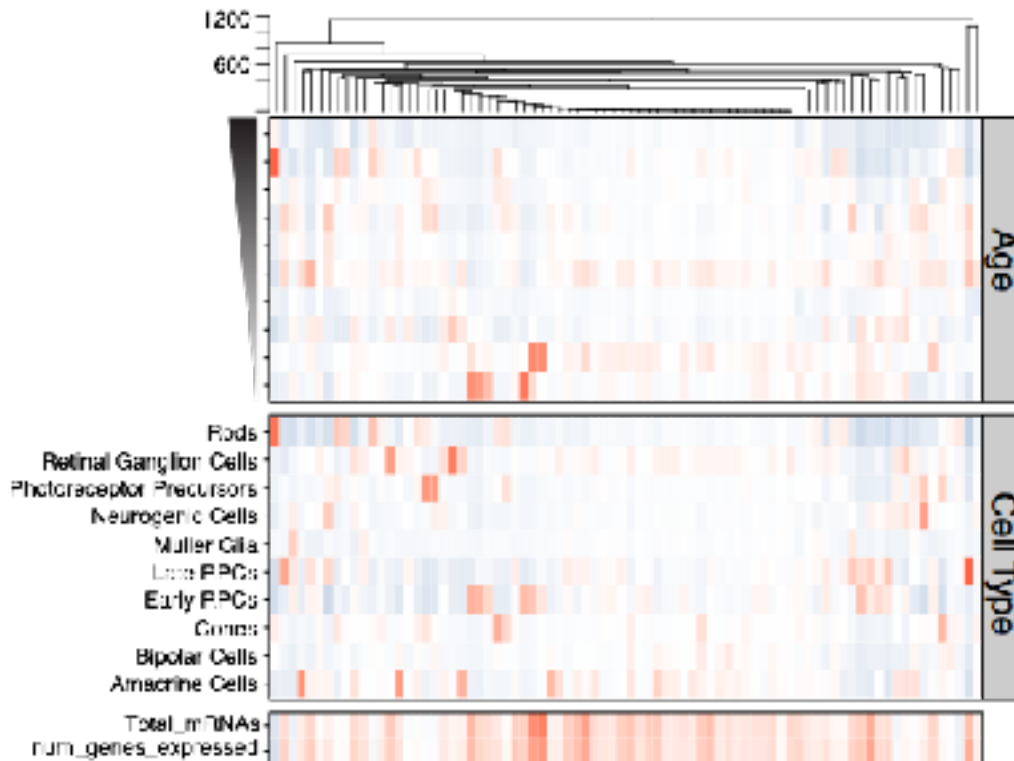
(gene weights for each biological process) (biological process in each sample)

pattern marker genes

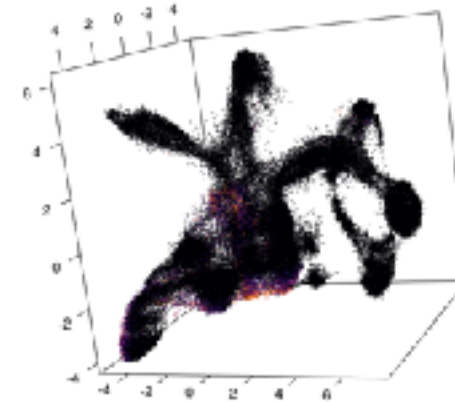
$$= \begin{matrix} \text{genes} \\ \begin{bmatrix} 0 & 1 \\ 1 & 0 \\ 1 & 1 \\ 1 & 2 \end{bmatrix} \end{matrix} \times \begin{matrix} \text{samples} \\ \begin{bmatrix} \text{Pattern 1} \\ \text{Pattern 2} \end{bmatrix} \end{matrix}$$

The algorithm looks for patterns that define the biological activity in a set of samples (e.g., time points) and the amplitude at which each gene contributes to that biological process.

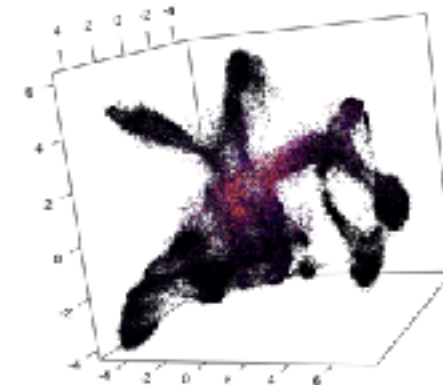
# Example of how matrix factorization uncovers axes of cellular identity in retinal development



Cell Cycle - G1/S



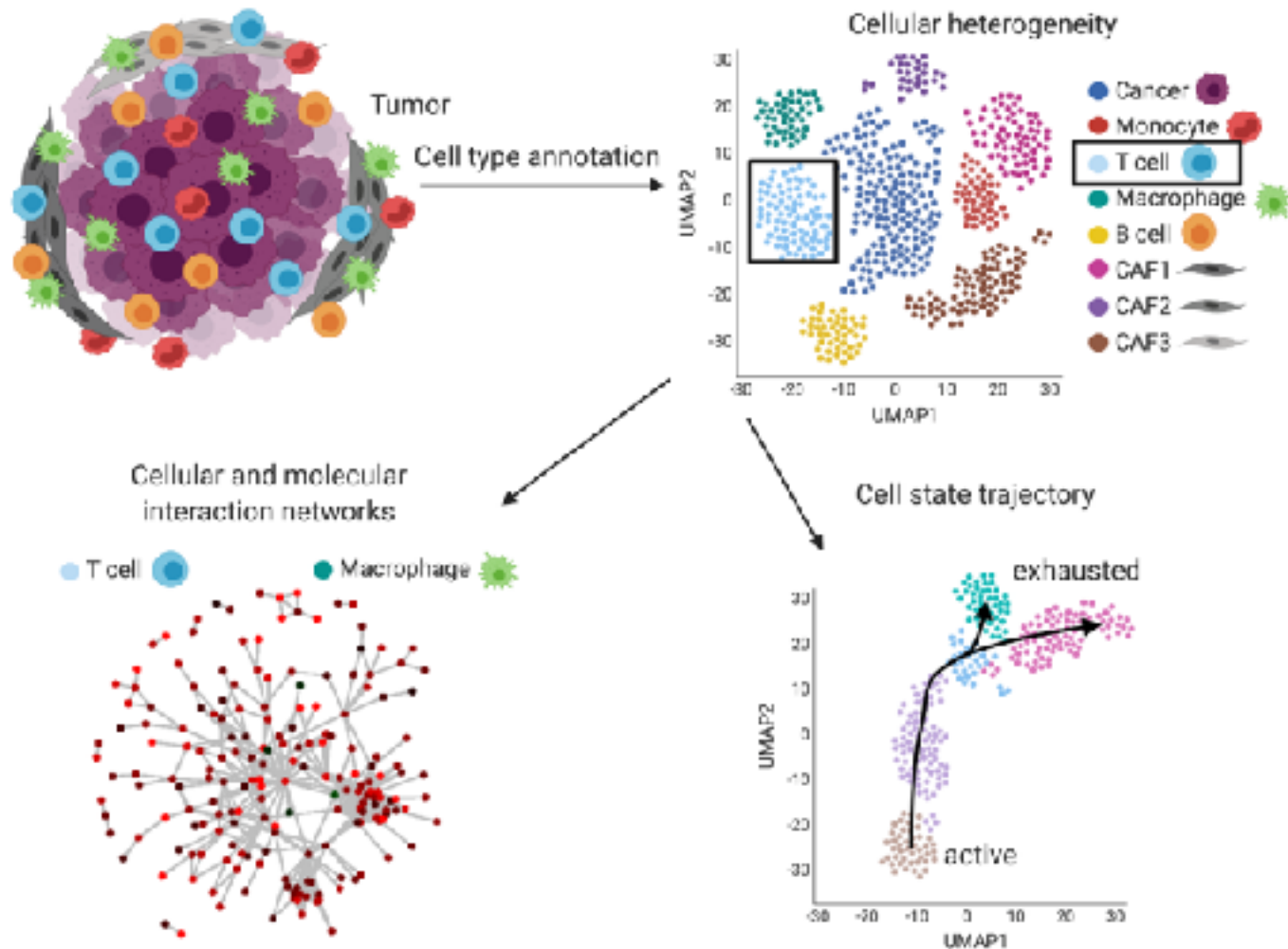
Neurogenic



Clark et al, *Neuron*, 2019

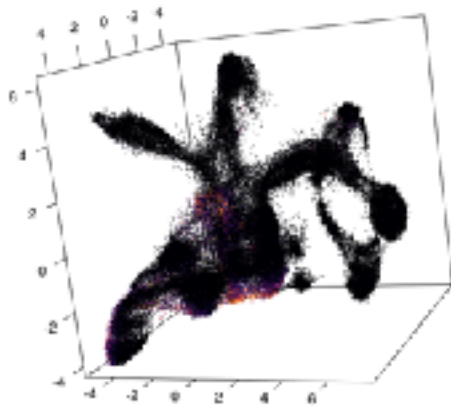
Stein-O'Brien et al, *Cell Systems*, 2019

# Inference of cellular state transitions and interactions requires new algorithms

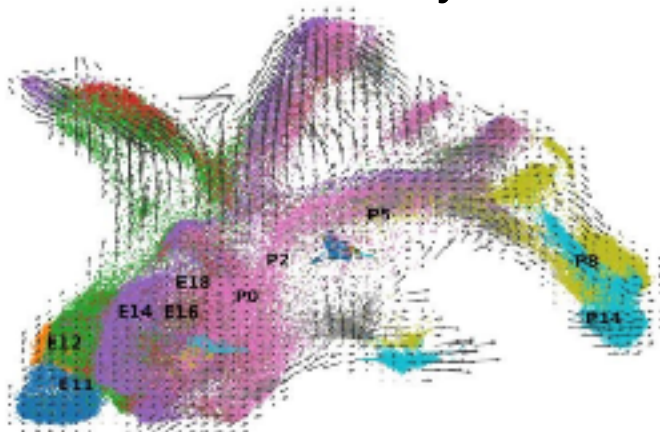


# Cell state transitions

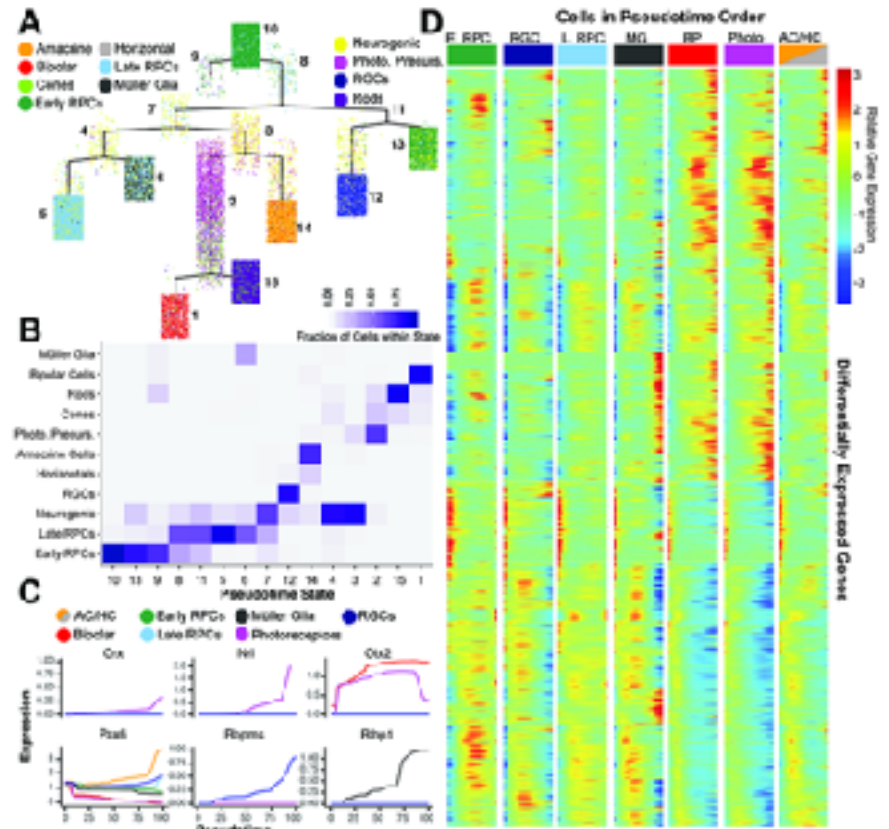
## Patterns of cell cycle - G1/S Trajectory inference or pseudotime



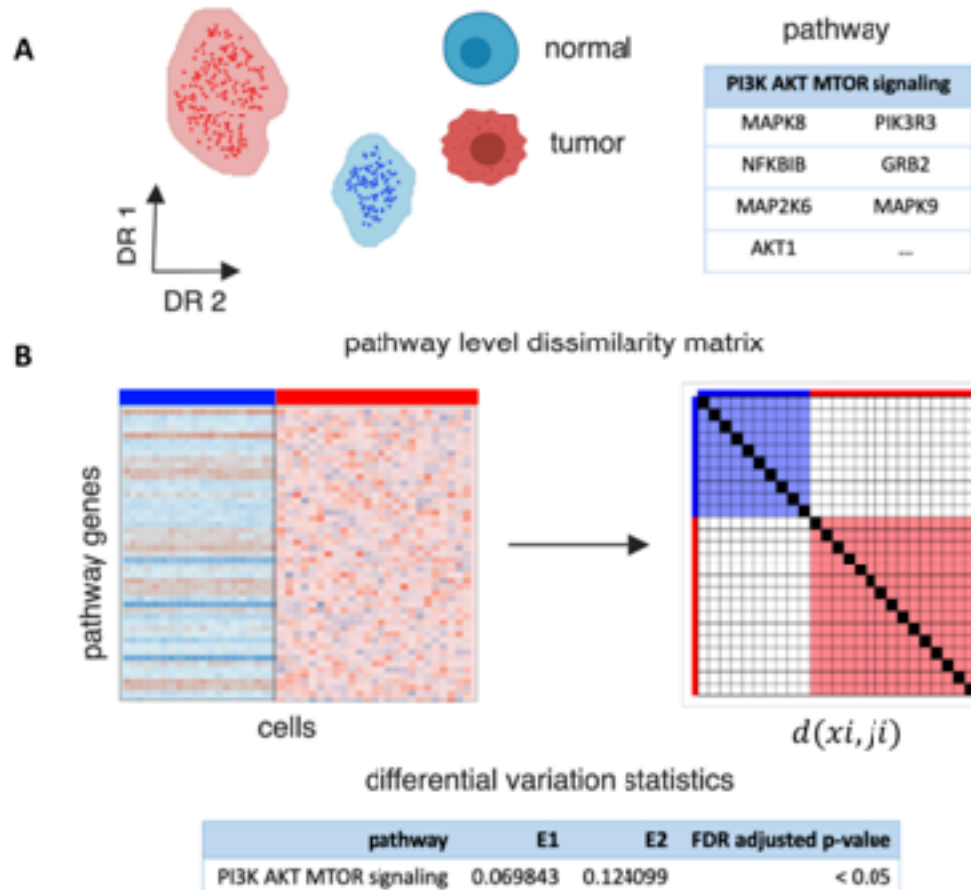
# RNA velocity



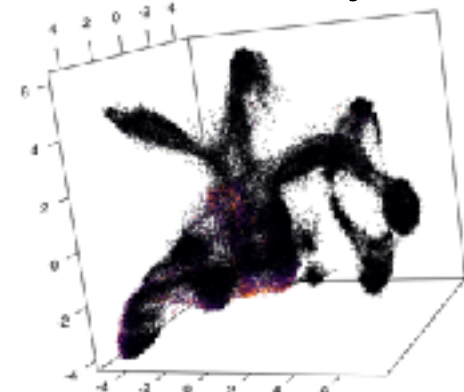
Melsted et al, *BioRxiv*, 2019

Clark et al, *Neuron*, 2019Stein-O'Brien et al, *Cell Systems*, 2019<sup>8</sup>

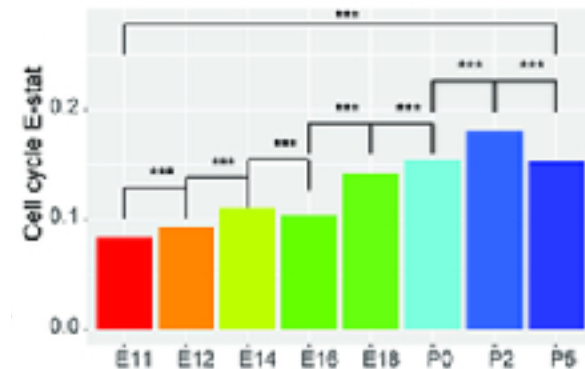
# Molecular heterogeneity can occur within distinct cellular subtypes



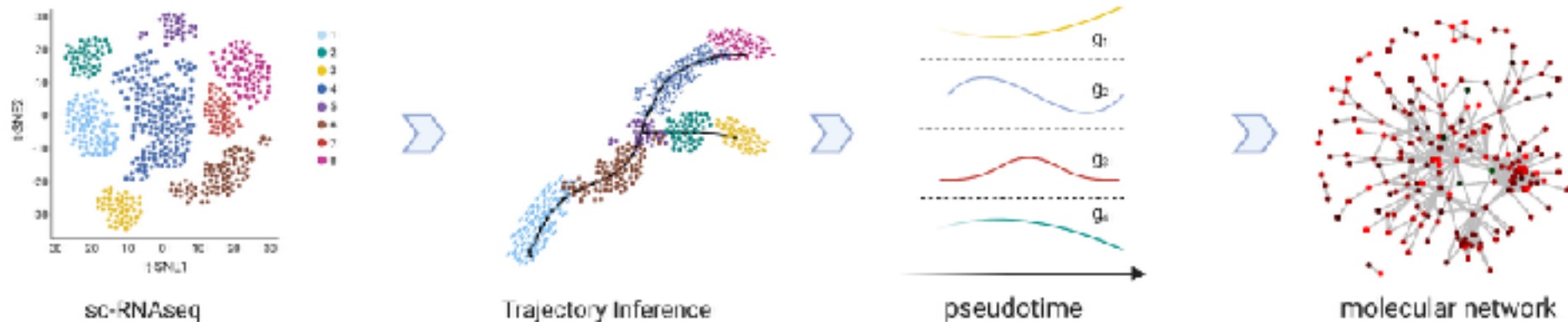
Patterns of cell cycle - G1/S



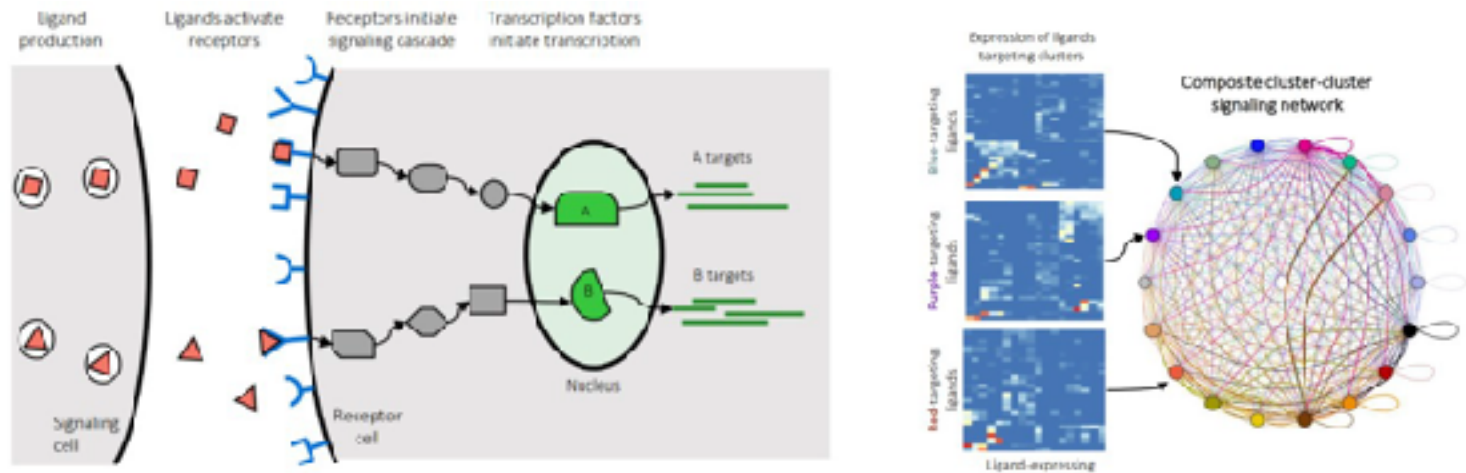
Temporal dysregulation of cell cycle



# Interaction networks can be informed by cell state transitions or regulatory networks



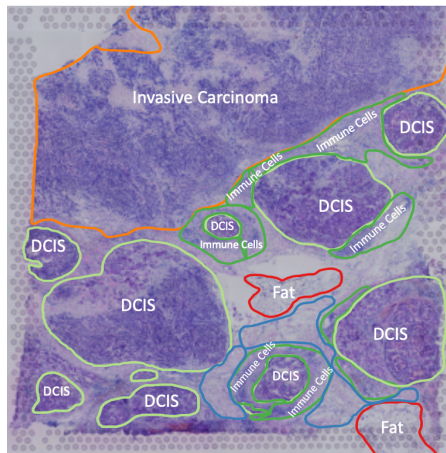
Deshpande et al, *BioRxiv*, 2019



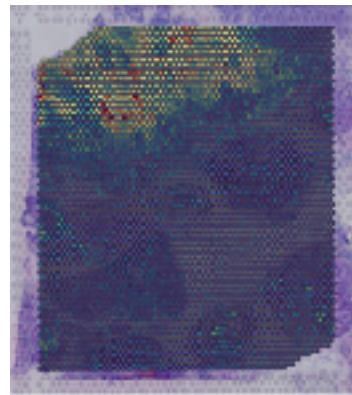
Cherry et al, *BioRxiv*, 2020

# Inter-cellular interactions can be further inferred with spatial transcriptomics

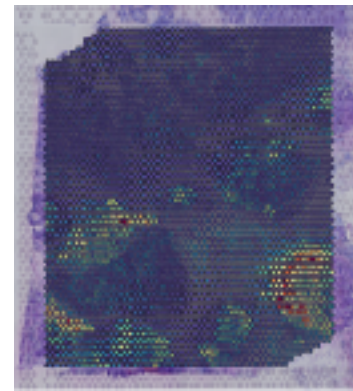
Visium  
IHC + expression



tumor patterns

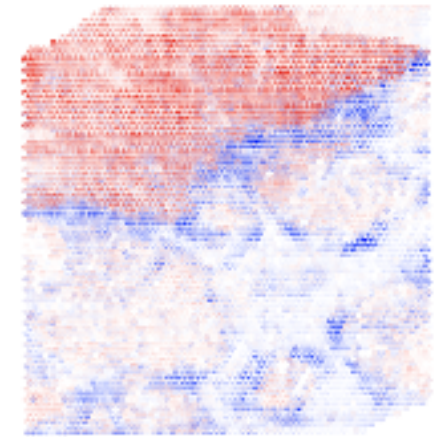


*TGFBI, VEGFA, FGFR1*



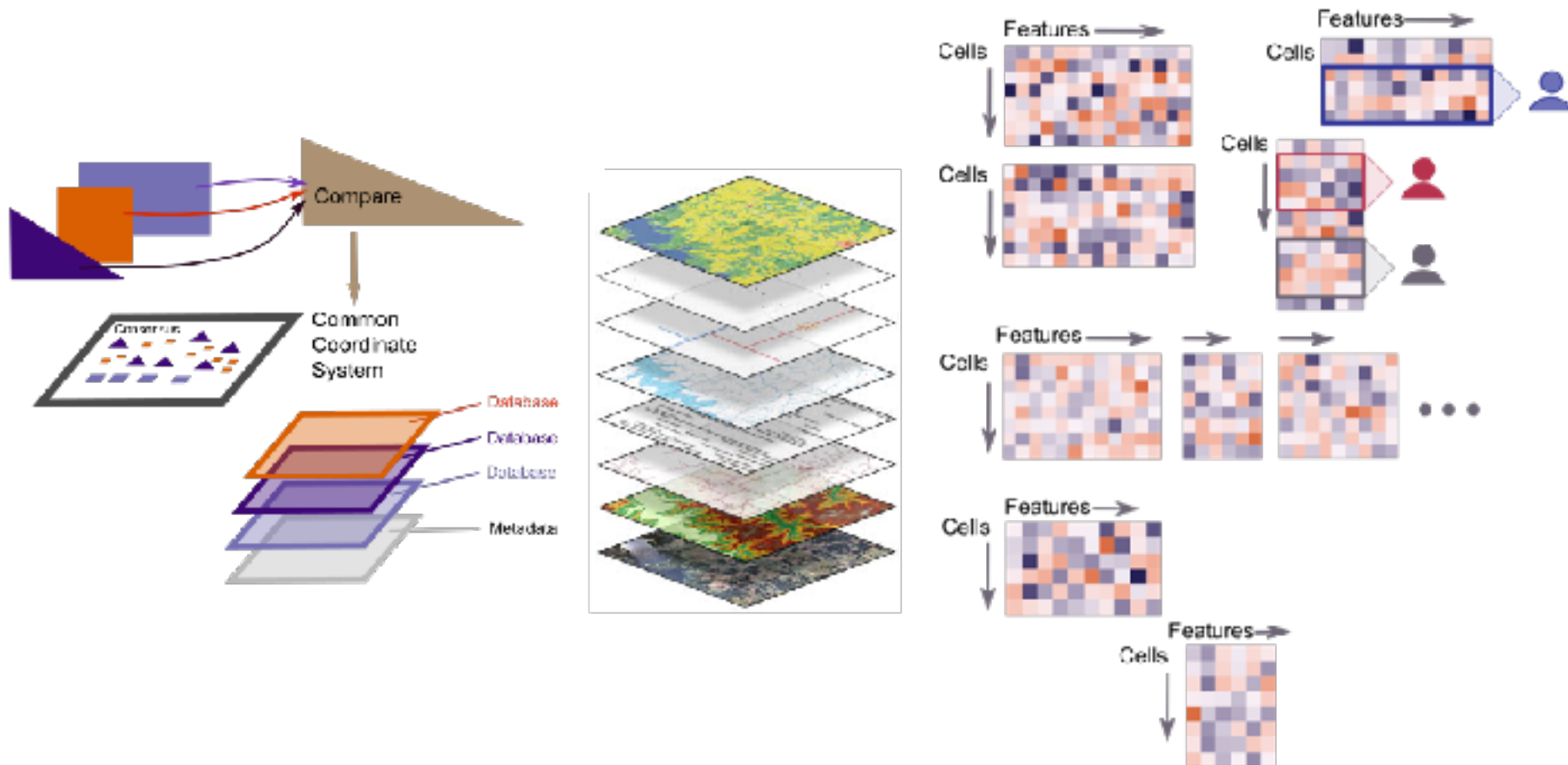
*PIK3R3*

immune / tumor  
interactions



Melanie Loth, Atul Deshpande in collaboration with 10X Genomics

# Multi-omics enables regulatory network inference and cross-study validation



Le Cao et al, 2019, <https://birsbiointegration.github.io/whitePaper/>

# Summary of methods and references



- **Matrix factorization and latent space analysis**

- Cleary et al, 2017
- Zhu et al, 2017
- Townes et al, 2017
- Stein-O'Brien et al, 2018
- Stein-O'Brien et al, 2019
- Welch et al, 2019
- Mohammadi et al, 2019
- Svensson et al, 2020

- **Heterogeneity analysis**

- Azizi et al, 2018
- Davis-Marcisak et al, 2019

- **Trajectory and velocity analyses**

- Trapnell et al, 2014
- Ji et al, 2016
- La Mannno et al, 2018
- Bergen et al, 2020
- Saalens et al, 2019

- **Network analyses**

- Efremova et al, 2020
- Browaeys et al, 2020
- Cherry et al, BioRxiv, 2020
- Deshpande et al, BioRxiv, 2019