

Cellino: Your Cells, Your Cure

A ultra -scalable biomanufacturing platform for
personalized regenerative medicines

NASEM Forum on Regenerative Medicine
Washington, DC
October 17, 2023

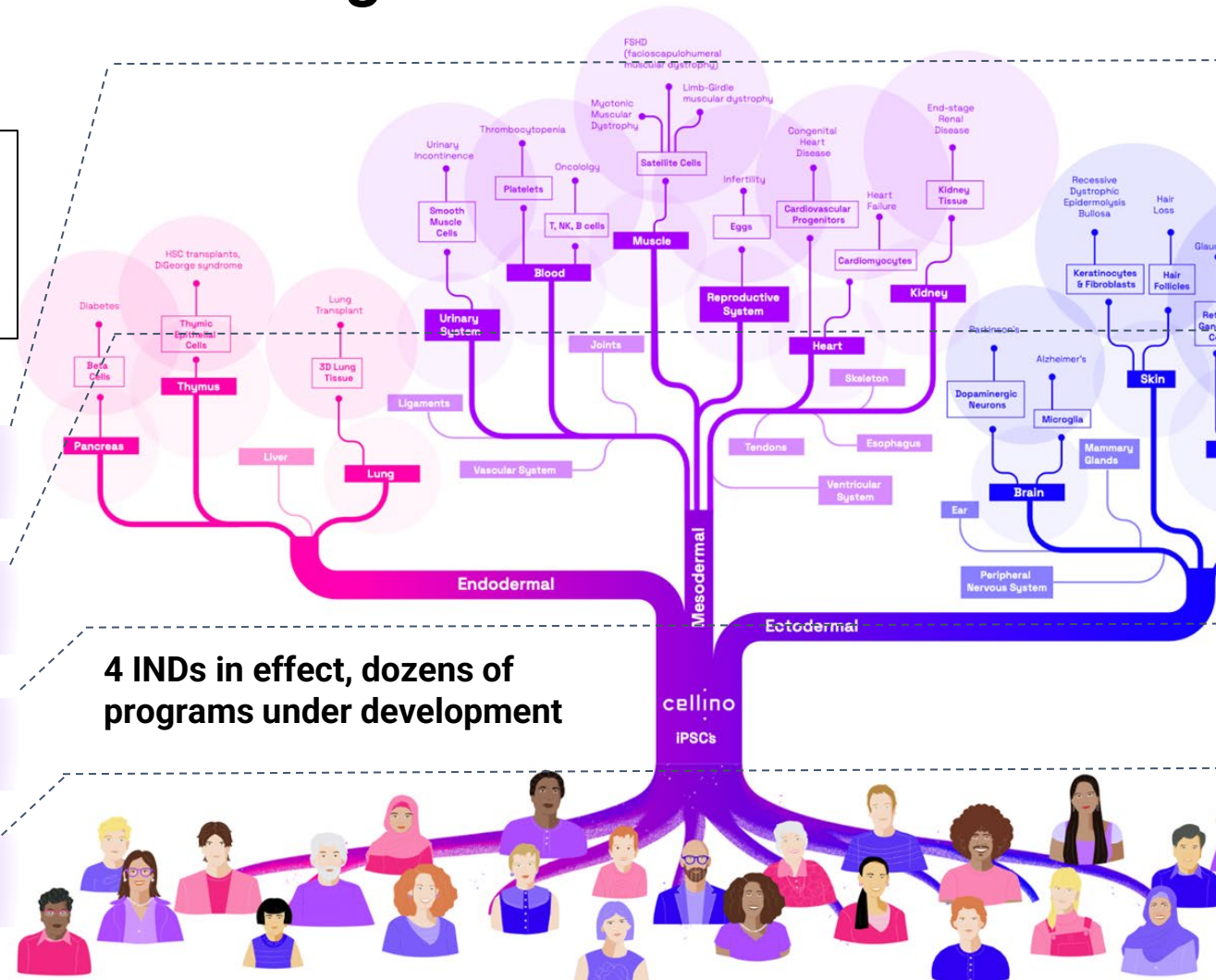
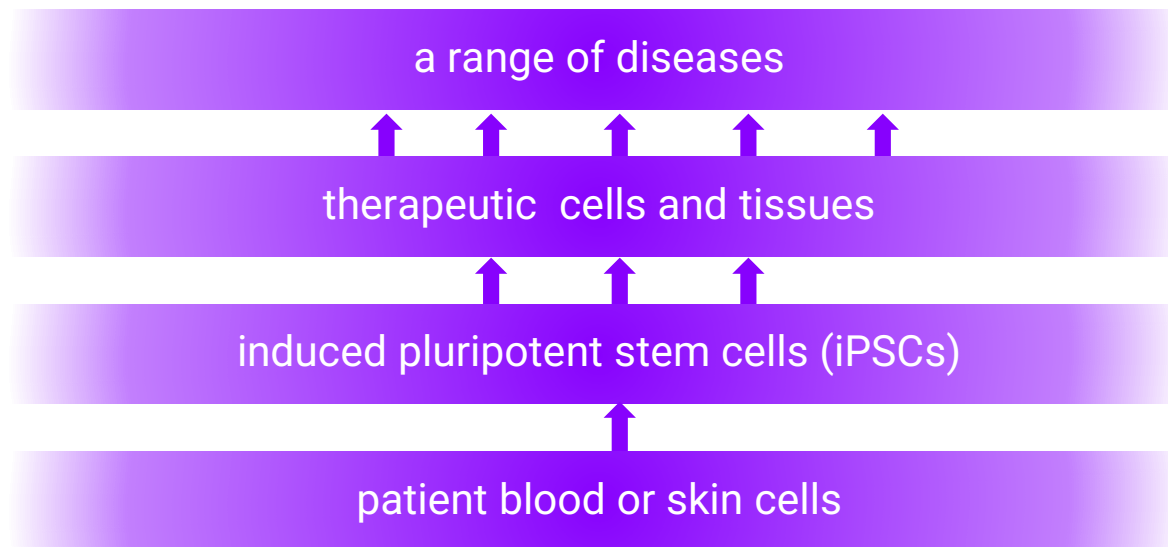
Nabiha Saklayen, Ph.D.
CEO and Co-Founder, Cellino



Autologous iPSC -derived cell therapies can address accessibility challenges associated with donor -derived regenerative medicines

Benefits

- no immunosuppression (more accessible for aging & immuno-compromised population)
- no donor matching (more accessible for genetically diverse population)



4 INDs in effect, dozens of programs under development

Current manufacturing limited by expert manual operations, open bioprocesses, and high -grade cleanrooms

OPEN BIOPROCESSES IN CLEANROOMS



*manual culture, scraping,
and transfers by hand*



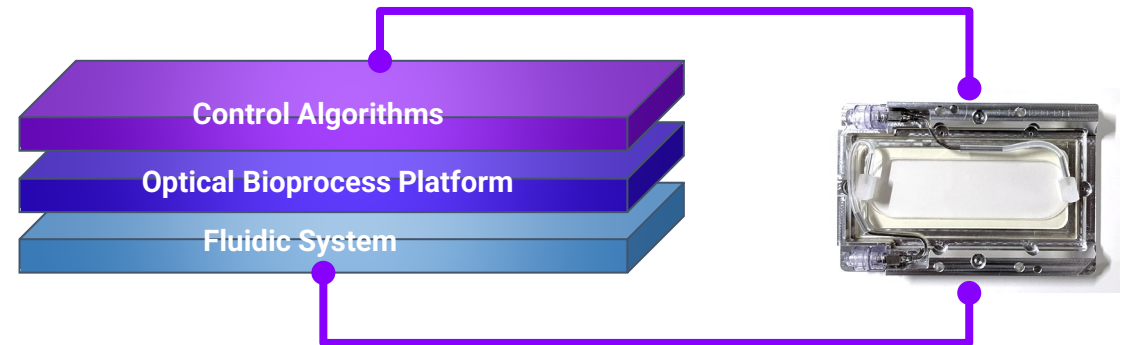
*Visual inspection and decisions
with a microscope*

REPEATED OVER 30-140 days (depending on cell type)

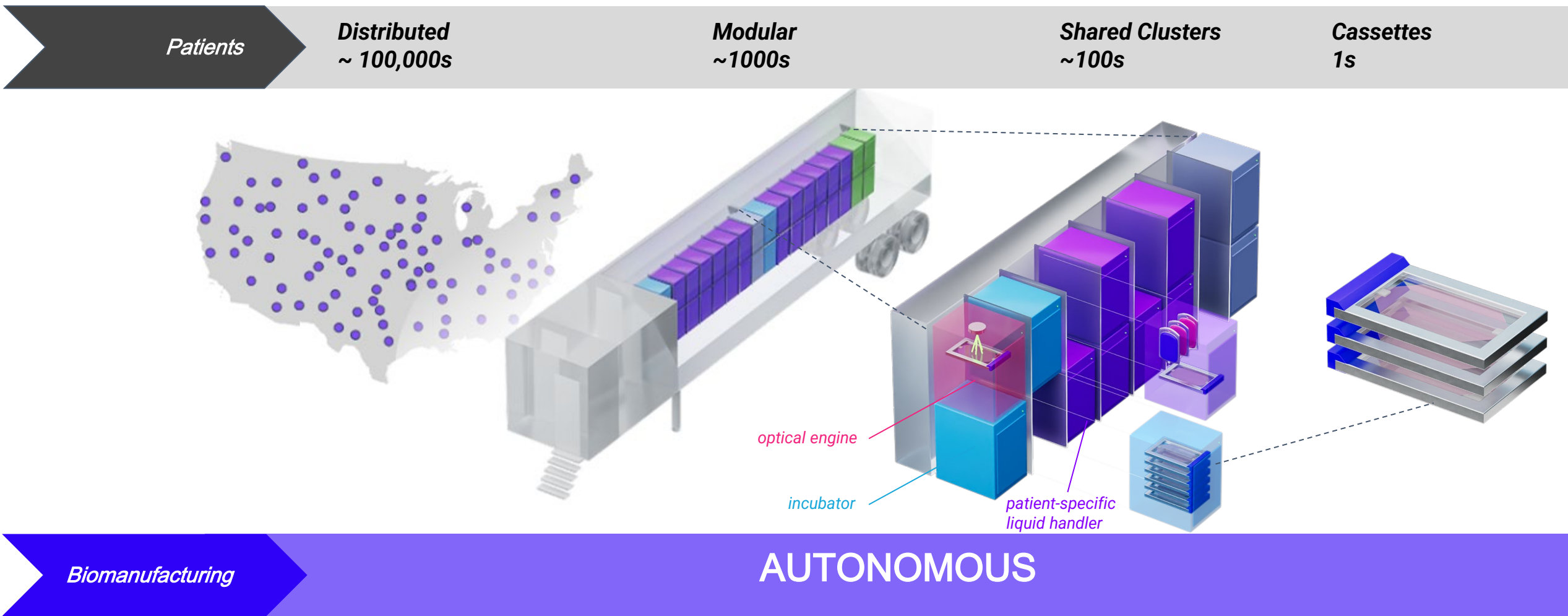
Recent advances in AI, optics, and fluidics enable the Cellino platform to be autonomous, closed, and modular

MANUAL

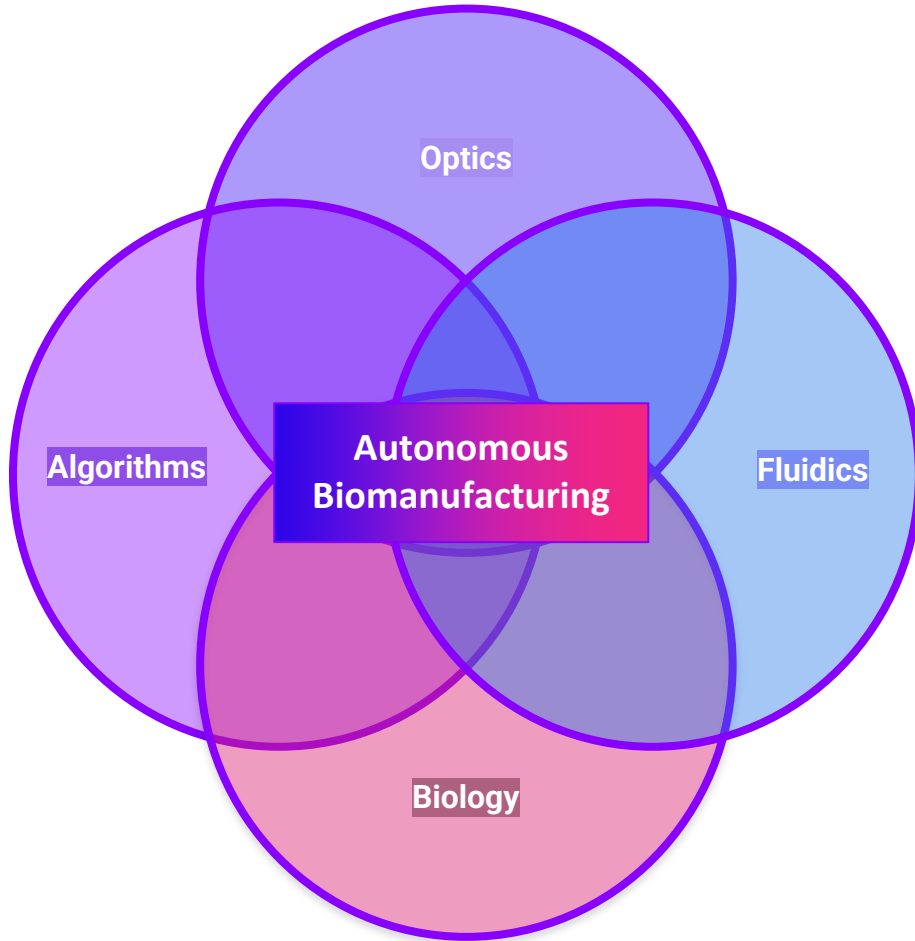
CLOSED
AUTONOMOUS
MODULAR



Cellino vision: Point -of-care, closed, and autonomous biomanufacturing to expand access to auto iPSC -derived therapies



Cellino's multidisciplinary team is building an autonomous biomanufacturing platform for patient access



Industry Experience

Biomanufacturing: preclinical/clinical/commercial manufacturing for cell therapies

Pharma/biotech: cell therapies, molecular biology, genomics

Med devices: 25 medical devices approved + serving over 500,000 patients

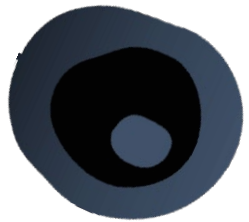
Photonics/semiconductors: w/ \$500MM+ products shipped

AI/ML: high-resolution, image-guided AI technologies in defense

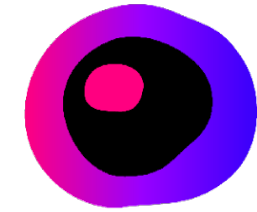
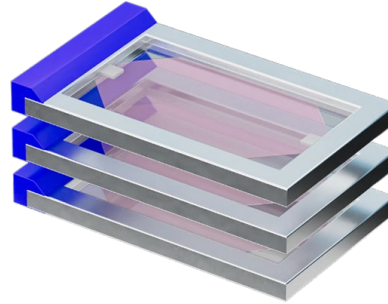
Fluidics: oil and gas, organ-on-chip



The platform makes in-process decisions using image-guided algorithms and removes unwanted cells using laser-induced bubbles



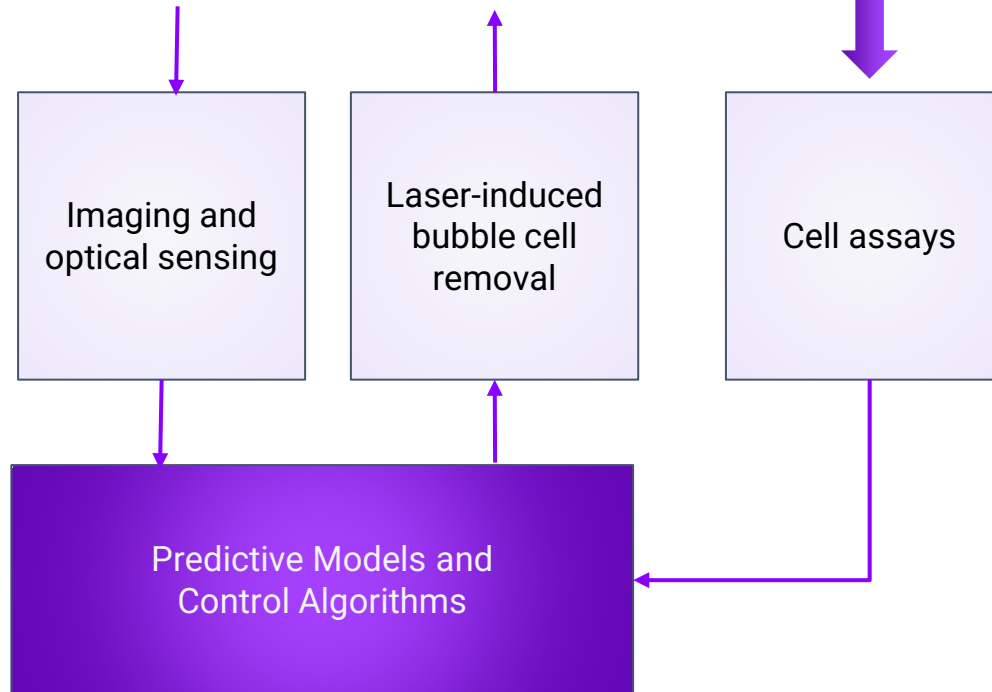
Cell Type A



Cell Type B

Benefits

- **passage-free culture**
- **minimal expert oversight**
- **single-cell precision**
- **closed-compatible**
- ...





Example case #1: replacing operator -driven selection of colonies with AI -based prediction of colony quality

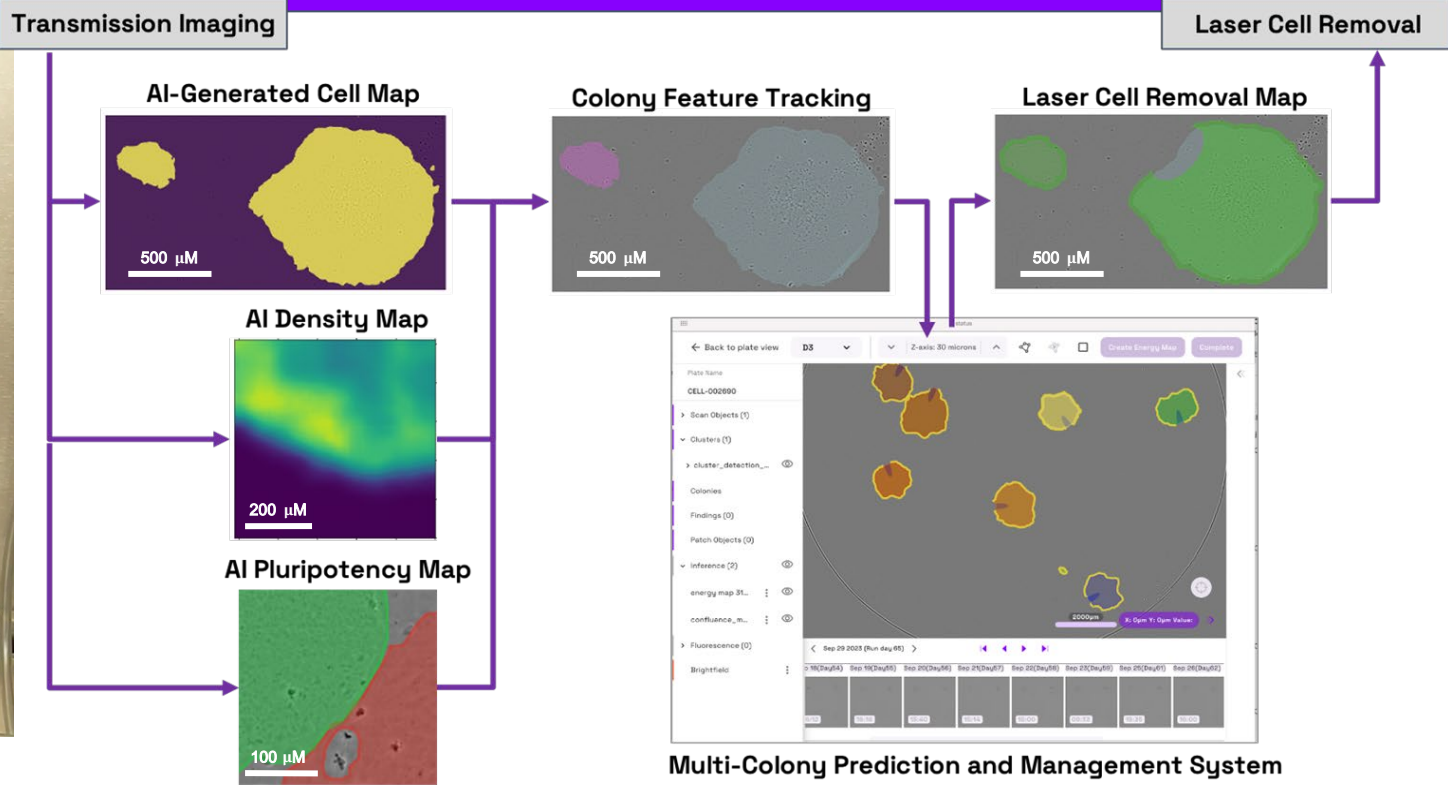
TRADITIONAL APPROACH

Human operator prediction of colony quality, manual scraping



CELLINO APPROACH

ML-based prediction of high -quality colony, ML - driven clonal management



Example case #2: replacing operator -driven well -to -well passaging with ML -driven laser -based cell removal

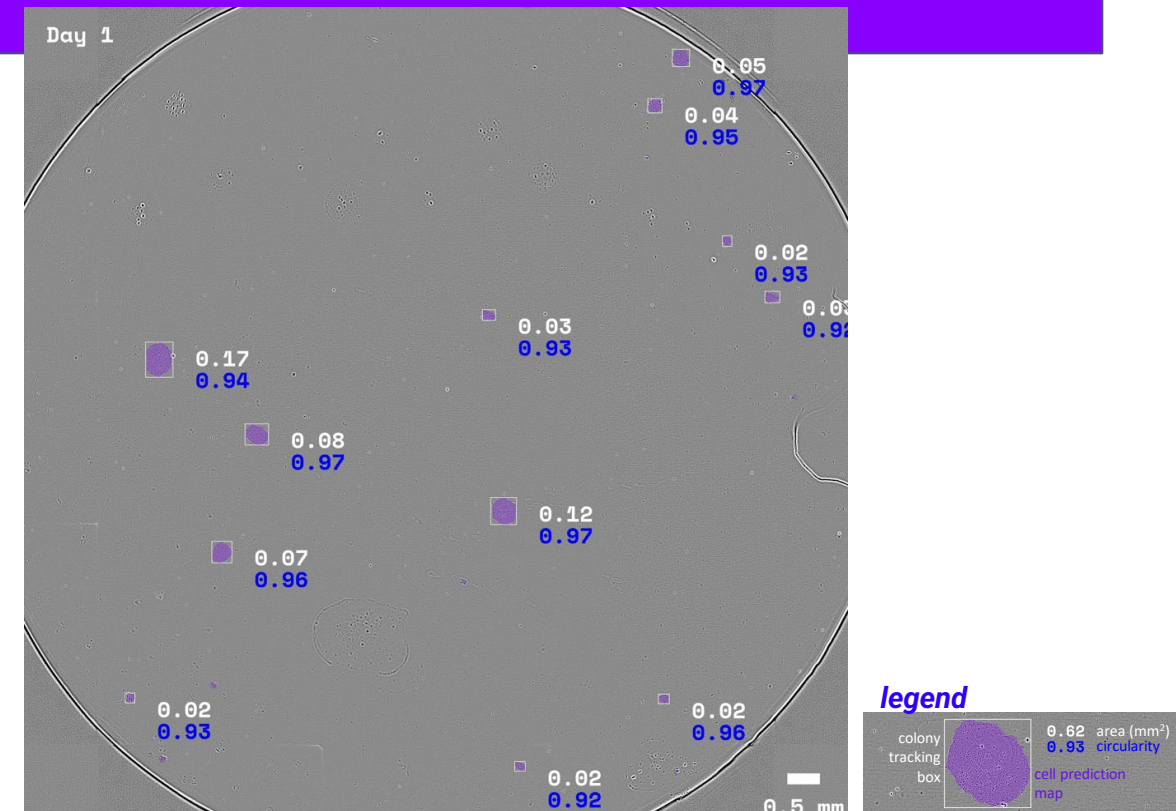
TRADITIONAL APPROACH

Manual well -to -well passaging



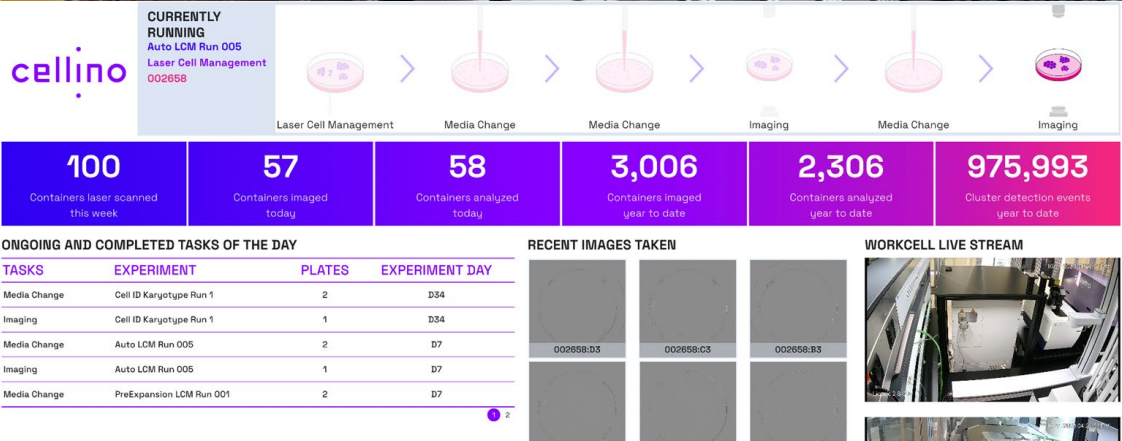
CELLINO APPROACH

Autonomous, label -free, PASSAGE-FREE bioprocess

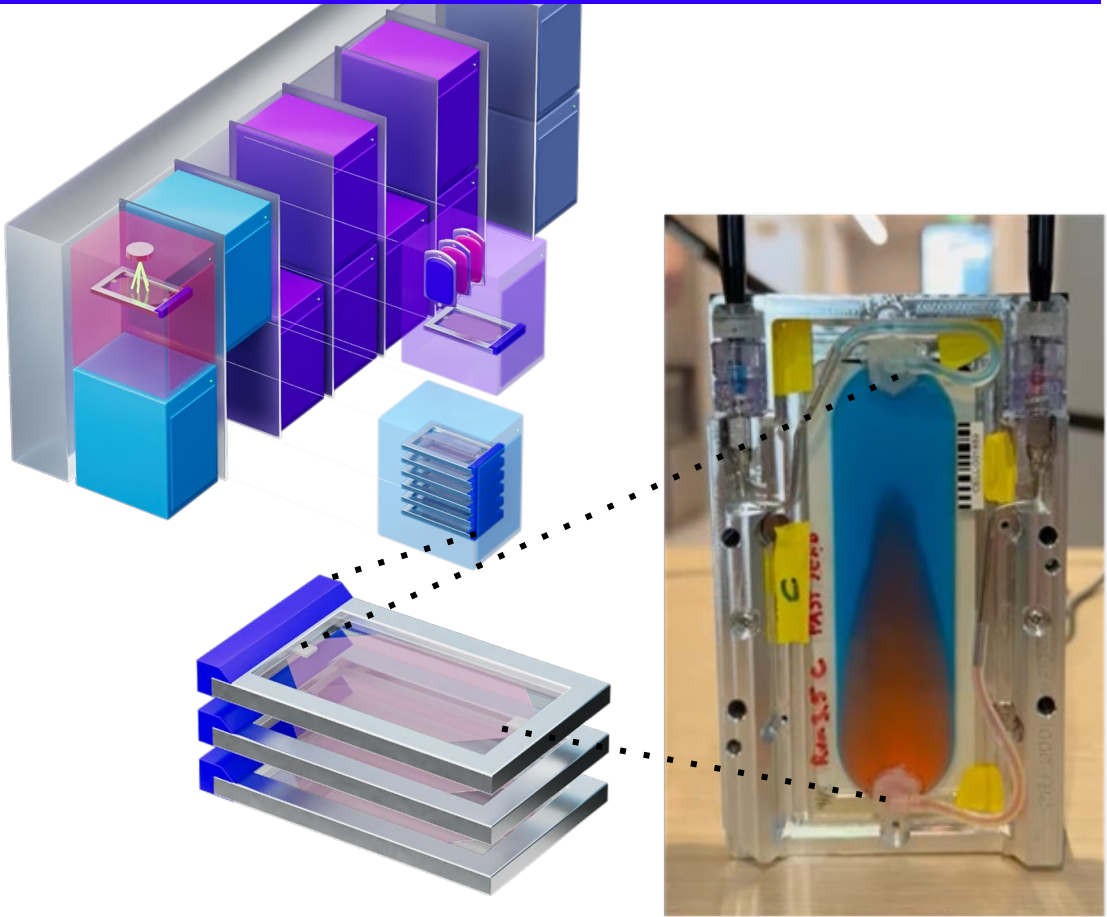


We are advancing our foundational manufacturing to support clinical programs

FOUNDATIONAL PLATFORM OPERATIONAL (Autonomous 1.0)



ULTRA-SCALABLE PLATFORM IN EARLY PROTOTYPE

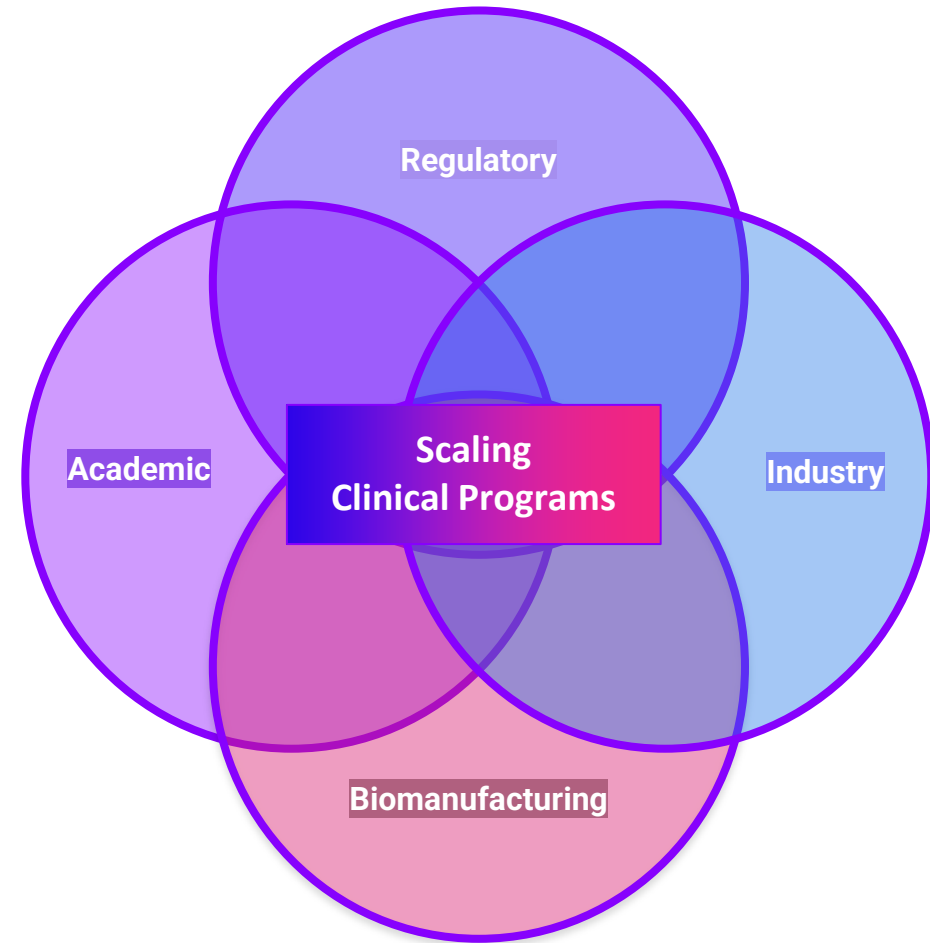




Entity	Cells Disease	IND	Ph 1/2a	Ph 2	Ph 3	Commercial
National Eye Institute NCT04339764	<i>Retinal Pigment Epithelial Cells</i> Age-related Macular Degeneration					

Robust academic/industry/regulatory/biomanufacturing collaborations needed to support rapidly advancing programs

- How do we establish a new standard approach compared to manual biomanufacturing?
- How do we define a “good” iPSC cell?
- How do we accelerate the path of standardization to meet clinical trial timelines?
- How do we implement good machine learning practices for AI-based biomanufacturing processes?



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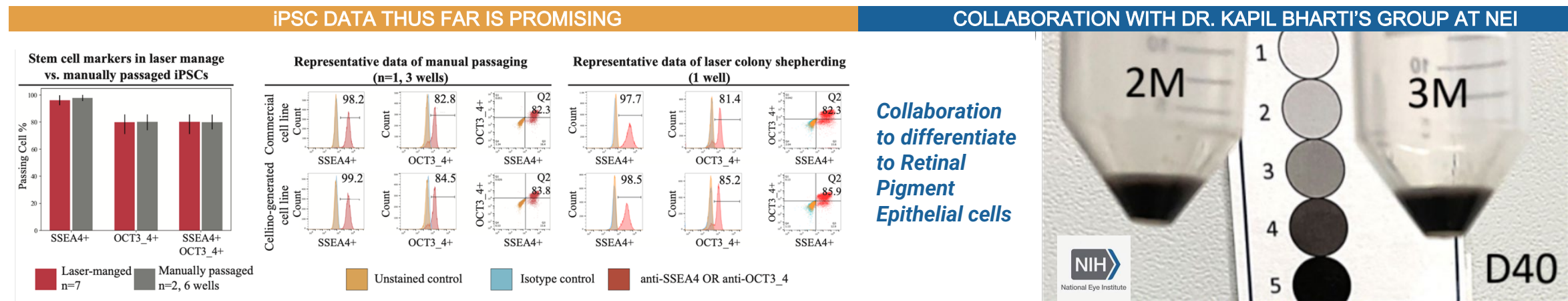
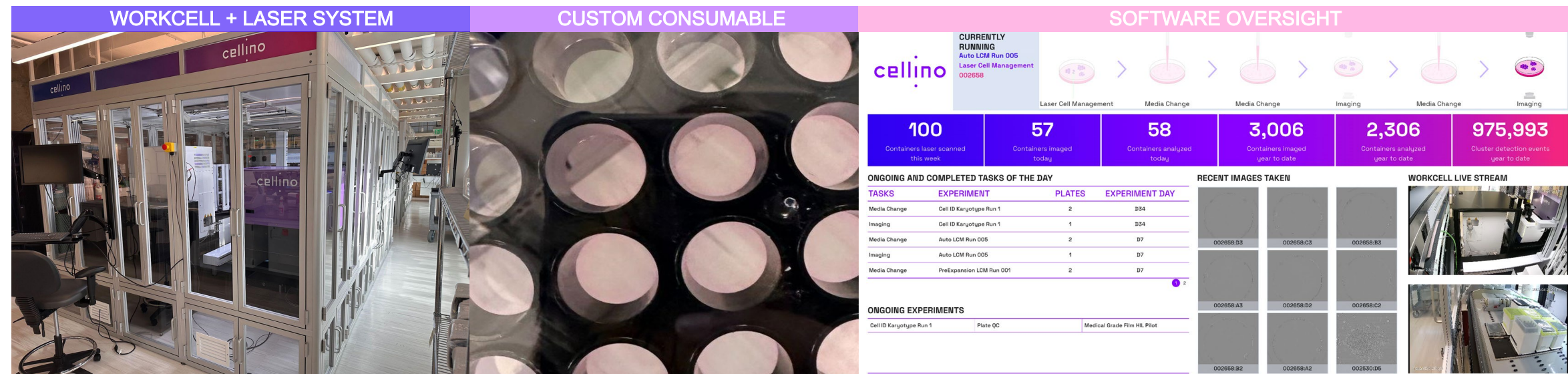
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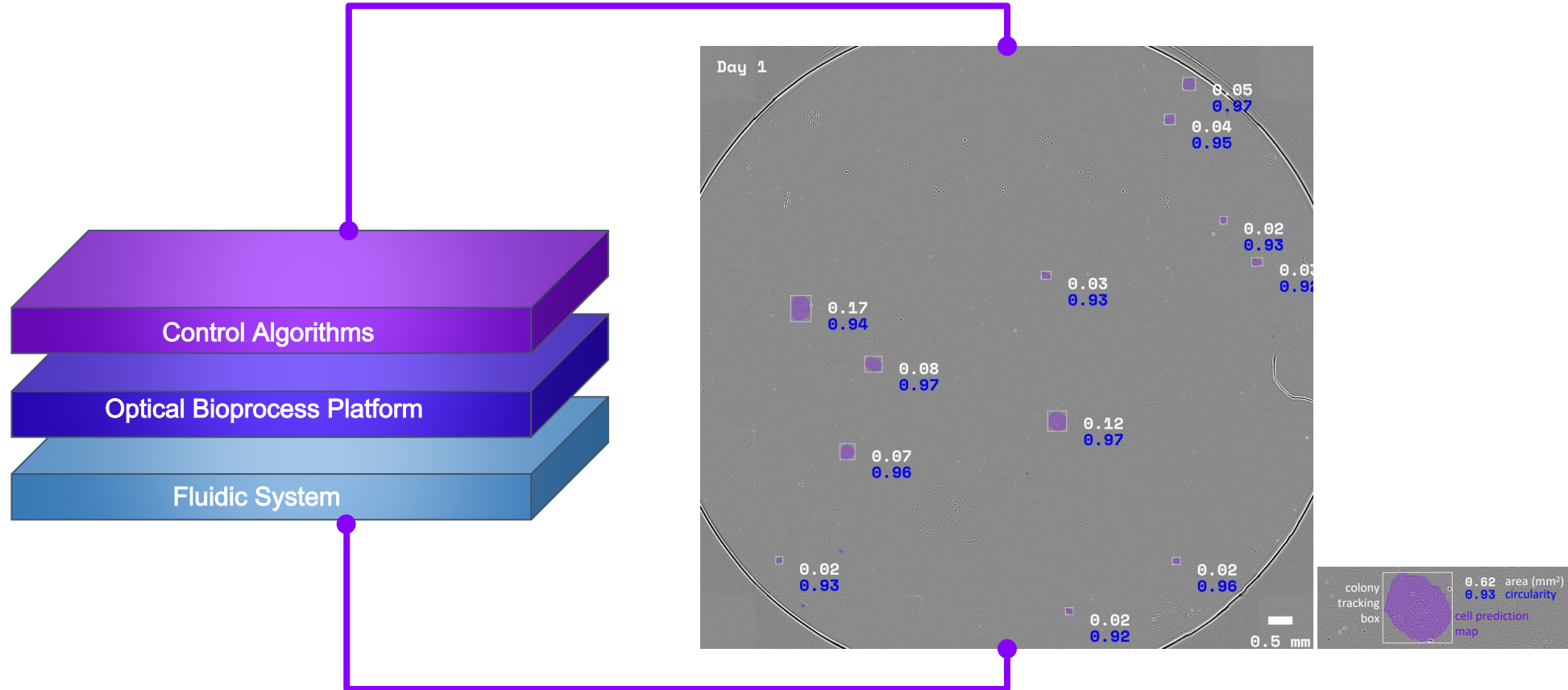
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Gen1 System Running Optical Bioprocess



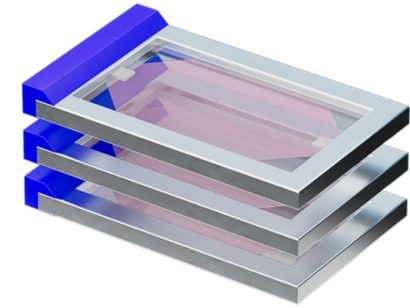
100



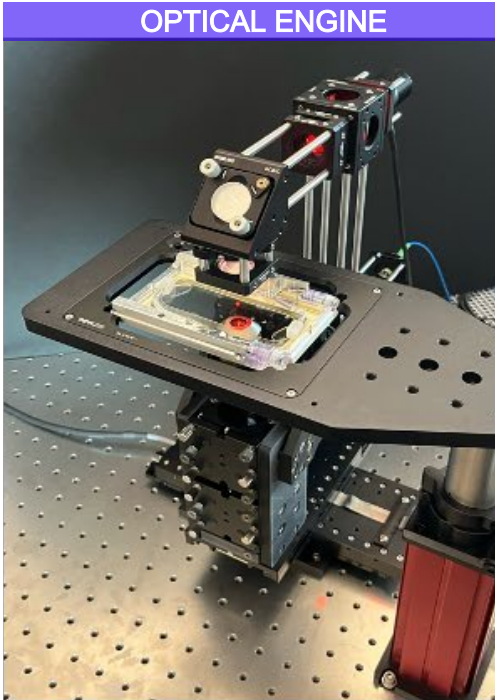
Gen2: Cassette System Development Started

Multiple Cassette Architectures for Biomanufacturing

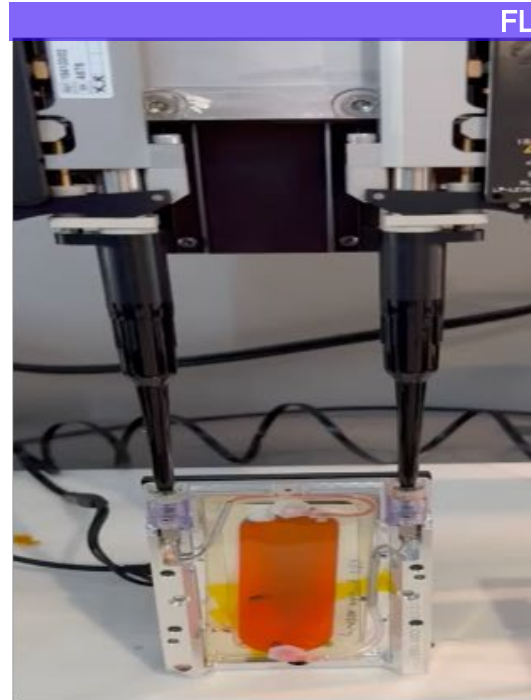
both reprogramming and differentiation capabilities



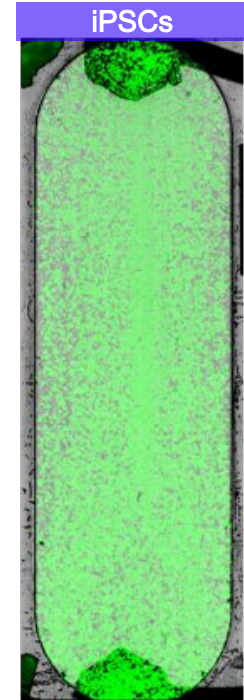
OPTICAL ENGINE



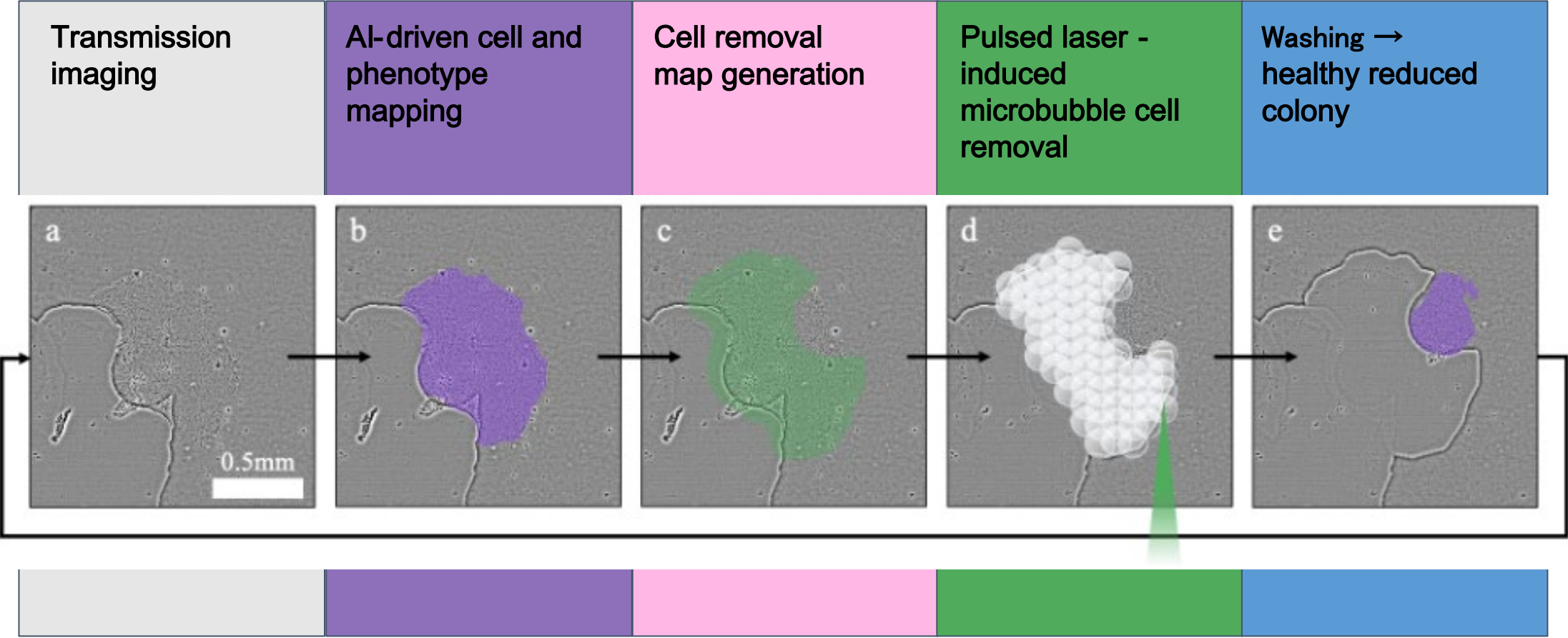
FLUIDICS



iPSCs



Cellino's Novel Optical Bioprocess For Scale



2023 BOLD GOALS FOR U.S. BIOTECHNOLOGY AND

DIAGNOSTICS

decrease health
inequities



1 address our increasingly aging,
and diversifying population

be patient-specific to minimize risk
of immune rejection

address major public health
sectors / diseases

expand
access



2 reach patients living everywhere

be POINT-OF-CARE at hospitals

decrease
cost 10-fold



4 be affordable
(not cost up to \$3M per patient)

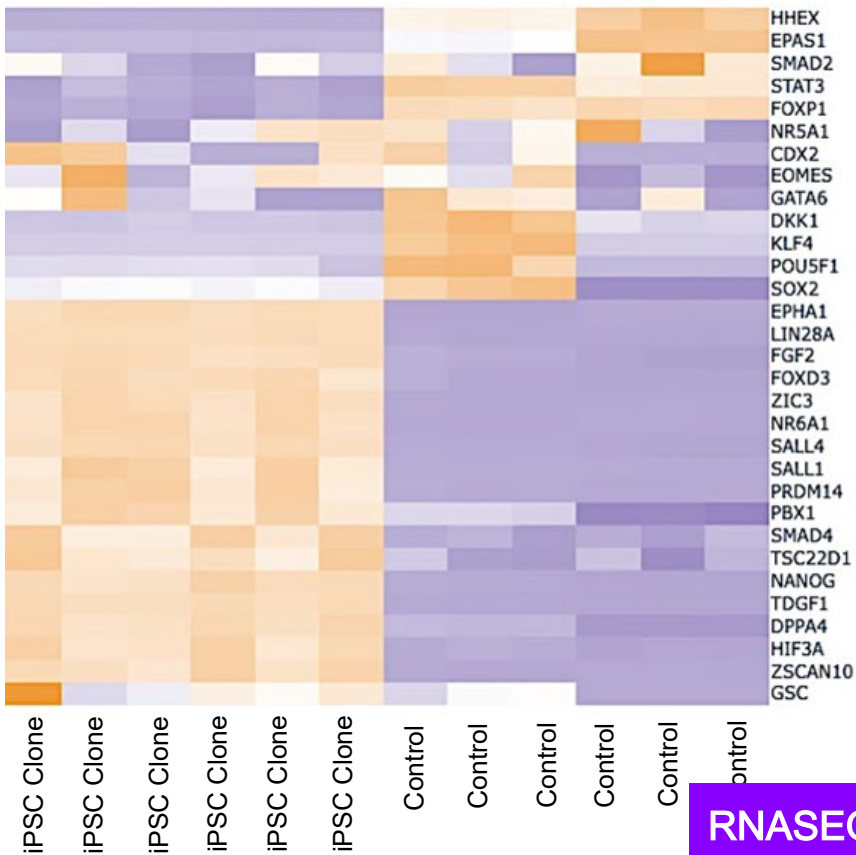
Cellino's Roadmap Towards Our Biomanufacturing Moonshot

2017 - 2020		2021	2022	2023
2017 Cellino founders met - laser processing of cells with custom consumables 2020 Decision to focus on personalized iPSC-based cell therapies	AI MODELS	- Built custom AIcloud infrastructure - Incorporated AI/ML into bioprocess	Detect cells, detect colonies, track colonies over time, calculate confluence, and automate colony selection	Calculate cell density, identify pluripotent vs non-pluripotent cells, time-series metrics such as growth/ proliferation rates
	BIOPROCESS	- Tested reprogramming techniques manually - Developed laser removal techniques (removing individual cells &entire colonies)	- iPSC101 - developed first automated Cellino iPSC process - Laser-based clonal isolation 250 days of automated culture 60 iPSC lines from 7 donors - Set up quality control metrics	- iPSC102 - developing prototype of passage-free clinical-grade process - Passage-free process - Human in loop→autonomous for 60 days - Scale quality metrics
	HARDWARE	- Custom consumable plates - Laser scanner2	- Workcell #1: Automated cell culture - Laser scanner3 &high throughput consumables	- Workcell #1: Autonomous cell culture - Workcell #2: automated QC prep - Laser scanner4 &fluidic chamber prototype
	DATA	- Health and viability of cells post-laser removal - POC iPSC colony generation from tested reprogramming techniques	Characterization of IPSC101/ automatically generated iPSCs that have undergone laser-based clonal isolation: sterility, pluripotency, functionality, genomic stability, etc.	- Further characterization of iPSCs undergoing passage-free laser-based cell maintenance process: karyotyping, pluripotency, etc - Differentiation to RPEs

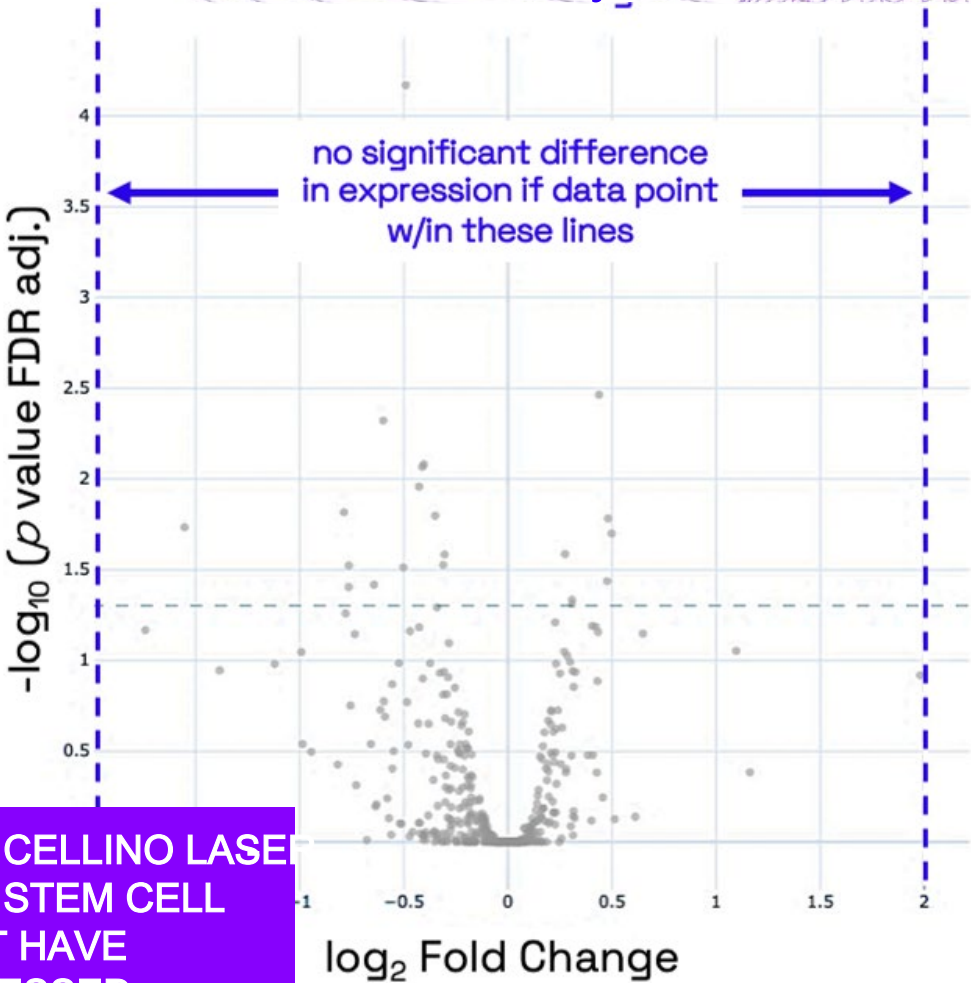
Does the laser have a negative impact on stem cell phenotype or oncogene expression of the remaining cells?

RNAseq showed laser -isolated iPSCs display stem cell signatures and do not have differentially expressed oncogenes.

Self-Renewal Gene Expression



Differential Gene Expression:
Laser-Isolated & Manually -Picked iPSCs



RNASEQ DATA SHOWS CELLINO LASE
ISOLATED IPSCS HAVE STEM CELL
SIGNATURES & DO NOT HAVE
DIFFERENTIALLY EXPRESSED
ONCOGENES

Does use of the laser have negative impacts on cell health?

A third -party vendor characterized laser -isolated iPSCs and manually -picked iPSCs and showed they were comparable for a range of assays.

Purpose

3rd-party analysis by **Elixirgen Scientific** to compare laser-isolated iPSCs to traditionally-produced iPSCs

Samples Tested

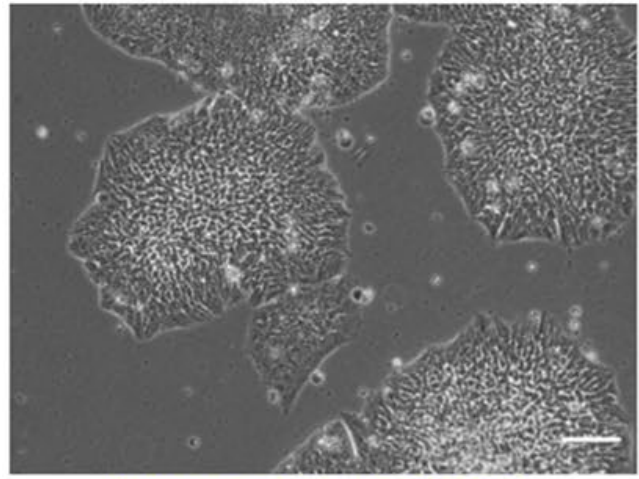
- Control lines (no laser):
 - n=1 Thermo episomal iPSC ctrl
 - n=1 CIRM episomal iPSC ctrl
 - n=3 Cellino Sendai ctrl (manually picked)
- Experimental lines (laser colony isolation):
 - n=2 Thermo episomal
 - n=3 Cellino Sendai

Analyses Performed

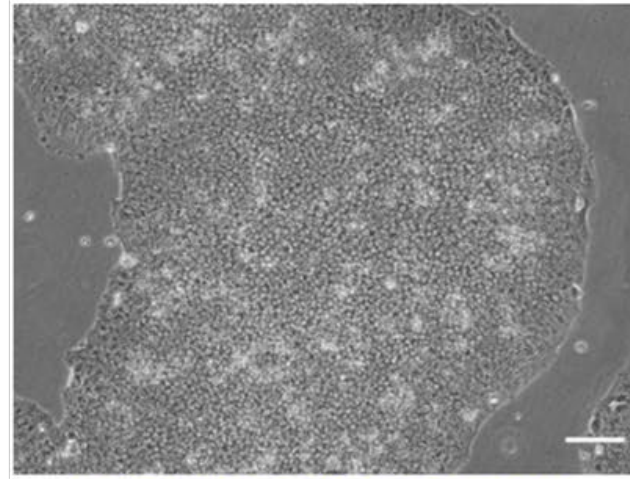
- ✓ visual inspection for morphology and (absence of) spontaneous differentiation
- ✓ p1 viability
- ✓ %Oct4+ (ICC)
- ✓ %TRA1-60, SSEA4+ (flow cytometry)
- ✓ karyotype g-banding (on a subset of samples listed above)

Does use of the laser have negative impacts on cell health?

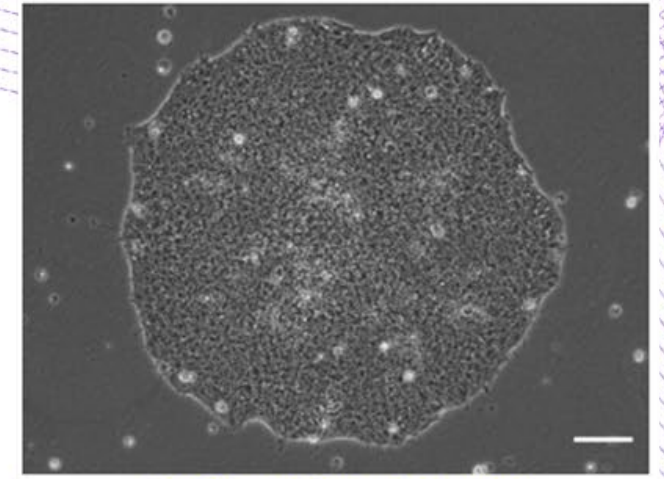
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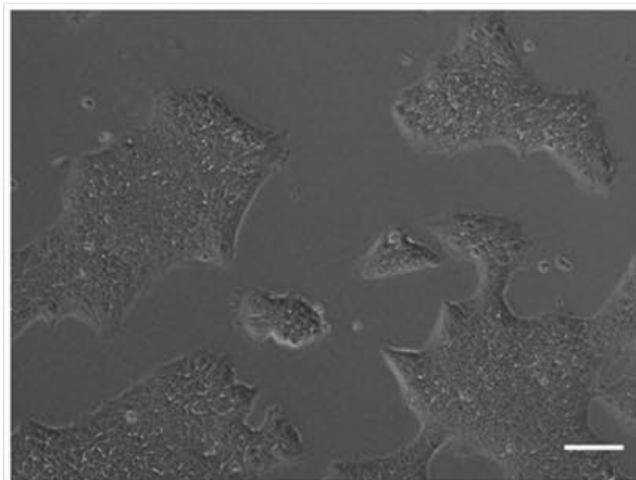
CIRM episomal ctrl



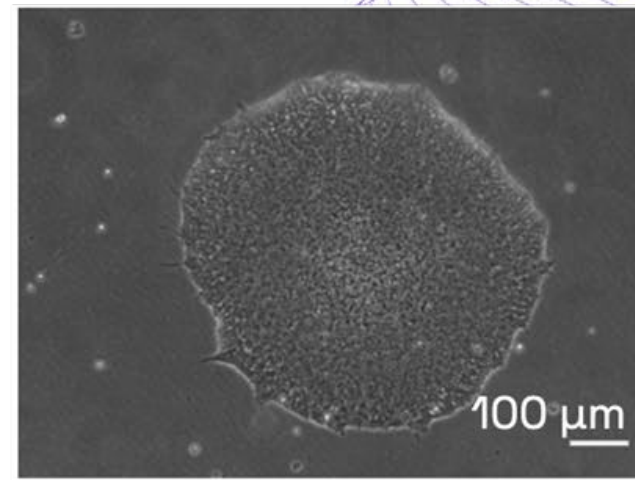
Thermo episomal ctrl



Cellino Sendai ctrl



Thermo episomal + laser



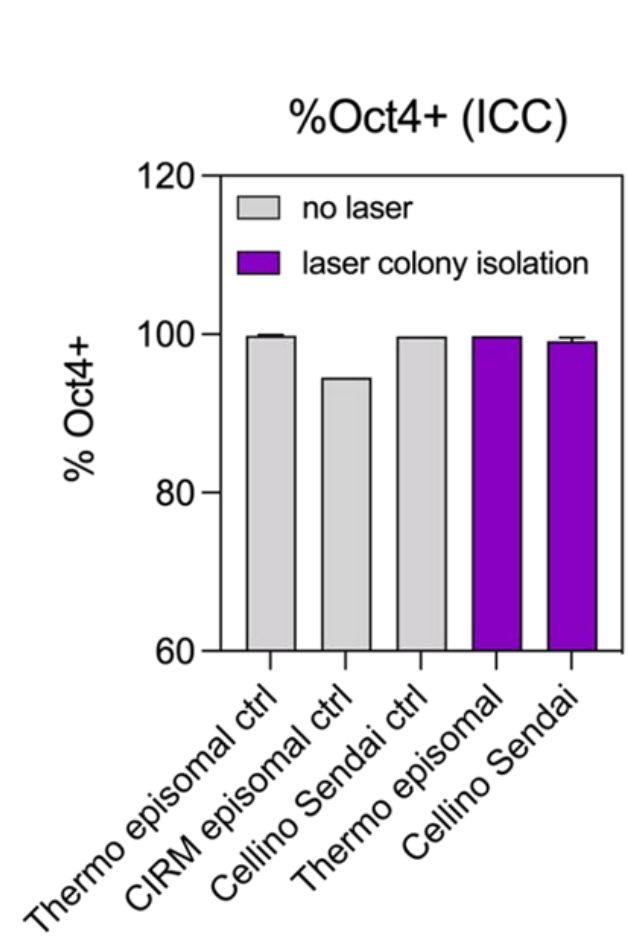
Cellino Sendai + laser

Does use of the laser have negative impacts on cell health?

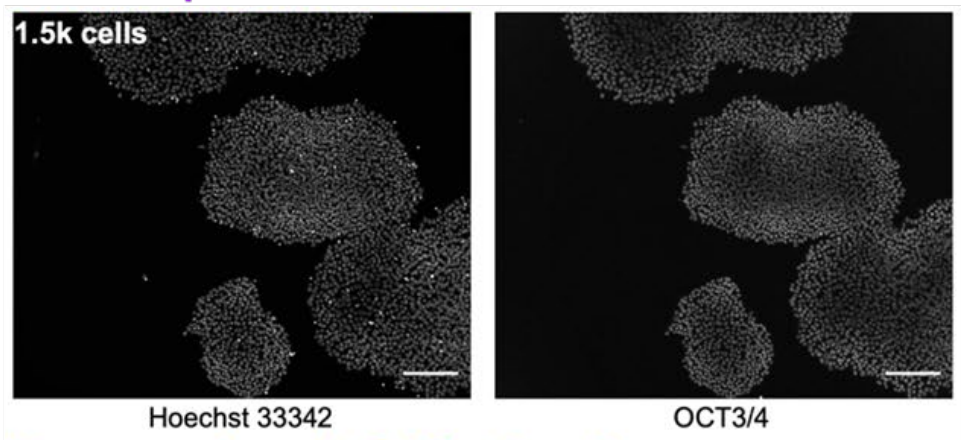
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Analysis & Conclusion

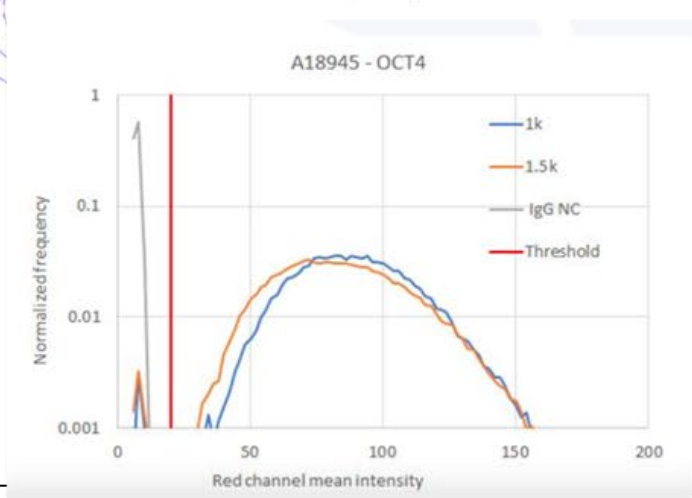
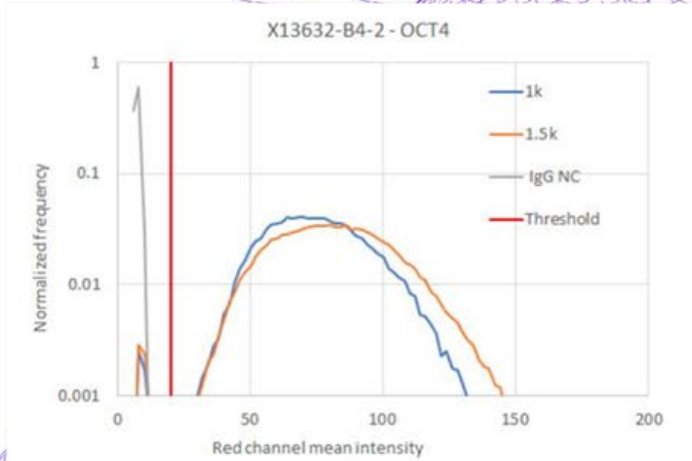
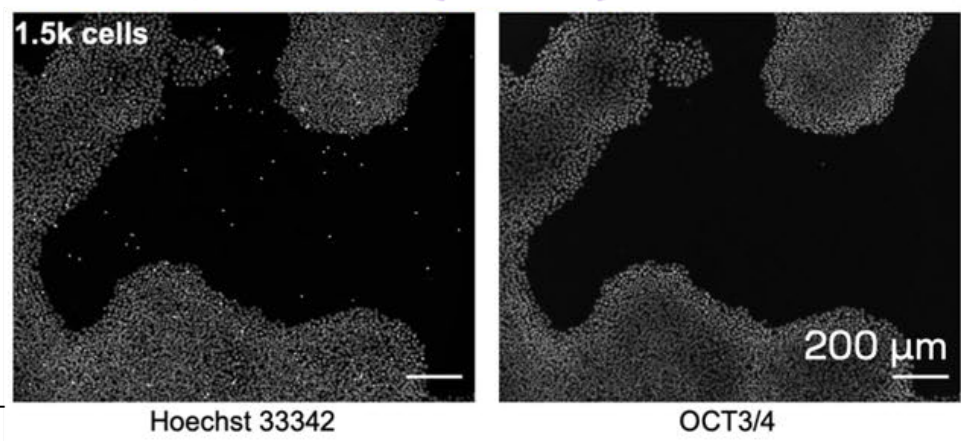
Laser-isolated iPSC lines show comparable %Oct4 expression, measured by ICC image analysis.



Thermo episomal + laser



Thermo episomal ctrl (no laser)

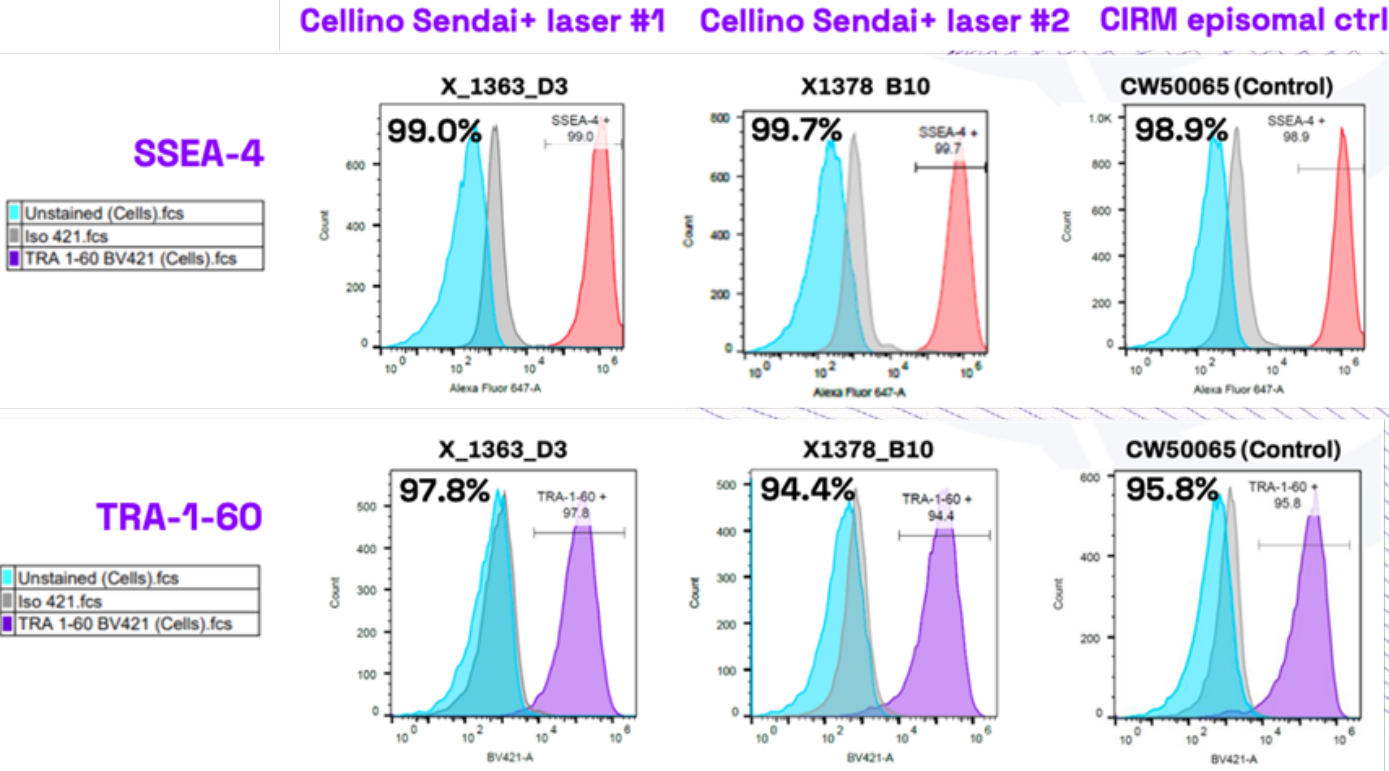
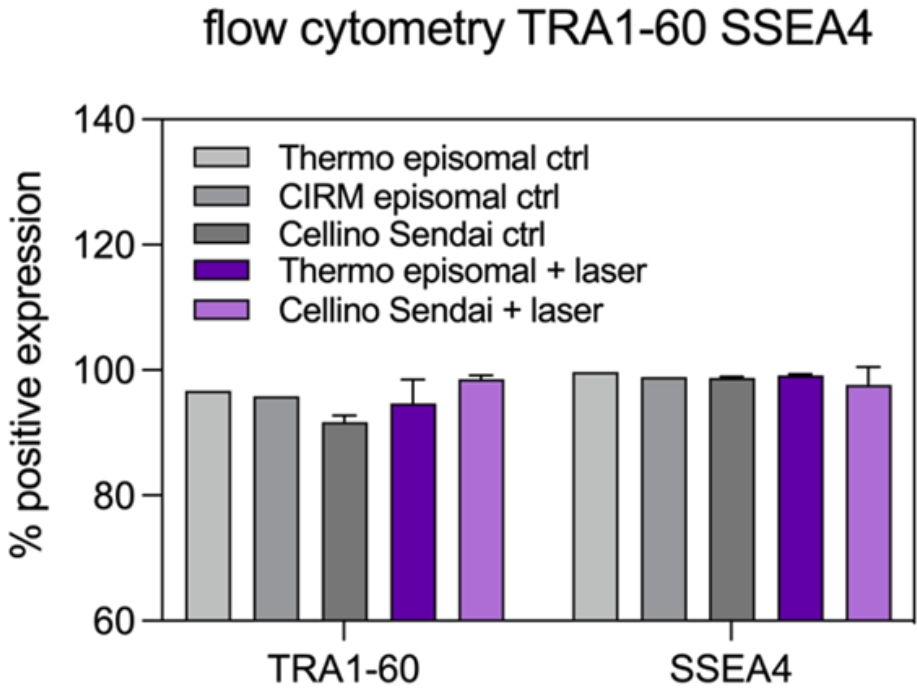


Does use of the laser have negative impacts on cell health?

A third -party vendor characterized laser -isolated iPSCs and manually -picked iPSCs and showed they were comparable for a range of assays.

Analysis & Conclusion

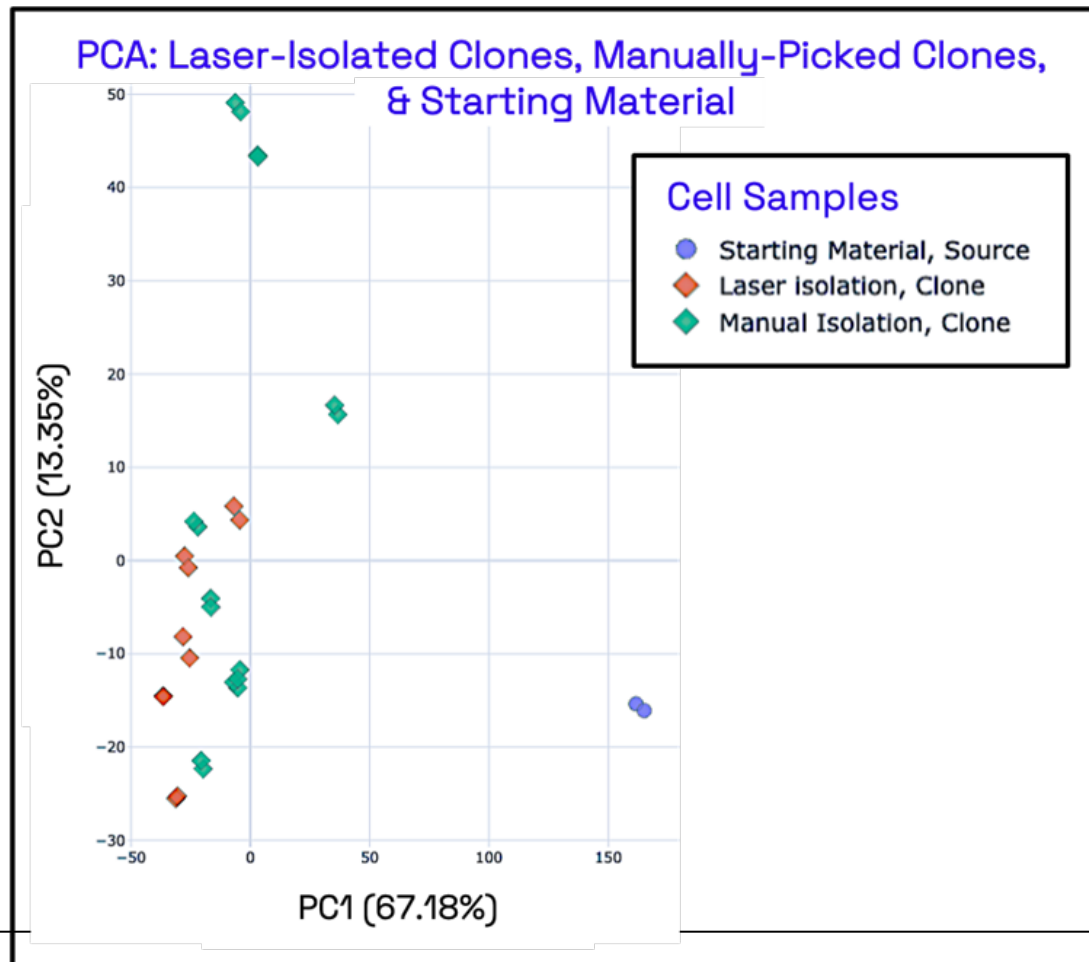
Laser-isolated iPSC lines show comparable TRA1-60 and SSEA4 expression, measured by flow cytometry.



Does use of the laser have negative impacts on cell health?

A third -party vendor characterized laser -isolated iPSCs and manually -picked iPSCs and showed they were comparable for a range of assays.

Takeaway: RNAseq analysis shows similar clustering between laser-isolated iPSC lines and manually-picked iPSC lines generated from 1 donor sample.



1 donor sample:

- CD34+ cells
- iPSCs generated w/ Sendai reprogramming:
 - laser isolated vs. manually picked

Does repeated use of the laser have negative impacts on cell health?

LATEST DATA: Laser cell management for 30 -60 days showing promising results in key assays

Increasing Autonomy

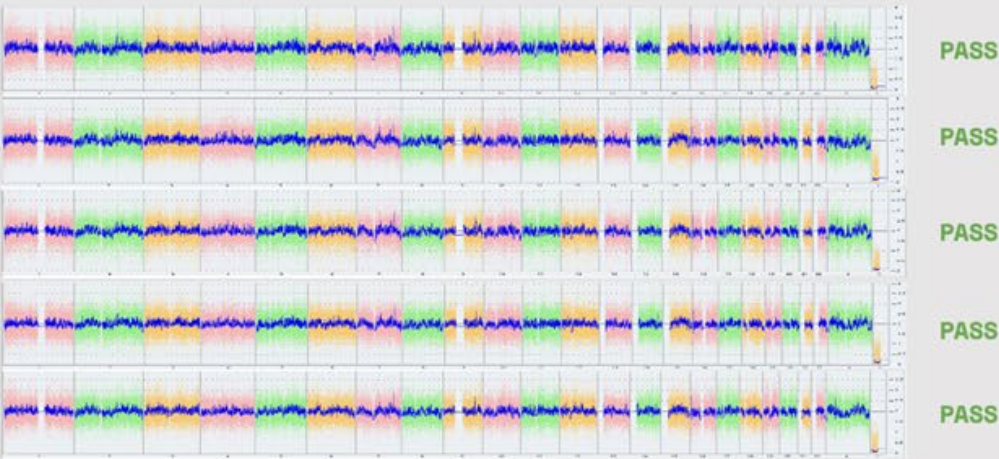
Experiments	Status	Days of LCM in single well	Wells processed	Cell lines	Results
1. Human determined laser scanning using laser scanner	✓	30, 60	4	Thermo line	Karyotyping 4/4 PASS Flow 3/4 PASS (¼ processing)
2. Human determined laser scanning using Cellino viewer	✓	30, 60	16	Thermo line Cellino line	Se V (processing)
3. Human determined laser scanning using Cellino viewer	✓	30, 60	16	Thermo line Cellino line	Karyotyping D58 7/8 PASS, 1/8 FAIL Flow D57 7/8 PASS, 1/8 FAIL WES (processing), RNA-Seq (processing)
4. Limited autonomous determined scanning, human - assisted (scan object at counterclockwise rotation) using Cellino viewer	Ongoing	30, 60 (Ongoing)	96	Thermo line Cellino line	Karyotyping (ongoing) Flow (ongoing) Se V (ongoing)
5. Fully autonomous determined laser scanning (scan frequency and directionality)	Future	60	TBD (> 96)	Thermo line Multiple cellino lines	Karyotyping Flow, Se V, WES, RNA-Seq

Does repeated use of the laser have negative impacts on cell health?

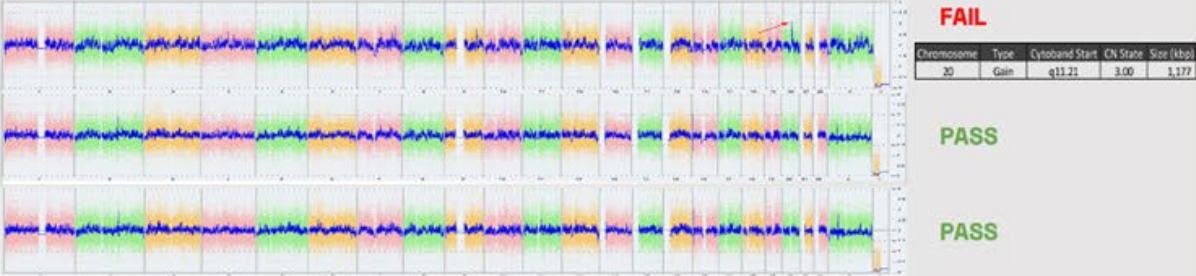
LATEST DATA: Laser cell management for 30 - 60 days showing promising results in key assays

D58 LCM - No chromosomal aberrations in 7 / 8 wells

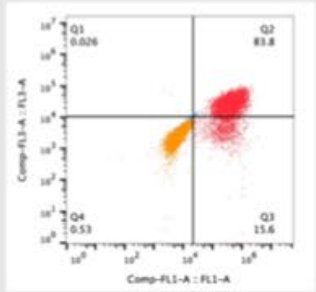
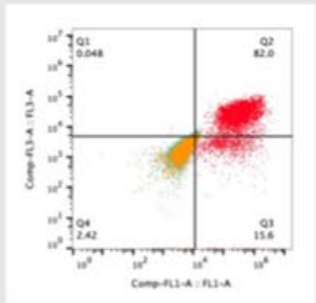
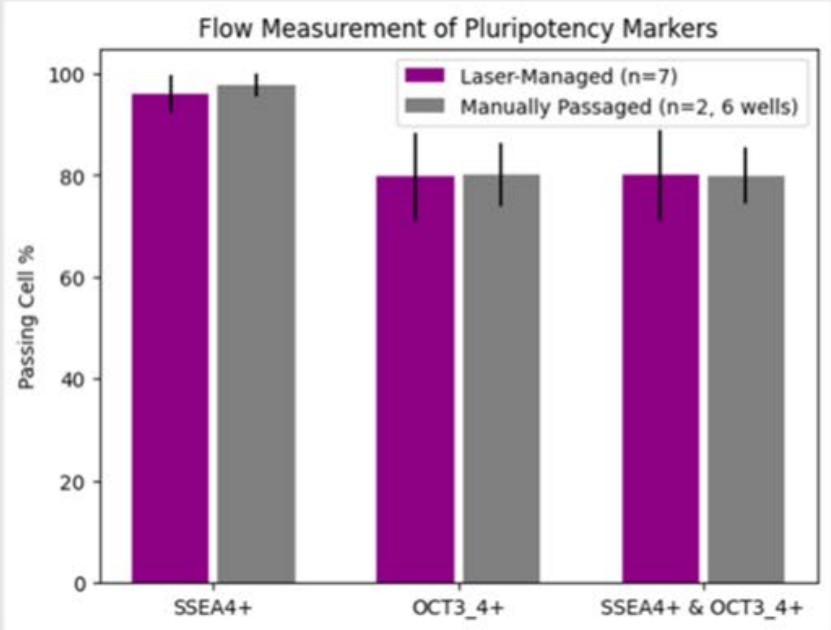
Thermo line A18945



Cellino line



D57 LCM - Equivalent OCT4 and SSEA4 expression to manually passaged wells (Thermo & Cellino lines, n=7)



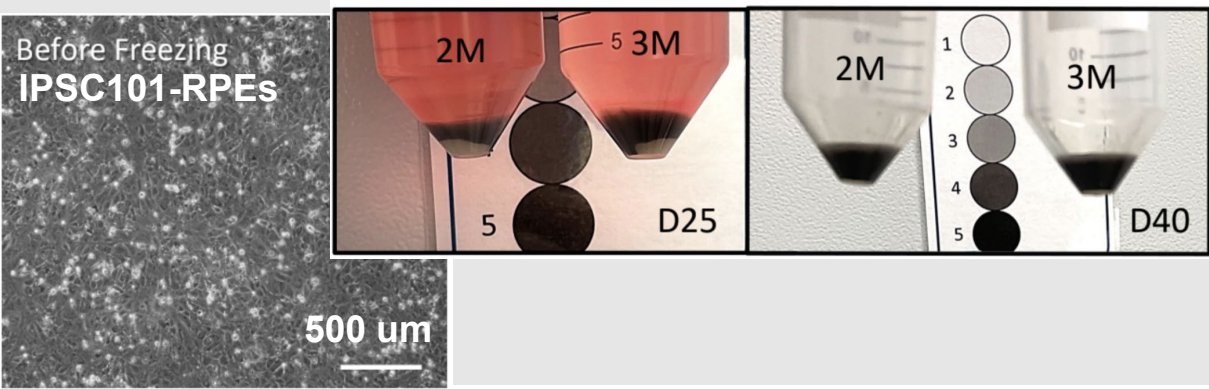
3. What is new about your approach? Why do you think you can be successful at this time?

We collected extensive data on iPSCs generated with Cellino bioprocess (clonal selection)

2022 IPSC LINE GENERATION

60 clonal iPSC lines generated & cryopreserved
7 donors: 2 African-American, 1 Latino, 4 Caucasian

COLLABORATION WITH NEI (DR. KAPIL BHARTI)



Quality Control by FACS, TEER and POS

Day-2	Day25			Day 40				
	Pigm.	TYRP1, %	MITF1, %	Pigm.	TYRP1, %	PMEL17, %	CRALBP, %	MACS, %
2M	4	57.75	93.85	5	98.95		78.75	65.8
3M	4	69.50	97.10	5	97.75	94.75		64.4

MODEL TO AUTOMATE COLONY SELECTION

86% of time, model’s #1 choice matched SME
trained on 3284 images of colonies

AUTOMATION STATS

250 days automated operations
520 wells underwent laser colony isolation
Imaging capacity: 3456 96-wells/day
laser colony isolation capacity: 576 96-wells/day

MANUFACTURING READINESS

3 QC vendor relationships established
1 expansion partner relationship established
9 internal QC assays developed
2 blood cell vendor relationships established

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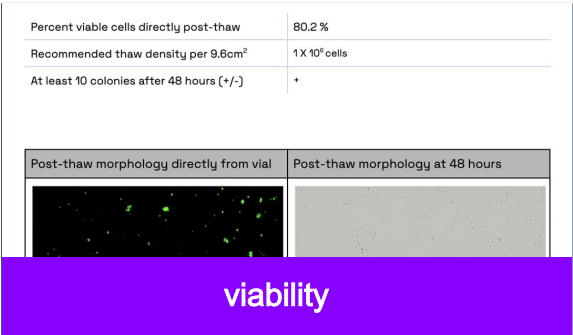
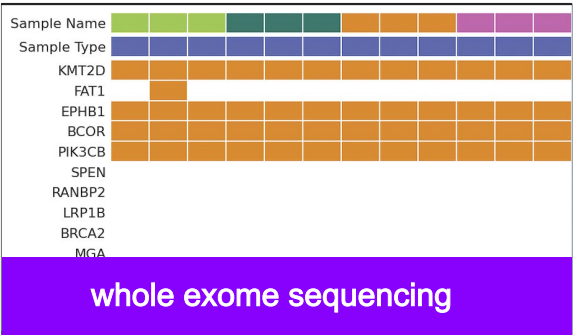
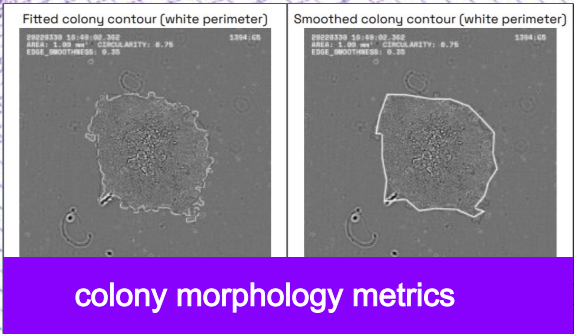
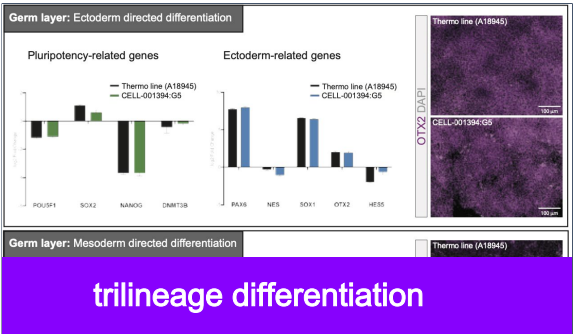
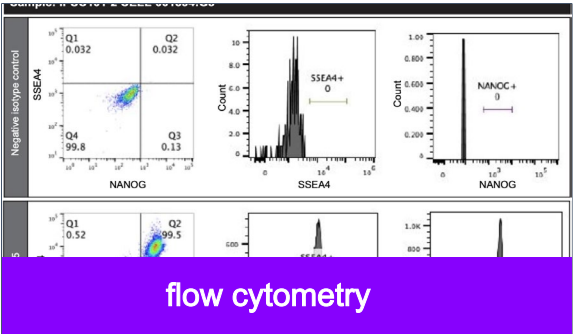
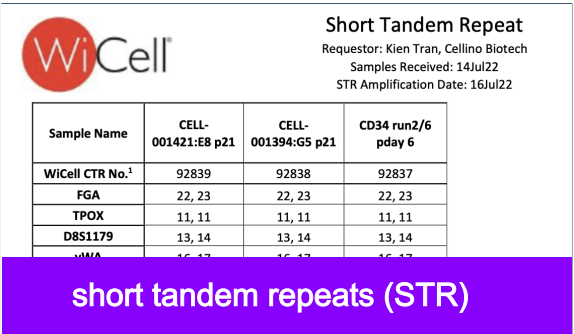
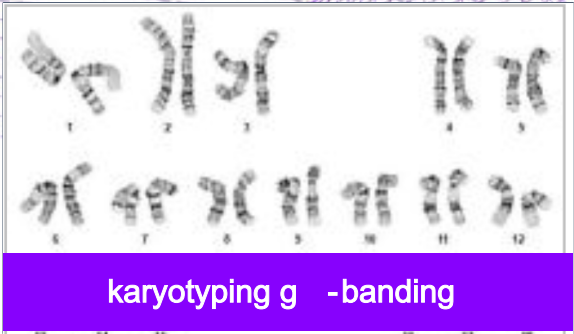
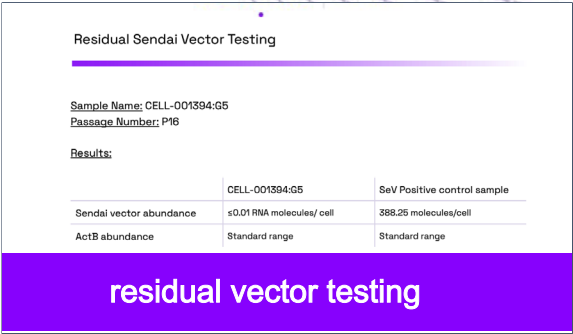
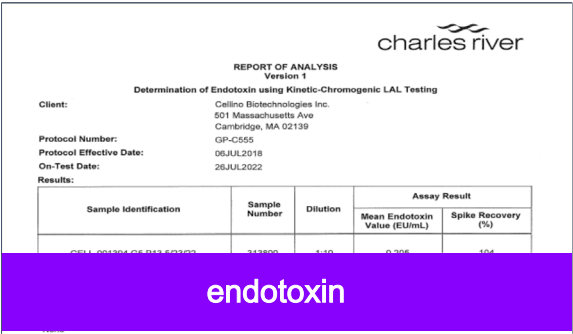
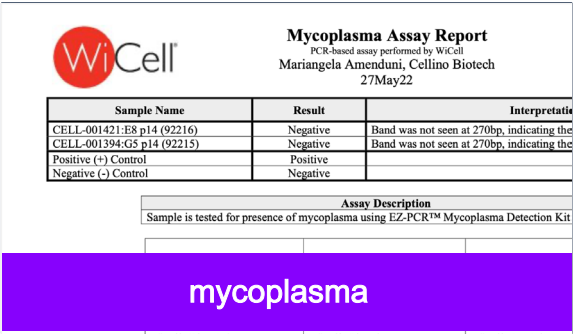
	ATTRIBUTE	TEST	METHOD	TESTED MATERIAL	RELEASE CRITERIA
RELEASE CRITERIA	Safety	Mycoplasma	WiCell (SOP-83.01)	Medium pre-cryopreservation	negative
		Bacterial & Fungal sterility	USP<71> and <61>, Ph.Eur.2.6.27 and 2.6.1, JP17<4.05> and <4.06>	Medium post-thaw	negative
		Endotoxin	CRL (GP-C555)	Medium post-thaw	negative
		Residual Sendai vector testing	qPCR	Cells pre-cryopreservation	< 0.01 copies/ cell
		Chromosome analysis	G Band karyotyping	Cells pre-cryopreservation	Identical to patient/ donor karyotype
		Oncogene integrity	WES, RNA-Sequencing	Cells pre-cryopreservation	Identical to patient/ donor oncogene sequence
	Identity	Patient/ donor identity	STR	Source material & iPSCs post-thaw	identical to patient/ donor
		Cell identity	Flow cytometry	Cells post-thaw	>90% SSEA4 NANOG +/-
	Cell Health	Viability	Dye exclusion test	Cells post-thaw	>60% post-thaw viability

	ATTRIBUTE	TEST	METHOD	TESTED MATERIAL
FOR INFO. ONLY	Colony information	Time-series colony growth tracking	Time-series image collection of colony growth pre-laser isolation	Cells in-process during reprogramming
		Single timepoint colony morphology	Image-based characterization to assess circularity, area, & edge smoothness, at time of laser colony isolation	Cells in-process during reprogramming
	Potency	Pluripotency	Directed differentiation towards endoderm, mesoderm and ectoderm (trilineage)	Cells post-thaw
	Expression profiling	Transcriptome analysis	RNA-Sequencing	Cells pre-cyropreservation

CELLINO only

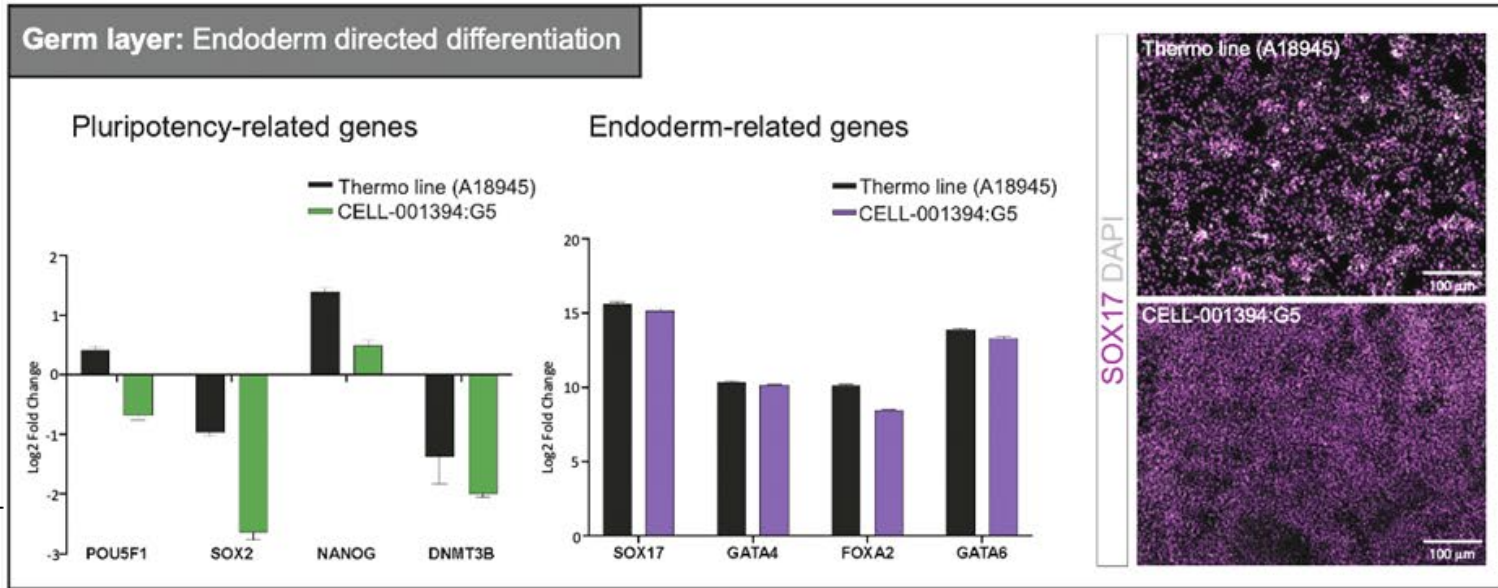
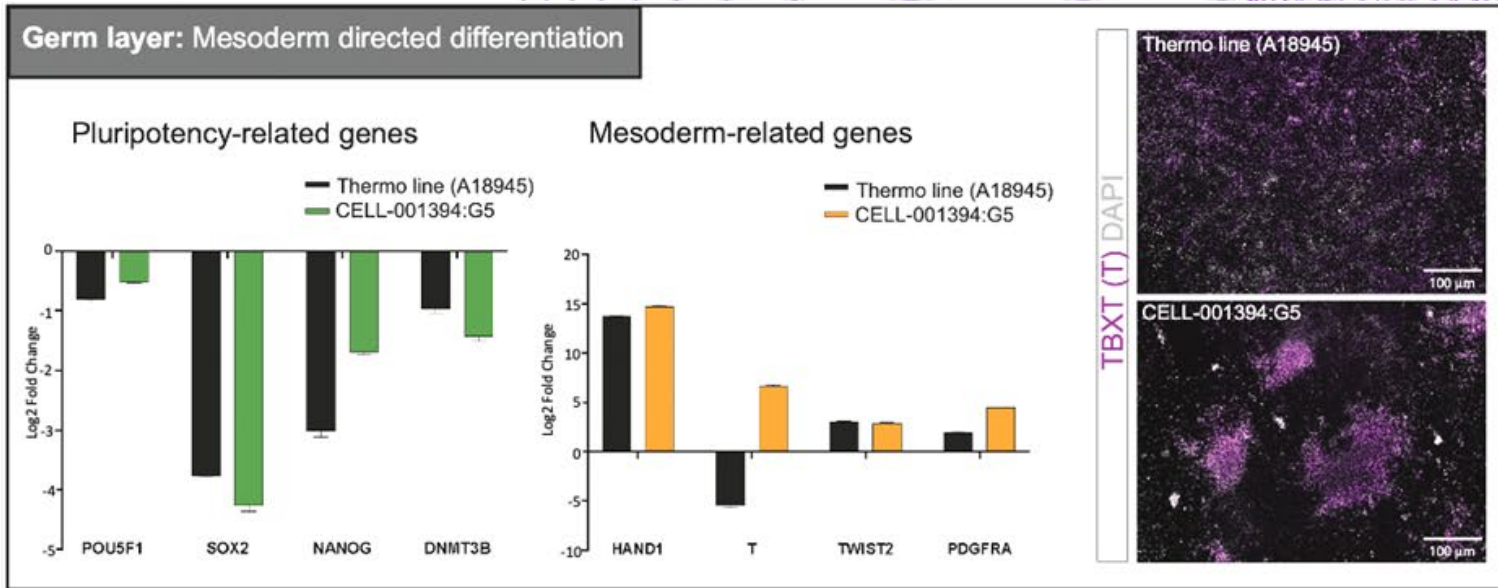
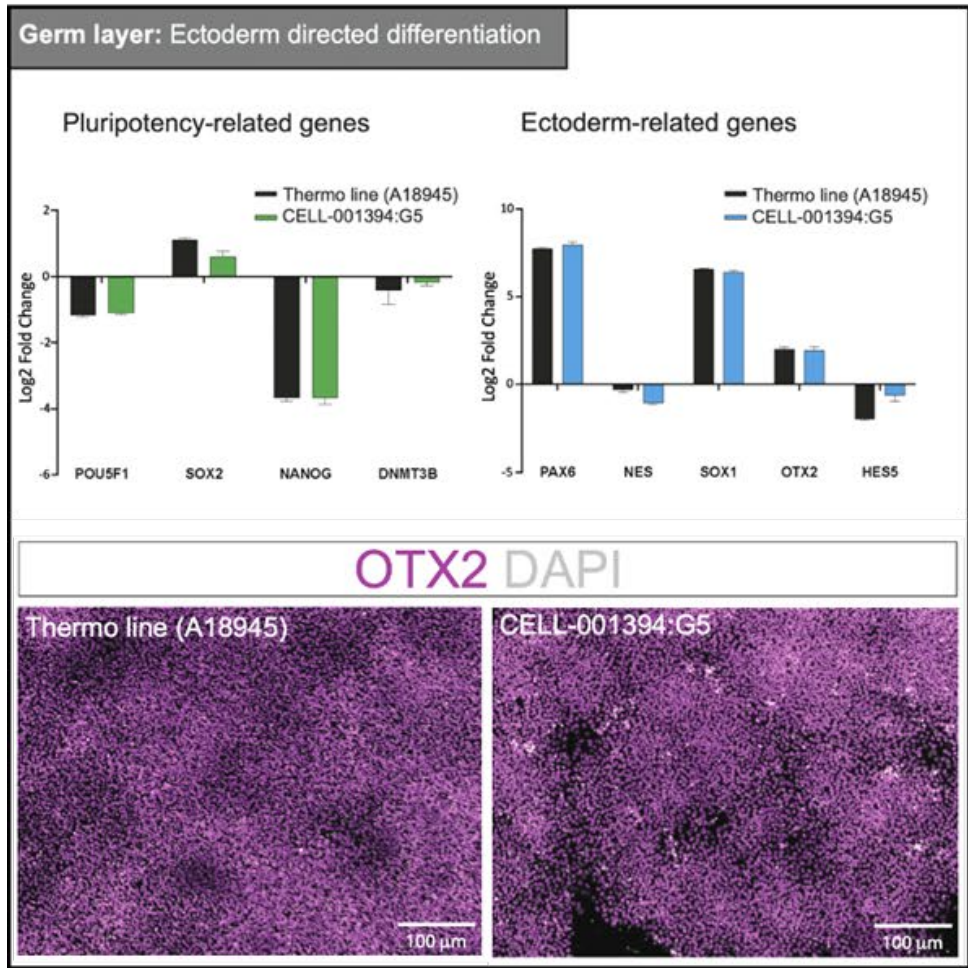
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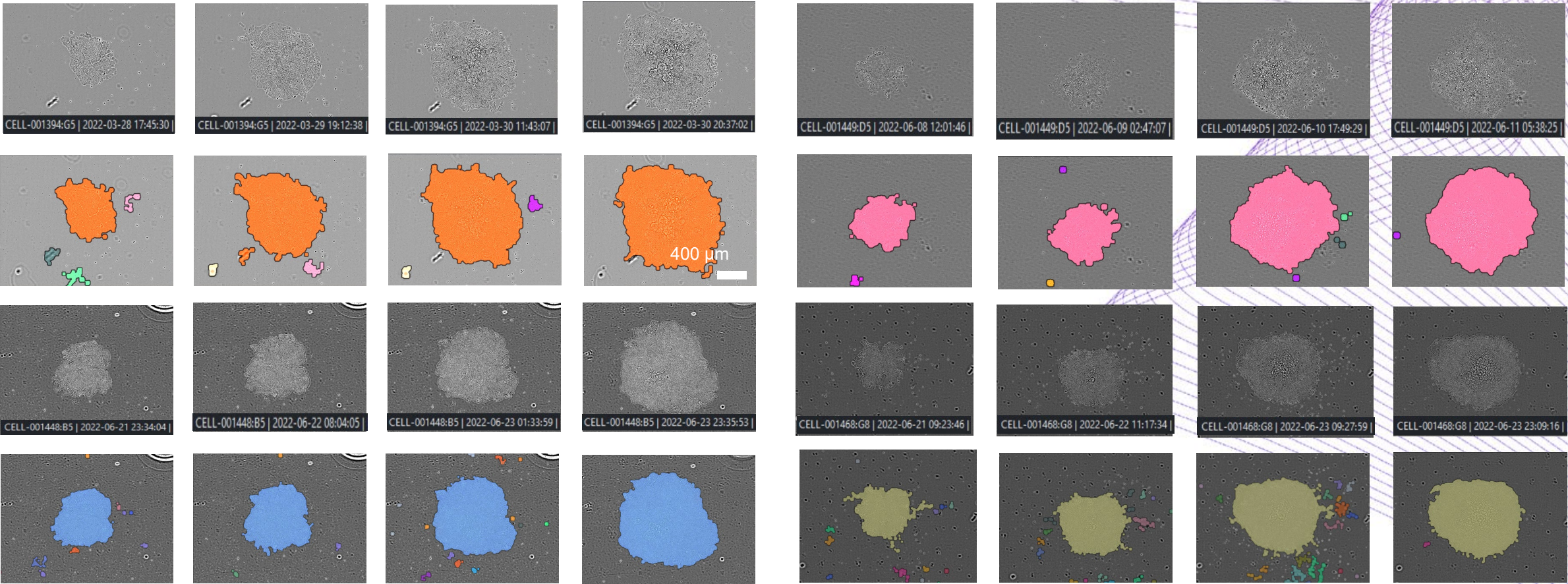
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3. What is new about your approach? Why do you think you can be successful at this time?

AI infrastructure in the cloud to track cells and run processes autonomously

Time-series tracking images of 4 iPSC colonies from automated reprogramming process



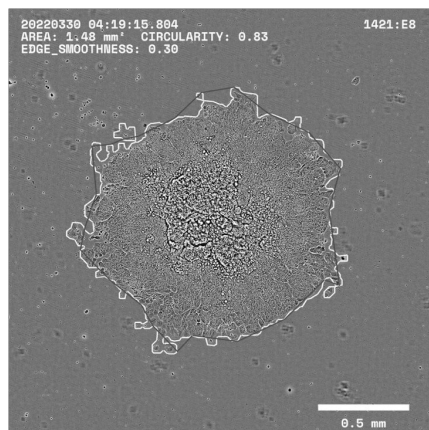
How did you develop the algorithm to predict colony selection?

We developed a cluster ranking algorithm to autonomize the selection of iPSC colonies (typically done by an SME)

What we did: We build an initial model that can, based on images of a colony, predict whether a colony would have been selected by a subject matter expert (SME) for downstream processing. **How we did this:**

1 Collect image training data

- 112 96-wells
- 3284 colonies total; 57 colonies selected by SMEs
- metrics: area, edge smoothness, density, circularity, perimeter, position w/in well, etc.



2 Determine correlation between image-able features and whether a colony is selected by an SME

$$r_{pb} = \frac{(\bar{y}_1 - \bar{y}_2) \cdot \sqrt{pq}}{s_y}$$

Point-biserial correlation

Used to correlate continuous features (eg area) w/ binary outcome (eg selected or not)

4 Test model on images of colonies

Testing: tested on a different batch run, w/ different donor material:

86% of time, model's #1 choice matched SME

3 Build probability model to predict, based on imageable features, which colony is likely to be selected by an SME

$$P(\text{selected}|\text{features}) = \frac{P(\text{features}|\text{selected})P(\text{selected})}{P(\text{features})}$$

Bayes rule

Used to calculate probability of colony being selected based on morphological features

Cellino ML Models - Dataset Scales

Model name	4X Confluence Model	4X Density Model	4X Pluripotency Model	Cluster Ranking Model
Signal used for extraction of "Ground Truth"	Calcein AM Live Stain	Genetically encoded nuclear fluorescence (EGFP tag on endogenous HIST1H2BJ protein)	Calcein Red-Orange AM Live Stain + Alkaline Phosphatase (AP) Live Stain	Colony object predictions
Contents of 1 datapoint	1 4X BRT image + 1 4X Confluence Map	1 4X BRT image + 1 4X Density Map (Derived from Nuc Segmentation)	1 4X BRT image + 1 4X Pluripotent cell map	Colony features (density, edge smoothness) + SME-based selection labels
Number of available raw (full well) datapoints	59	137	75	171
Number of (full well) datapoints used for training current version	44	27	N/A (training in progress)	112
Number of cell lines covered in dataset	3	1	3	2
Names of cell lines used	A18945 (ThermoFisher Episomal line); AICS61 (Allen Institute Sendai Line w/ Nuc. EGFP); PBMC-derived iPSC Line (Cellino In-House, new)	AICS61 (Allen Institute Sendai Line w/ Nuc. EGFP)	A18945 (ThermoFisher Episomal line); AICS61 (Allen Institute Sendai Line w/ Nuc. EGFP); PBMC-derived iPSC Line (Cellino In-House)	2X CD34+ Cell-derived iPSC clones (Cellino In-House)

Cellino ML Models - Performance Metrics

Model Name	4X Confluence Model		4X Cell Density Model (In Progress)		Colony Ranking Model	
Evaluation Metric	Precision	Recall	Pearson's r	Classification Accuracy	Mean Reciprocal Rank	% Match to Expert Selection
Explanation	Fraction of true positives over predicted positives (i.e. how good is the model in only detecting true cells?)	Fraction of true positives over all positives (i.e. how good is the model in detecting all true cells?)	Strength of linear relationship between true and predicted local cell density (between -1 to 1)	Fraction of correct region classifications as empty, normal density, or high density	Average of reciprocal of predicted rank of SME-selected colony (1 for perfect model, lower for worse performance)	Percentage of cases where model also selects SME-selected colony
Average value for test dataset	%98.98	%95.24	0.9-0.95	70-87%	0.9	%83.3