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Genome Editing In the Clinic

Fyodor D. Urnov, PhD

Project Leader and Senior Scientist,
Sangamo BioSciences, Inc.

NAS International Summit on Human Gene Editing
December 3, 2015



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Genome editing has expanded the definition of the term
“druggable target”:

If it's in the DNA, it's a druggable target

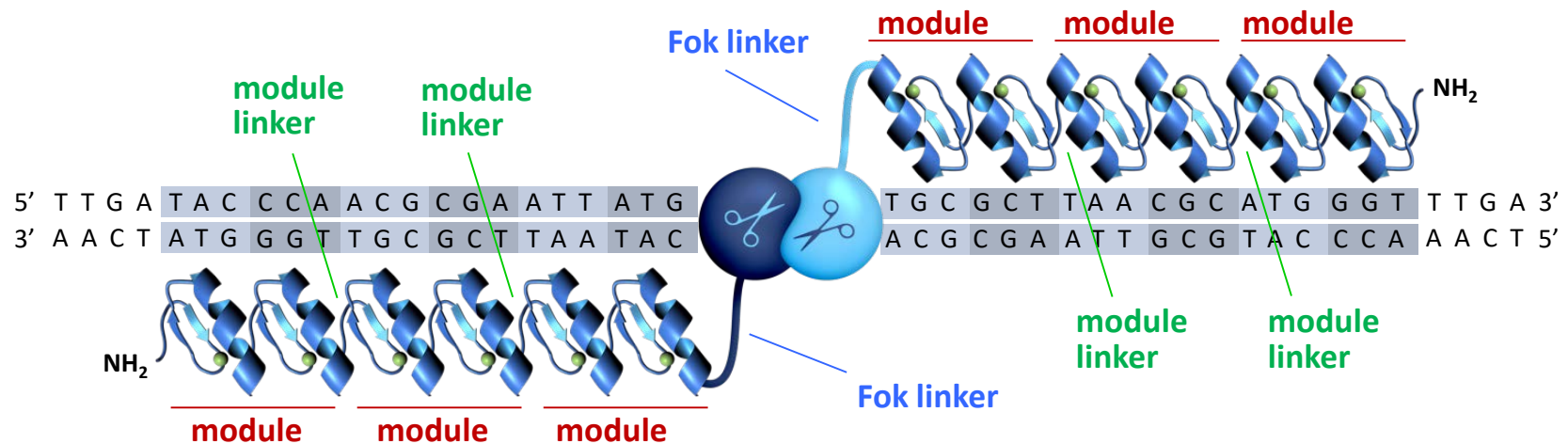
Bedside to Bench to Bedside

The use of somatic cell genome editing to create a disease-protective genetic state:

1. (T cells and HSPCs) CCR5 editing → HIV
2. (HSPCs) BCL11A enhancer editing → β -thalassemia / SCD
3. (Liver) *In vivo* targeted gene addition → monogenic disease

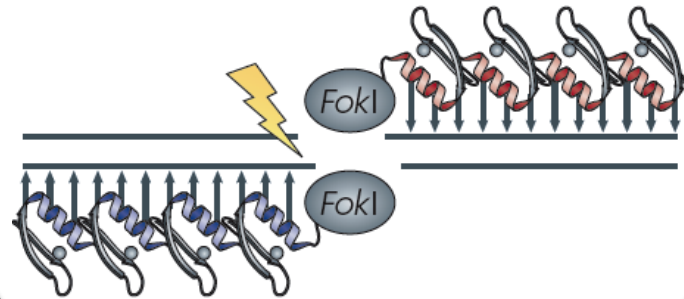
Pharmaceutical Drug Development Applied to Genome Editing

- Generate therapeutic reagents maximized for potency / specificity (the “medicinal chemistry” of genome editing)



- Using established regulatory frameworks to characterize the product appropriately for safety:
 - Tailored to the nature of the technology
 - Appropriate for the intended product and indication (risk-benefit)
 - *In vitro* (using multiple distinct assays / readouts)
 - *In vivo* (using appropriate models)

Human Genome Editing With Engineered ZFNs: Targeted Disruption



HDR

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Gene correction



Targeted gene addition

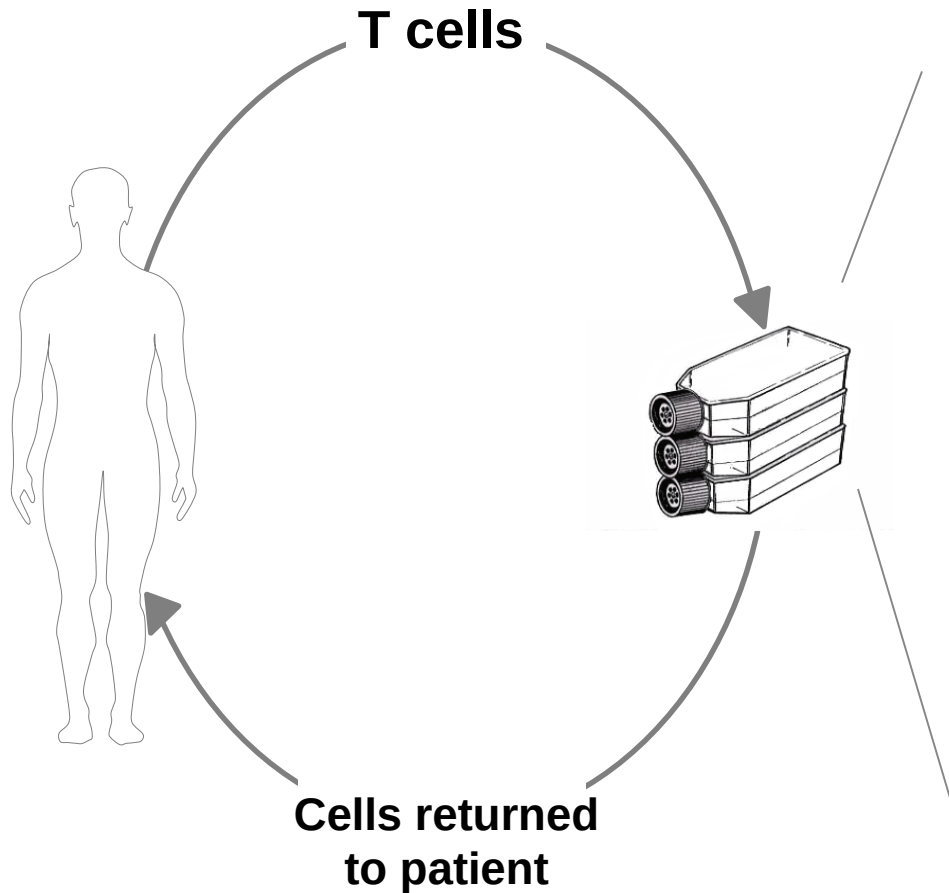


Transgene stacking

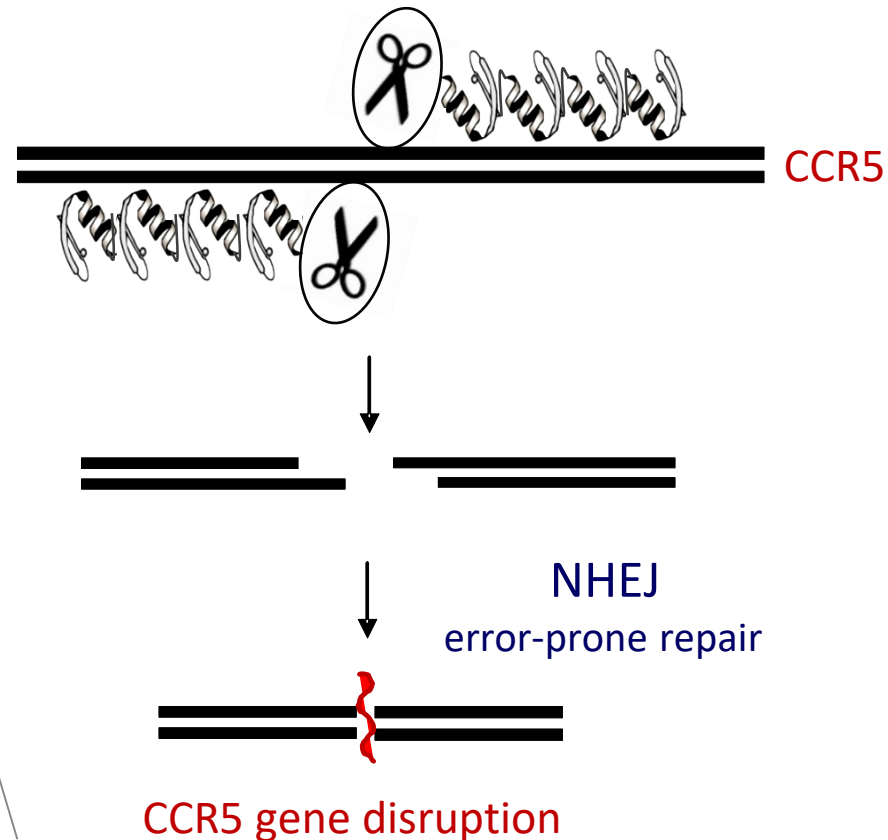
Homozygous Defect in HIV-1 Coreceptor Accounts for Resistance of Some Multiply-Exposed Individuals to HIV-1 Infection

Rong Liu,* William A. Paxton,* Sunny Choe,* Daniel Ceradini,* Scott R. Martin,* Richard Horuk,† Marcy E. MacDonald,‡ Heidi Stuhlmann,§ Richard A. Koup,* and Nathaniel R. Landau*

designated EU2 and EU3, required about 1000-fold more virus to establish infection than control cells from unexposed donors. While a small fraction of the cells did become infected with this high inoculum, the virus failed



Genome Editing for HIV

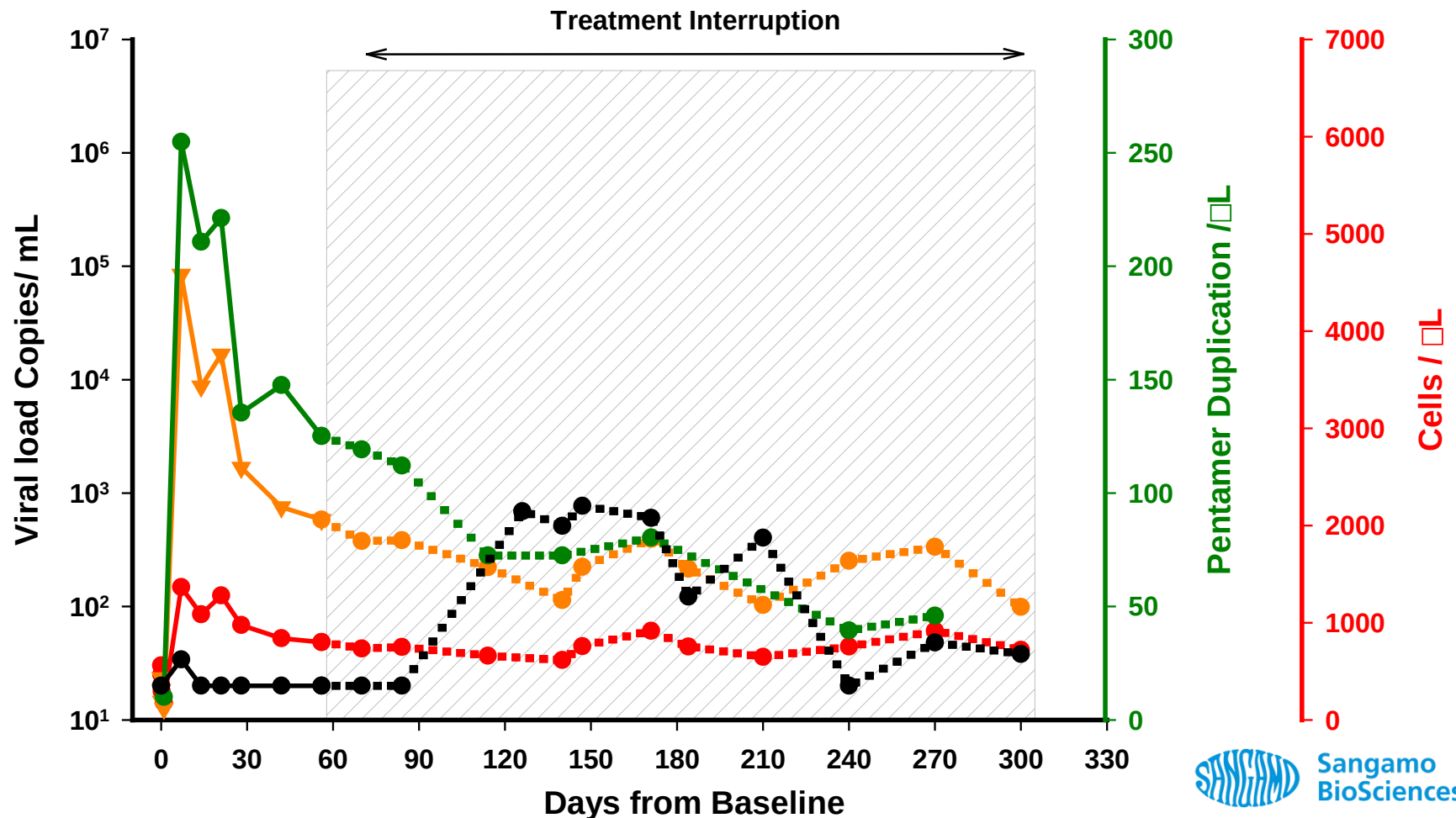


Preclinical efficacy: Perez et al Nature Biotechnology 2008
Manufacturing: Maier et al Human Gene Therapy 2013
Clinical data from first cohort: Tebas et al NEJM 2014

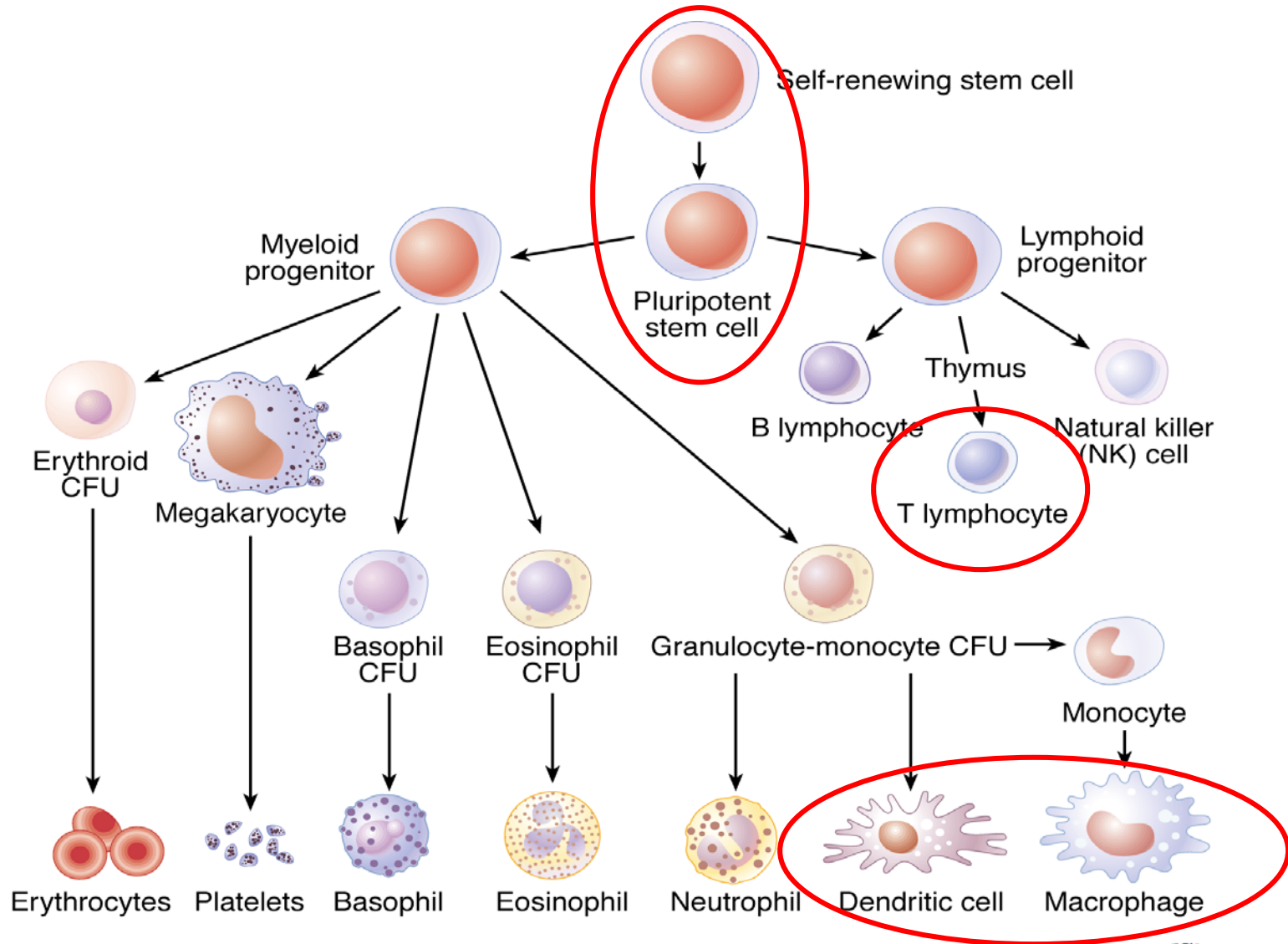
In-clinic Efficacy Readouts: Genome Modification, CD4/8 Count, Viral Load

Subject 04-046

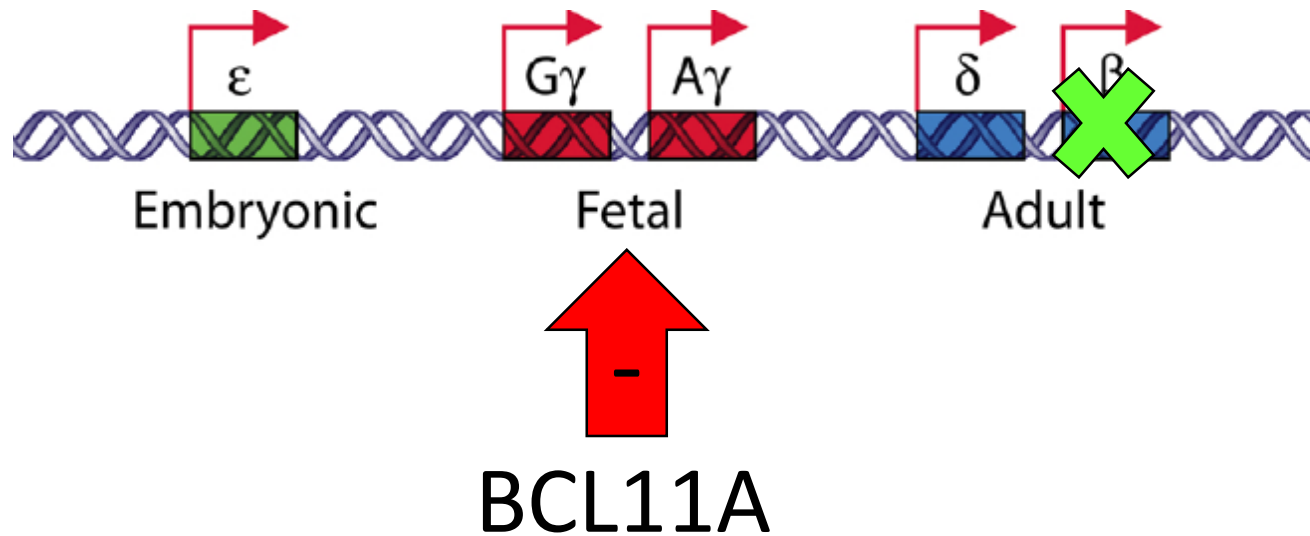
- Absolute CD4 Count
- ▼ Absolute CD8 Count
- Pentamer
- Viral Load



Editing Hematopoietic Stem Cells Endows All Their Progeny With the Desired Genomic Signature



Disease-Protective Genetic Variation in β -thalassemia



2008:

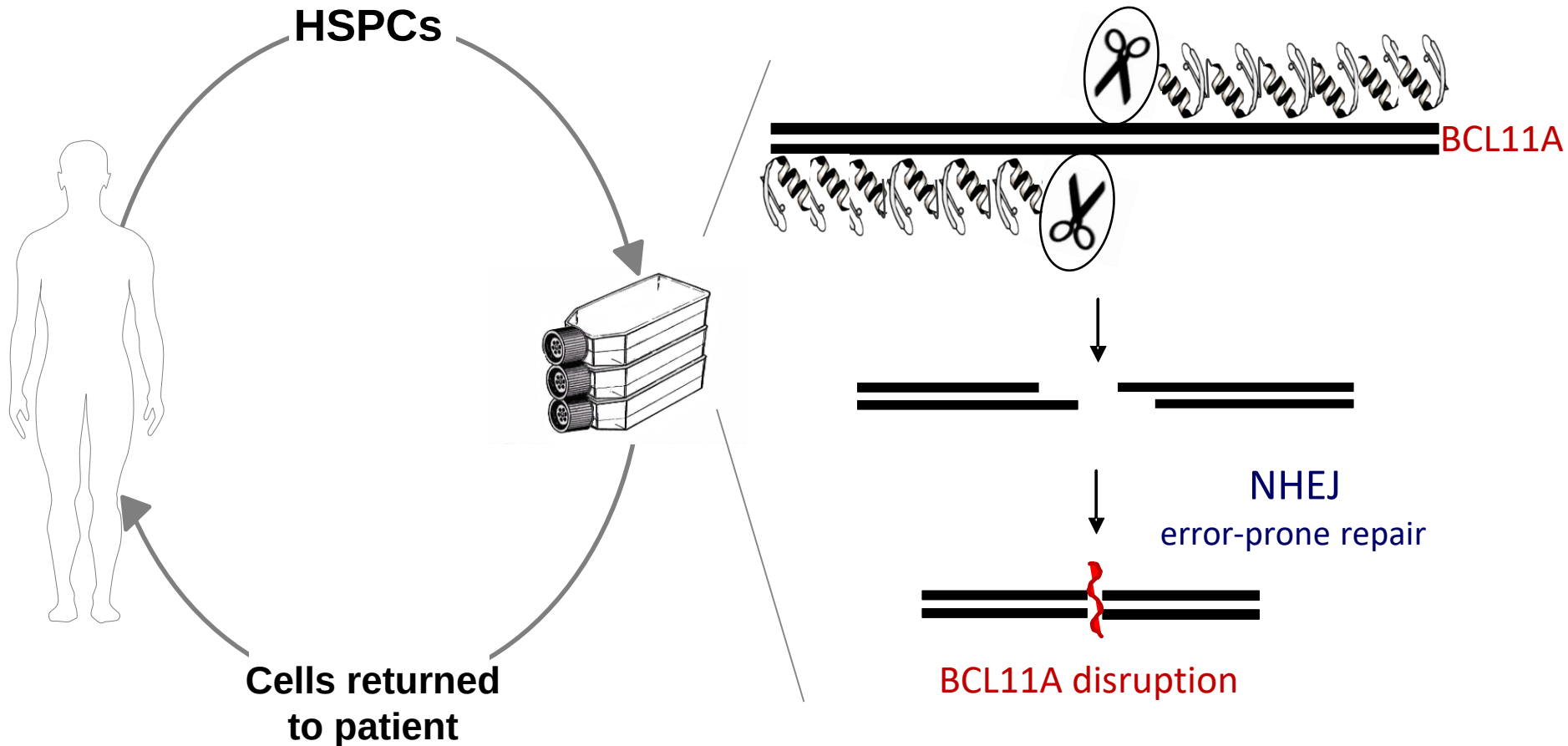
PNAS

Genome-wide association study shows *BCL11A* associated with persistent fetal hemoglobin and amelioration of the phenotype of β -thalassemia

Manuela Uda*, Renzo Galanello†, Serena Sanna*, Guillaume Lettre*§, Vijay G. Sankaran*¶, Weimin Chen||, Gianluca Usala*, Fabio Busonero*, Andrea Maschio*, Giuseppe Albai*, Maria Grazia Piras*, Natascia Sestu*, Sandra Lai*, Mariano Dei*, Antonella Mulas*, Laura Crisponi*, Silvia Naitza*, Isadora Asunis*, Manila Deiana*, Ramaiah Nagaraja**, Lucia Perseu*, Stefania Satta†, Maria Dolores Cipollina†, Carla Sollaino†, Paolo Moi†, Joel N. Hirschhorn*§, Stuart H. Orkin*¶††, Gonçalo R. Abecasis||, David Schlessinger**, and Antonio Cao*††

BCL11A ↓ = fetal globin ↑ = disease ameliorated or eliminated

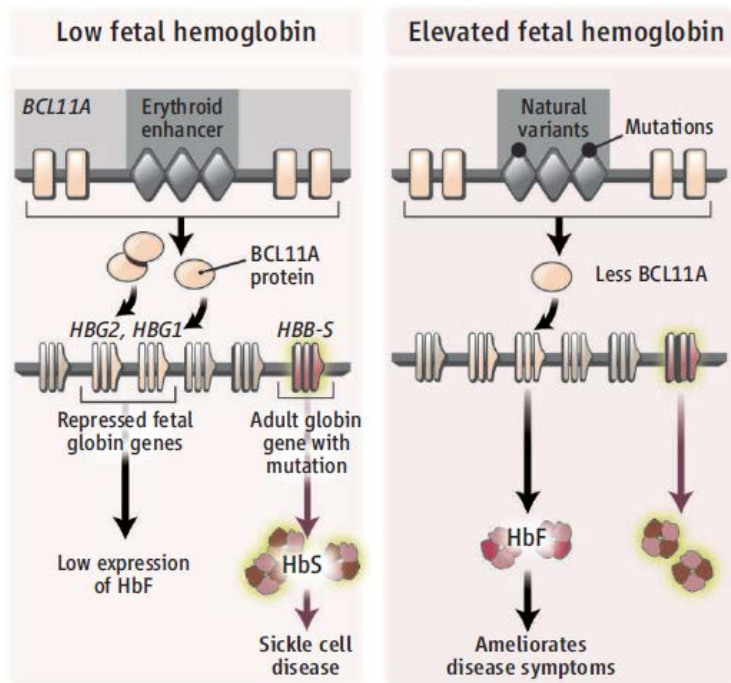
Genome Editing for β -thalassemia and Sickle Cell Disease: a “Simple” Plan



The Achilles Heel of the BCL11A Erythroid Enhancer

Bauer/Orkin – *Science* Oct 2013

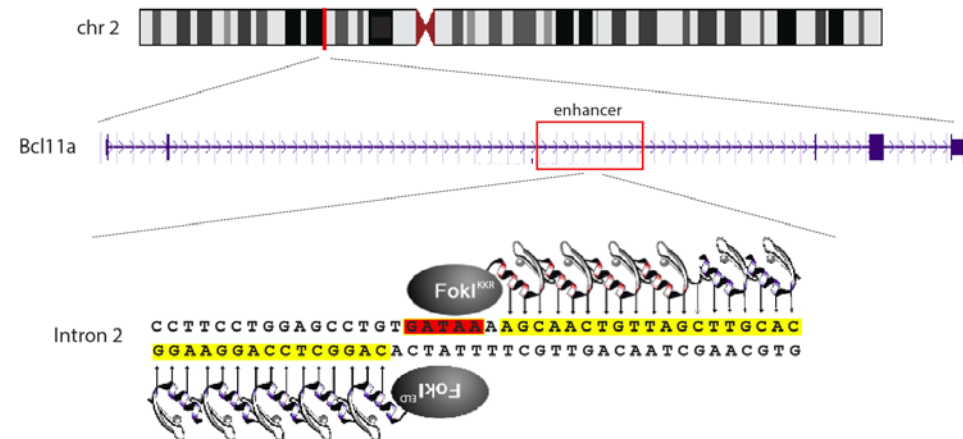
“We propose the GWAS-identified enhancer of BCL11A as a particularly promising therapeutic target for genome engineering in the β -hemoglobinopathies.”



Vierstra et al *Nat Meth* 2015

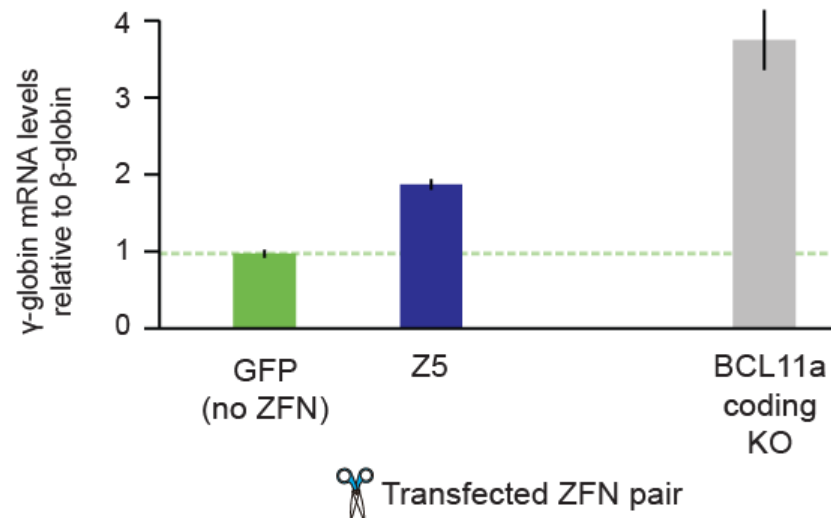
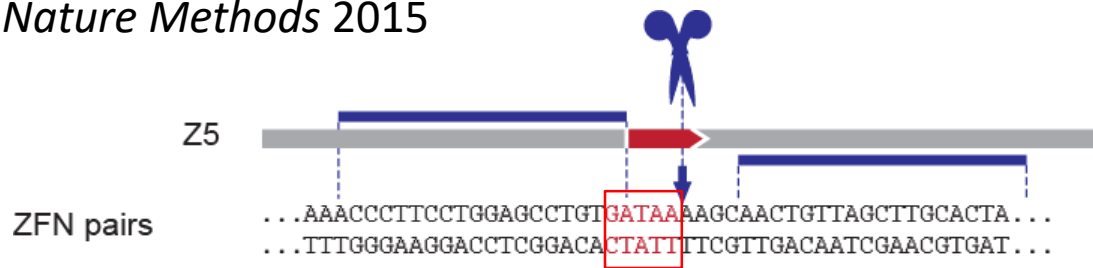
A genome editing-based “walk” across the enhancer using two orthogonal nuclease platforms illuminated a single, specific binding site for GATA1 as key to enhancer function in erythroid-specific BCL11A activation and fetal globin suppression.

(also see Canver et al *Nature* 2015)



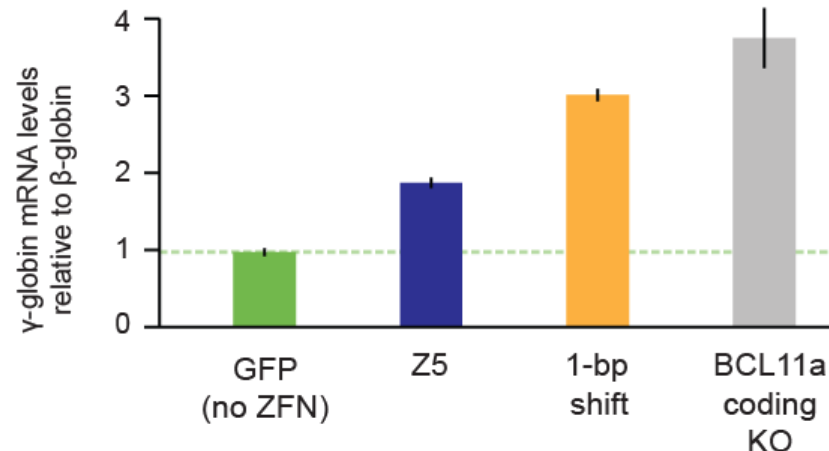
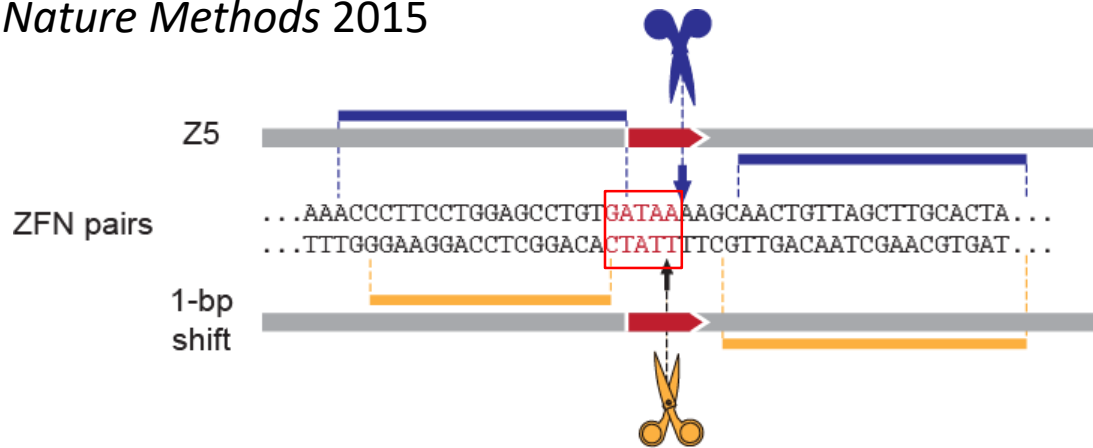
Initial Hit Yielded Enhanced γ -globin Levels

Vierstra et al *Nature Methods* 2015



Fine-tuning Cut Site Further Enhances γ -globin Levels

Vierstra et al *Nature Methods* 2015



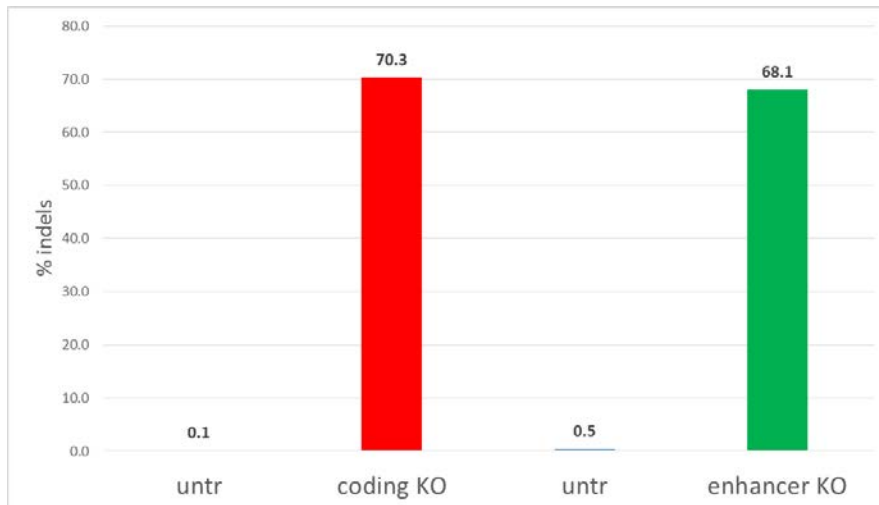
Transfected ZFN pair

BCL11A Knockout vs Enhancer Disruption: Equivalent Effects on Fetal Globin in PB-derived HSPCs

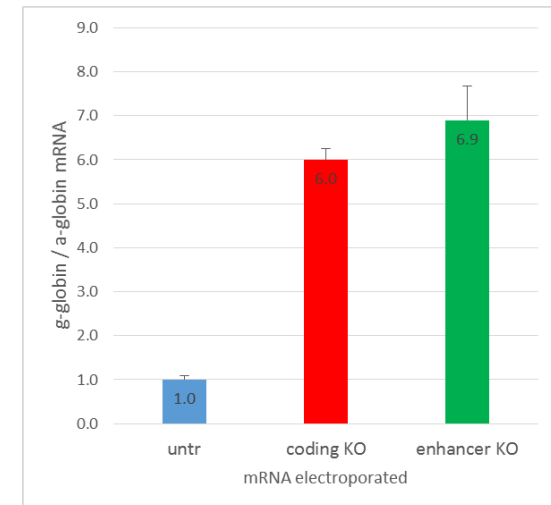


- erythropoiesis proceeds indistinguishably to control cells (Vierstra et al 2015)
- equivalent elevation of gamma globin in RBCs obtained from PB-derived HSPCs

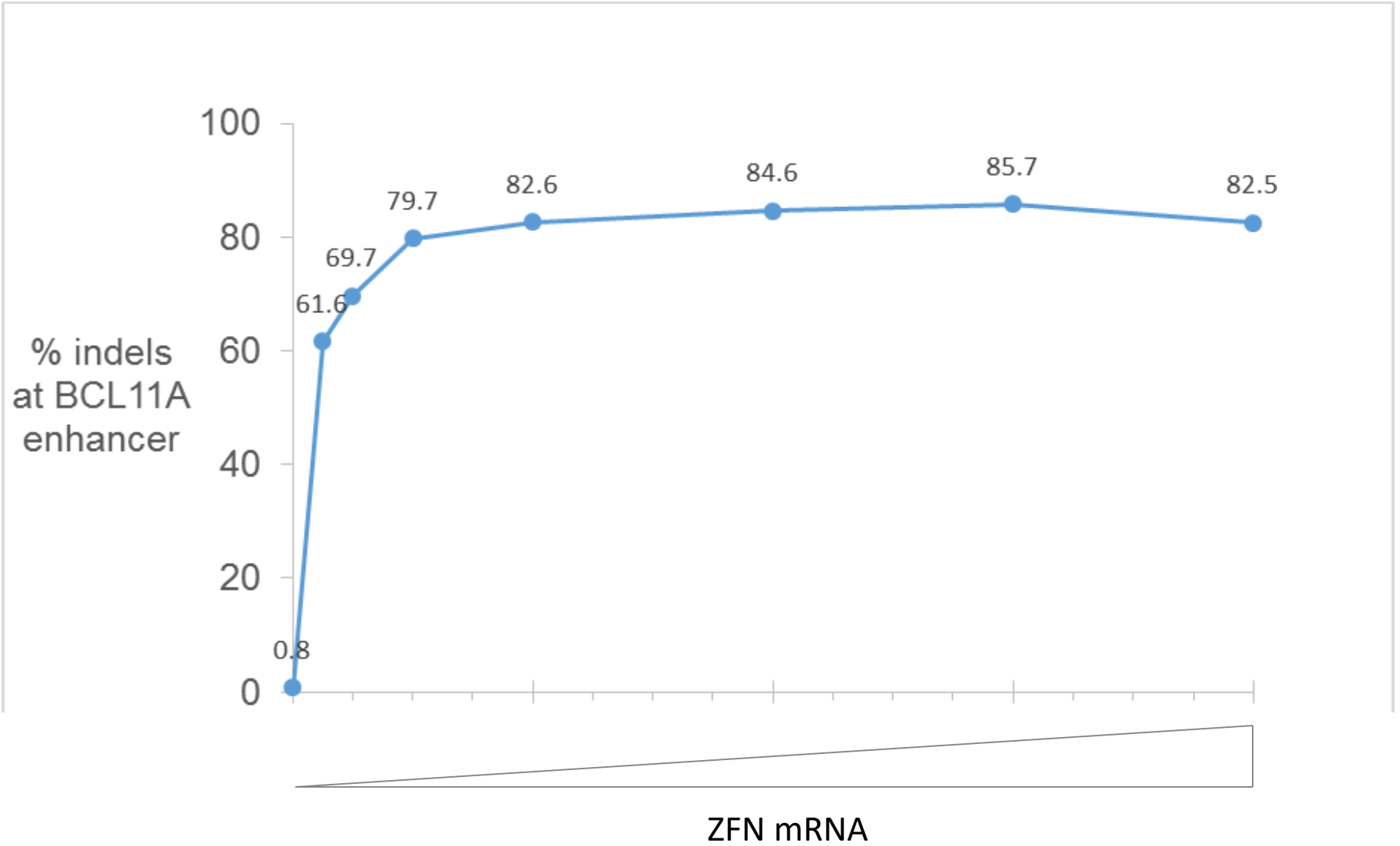
Editing



Level of fetal globin mRNA



85% On-Target Editing in HSPCs Using Clinical Lead ZFNs



Genome Editing En Route to the Clinic: CD34 GMP Cell Manufacturing Process

Day -2 & -1



Apheresis from G-CSF/Plerixafor mobilized subjects

Day 0



Platelet depletion

Day 0



CliniMACS
CD34 enrichment



Day 0-1



Rest cells in culture

Day 2



Maxcyte ZFN mRNA
electroporation

Day 2-4

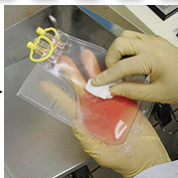


Static culture step to allow
ZFN gene expression

Day 4



Formulate CD34 cells



Gene modified CD34 cells
Final Product after QC
testing and release

From GWAS to the Clinic

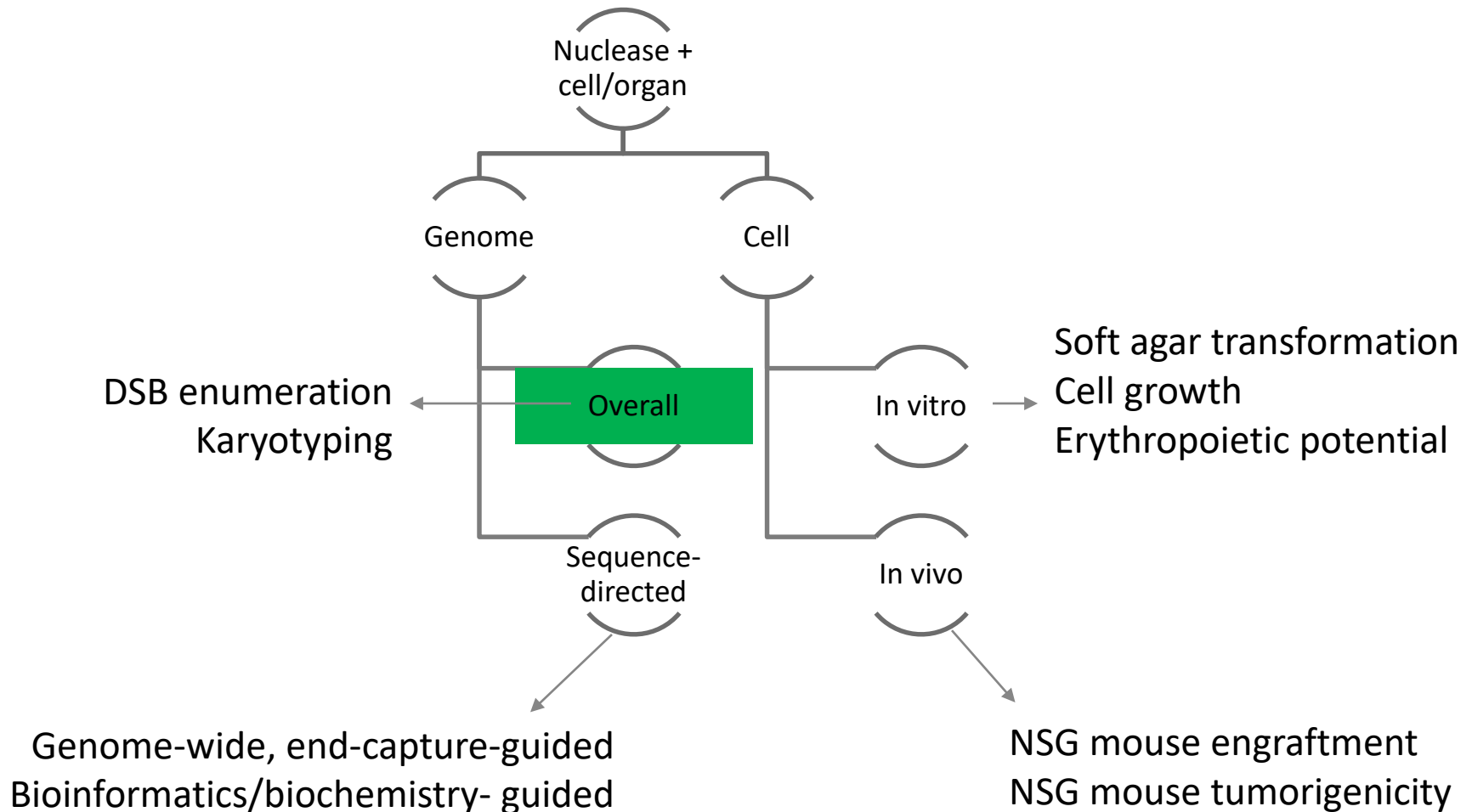
Proposal (Bauer et al (2013)):

“We propose the GWAS-identified enhancer of BCL11A as a particularly promising therapeutic target for genome engineering in the β -hemoglobinopathies”

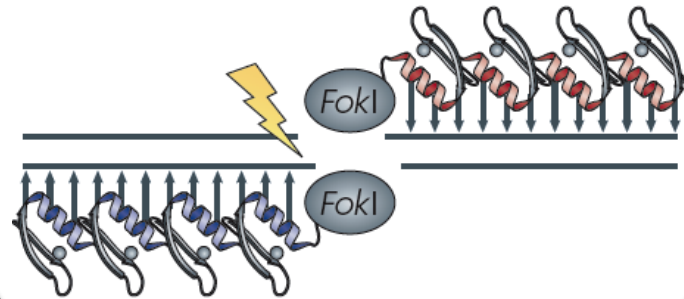
Findings:

- An editable “Achilles Heel” in the enhancer
- Enhancer KO elevates disease-protective fetal globin
- Efficient knockout at clinical scale
- **On-track for IND filing H1 2016 for our beta-thalassemia program**

Readouts of Genome Editing Safety: Charted Path Based on Two Open INDs For ZFN-Driven HSPC Editing



Human Genome Editing With Engineered ZFNs: Targeted Gene Addition



HDR

*

Gene correction

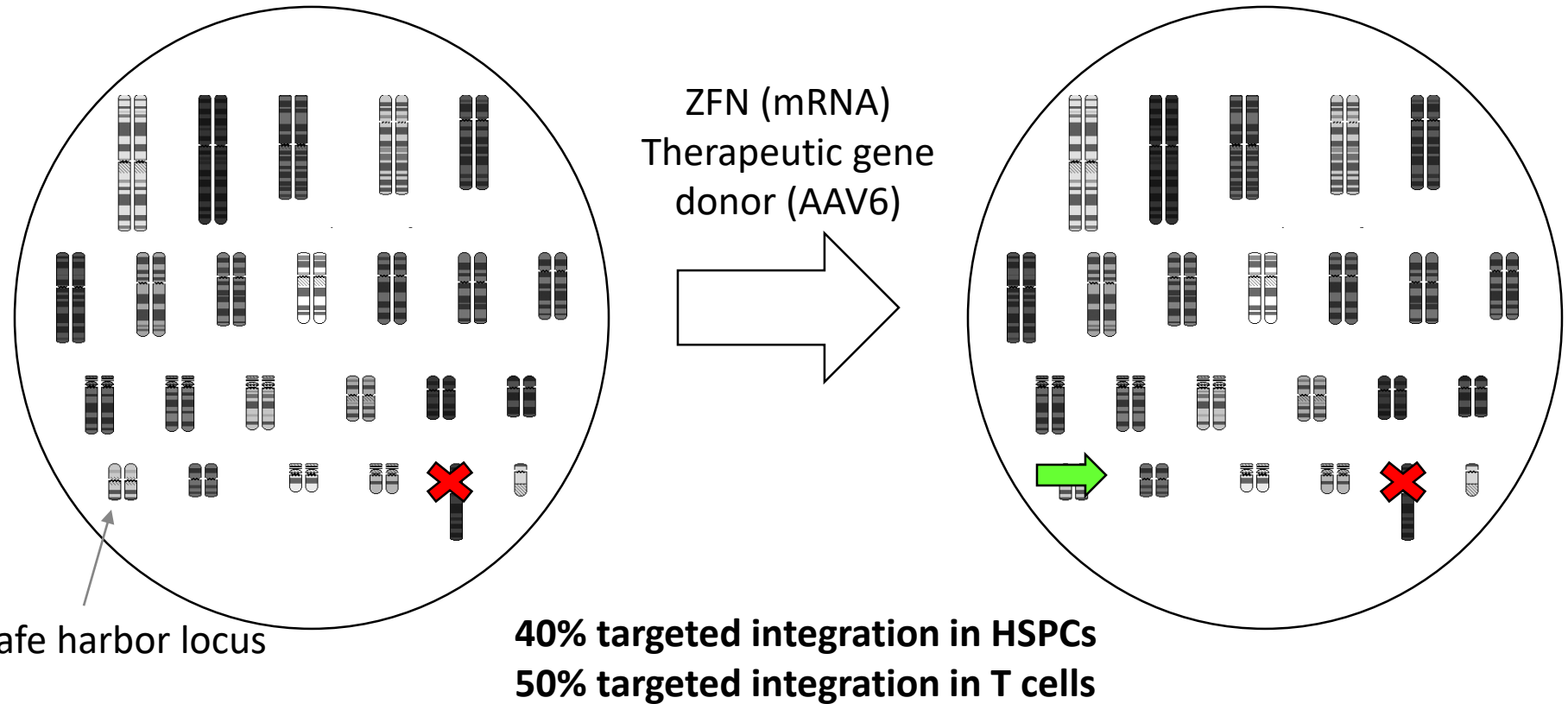


Targeted gene addition



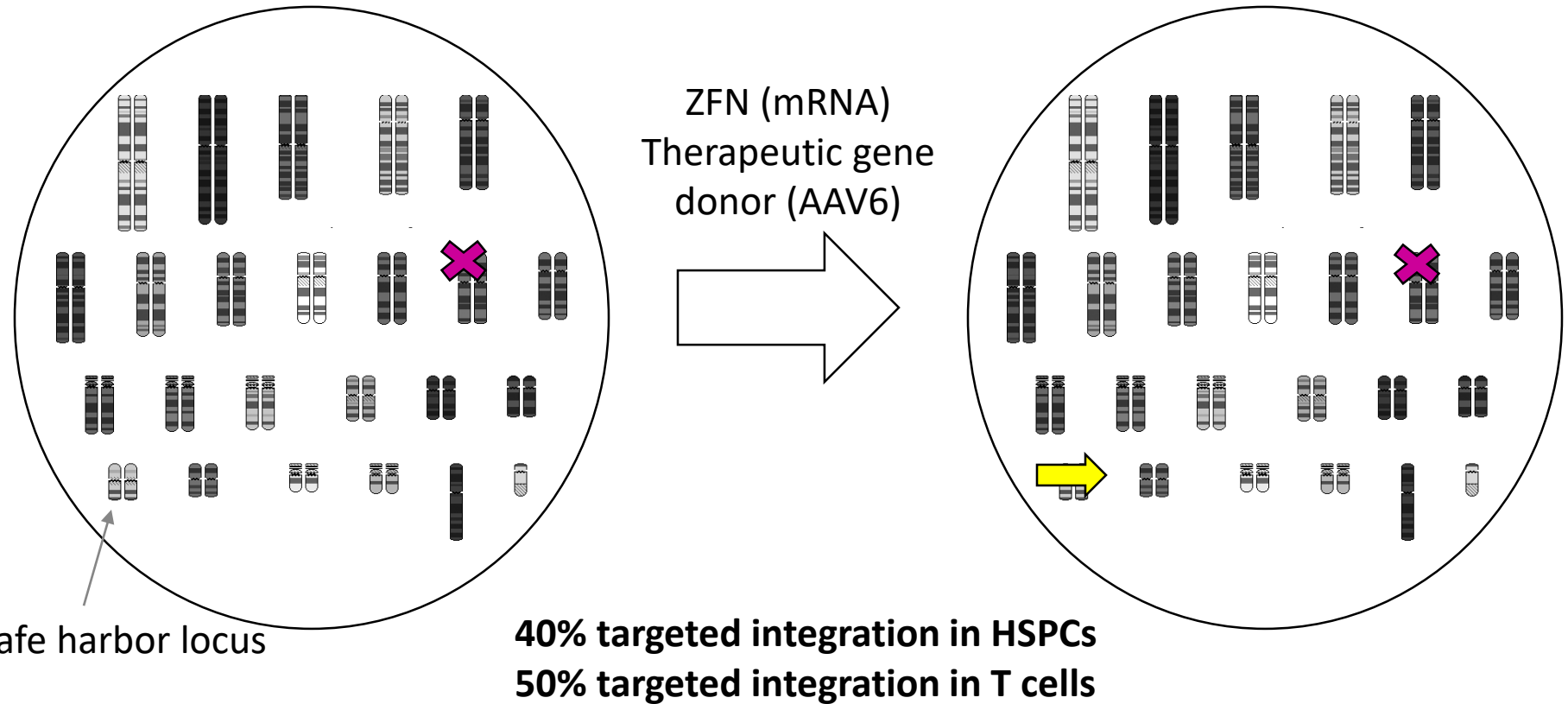
Transgene stacking

Targeted Addition of Therapeutic Gene to a “Safe Harbor”



Hockemeyer et al *Nature Biotechnology* 2009
DeKolver et al *Genome Research* 2010
Lombardo et al *Nature Methods* 2011
Wang et al *Nature Biotechnology* 2015; *NAR* 015

Targeted Addition of Therapeutic Gene to a “Safe Harbor”



Hockemeyer et al *Nature Biotechnology* 2009
DeKolver et al *Genome Research* 2010
Lombardo et al *Nature Methods* 2011
Wang et al *Nature Biotechnology* 2015; *NAR* 015

Targeted Gene Integration via Systemic AAV Delivery of ZFP Therapeutics: *In Vivo* Treatment of Monogenic Disease

Human albumin
ZFN Pair and
Corrective Gene

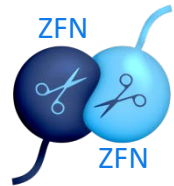
Nucleic Acids Coding for
ZFNs and Corrective Gene
in AAV Vectors

Peripheral IV
Administration

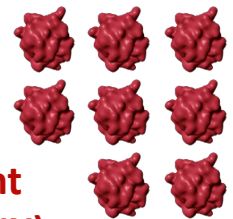
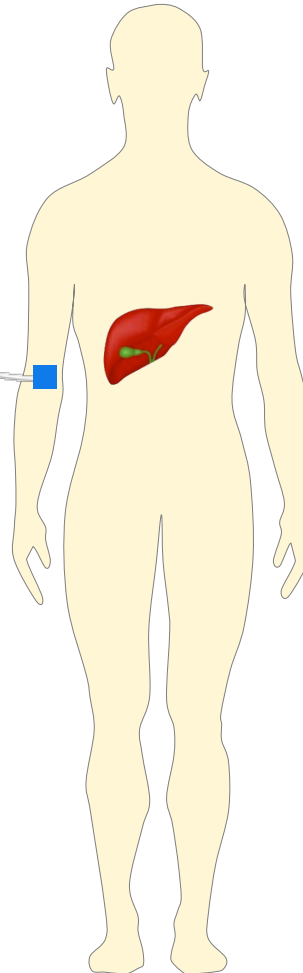
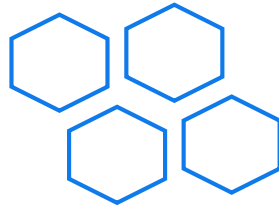
AAV Traffics to Liver
And ZFNs Permanently
Modify the Genome

Liver Cells Secrete
Corrected Protein

In Vivo
Protein
Replacement
(e.g. Factor IX)



AAV2/6 Vectors

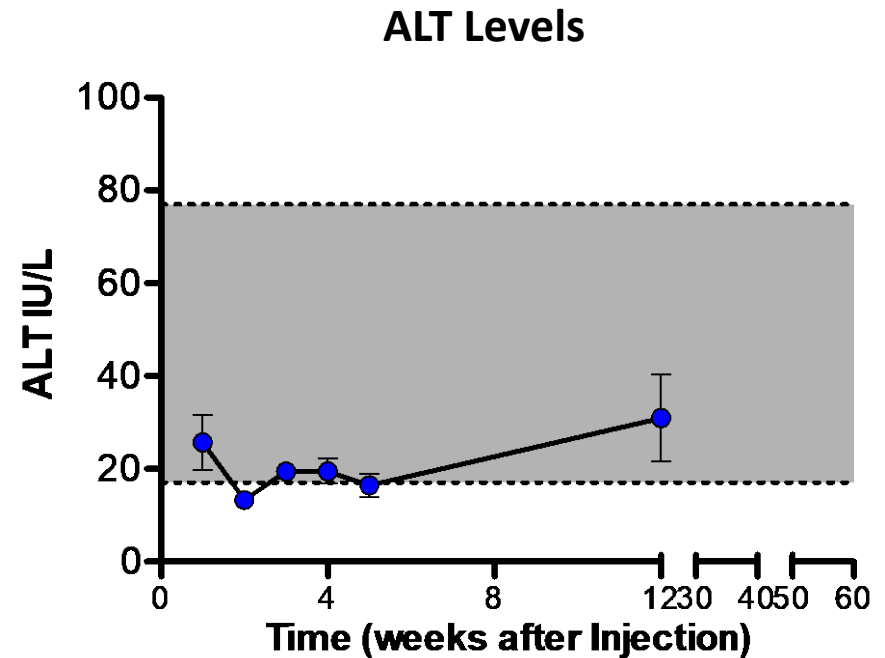
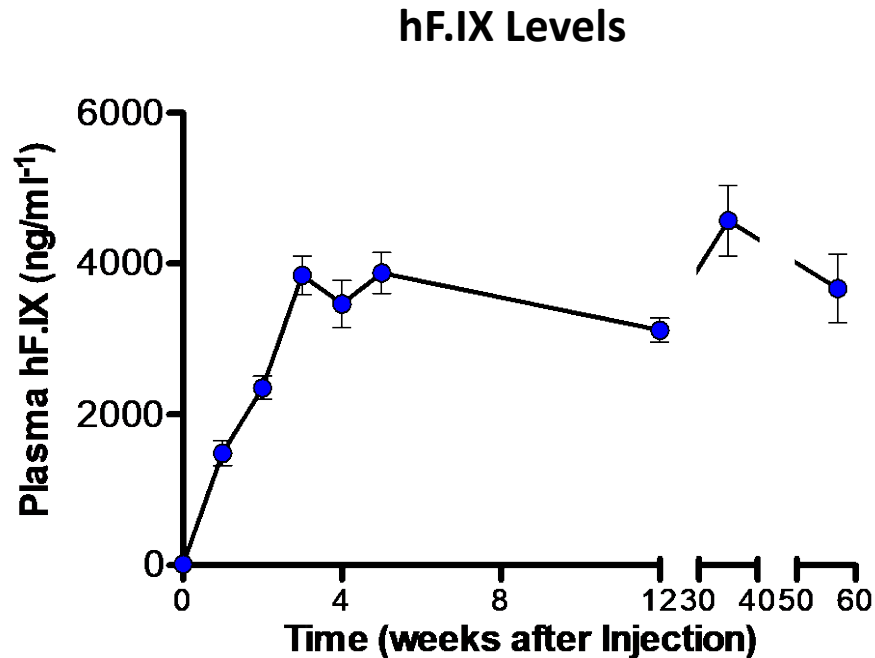


Homology Corrective Gene Homology

Also see Li, Holmes, High et al *Nature* 2011

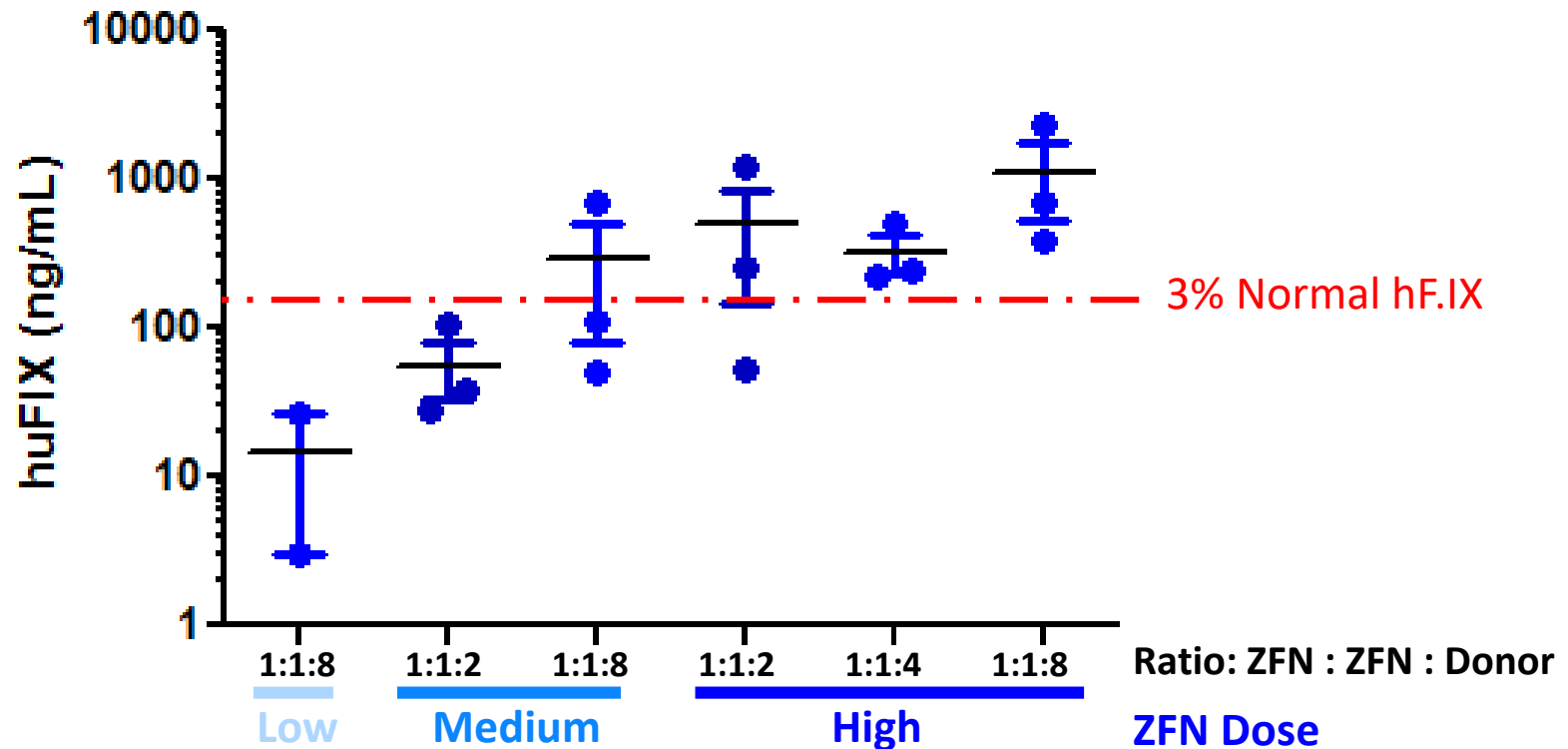
***In Vivo* Genome Editing: Robust Circulating hF.IX Levels**

Following Its ZFN-Driven Addition to Albumin – **at 60 wks**



Therapeutic hF.IX levels observed for 60 weeks (longest time point assayed)
No change in ALT levels

NHP Genome Editing: Therapeutic hF.IX Levels Following Its Addition to ALB



IND reviewed by FDA and is open – first study of in vivo genome editing

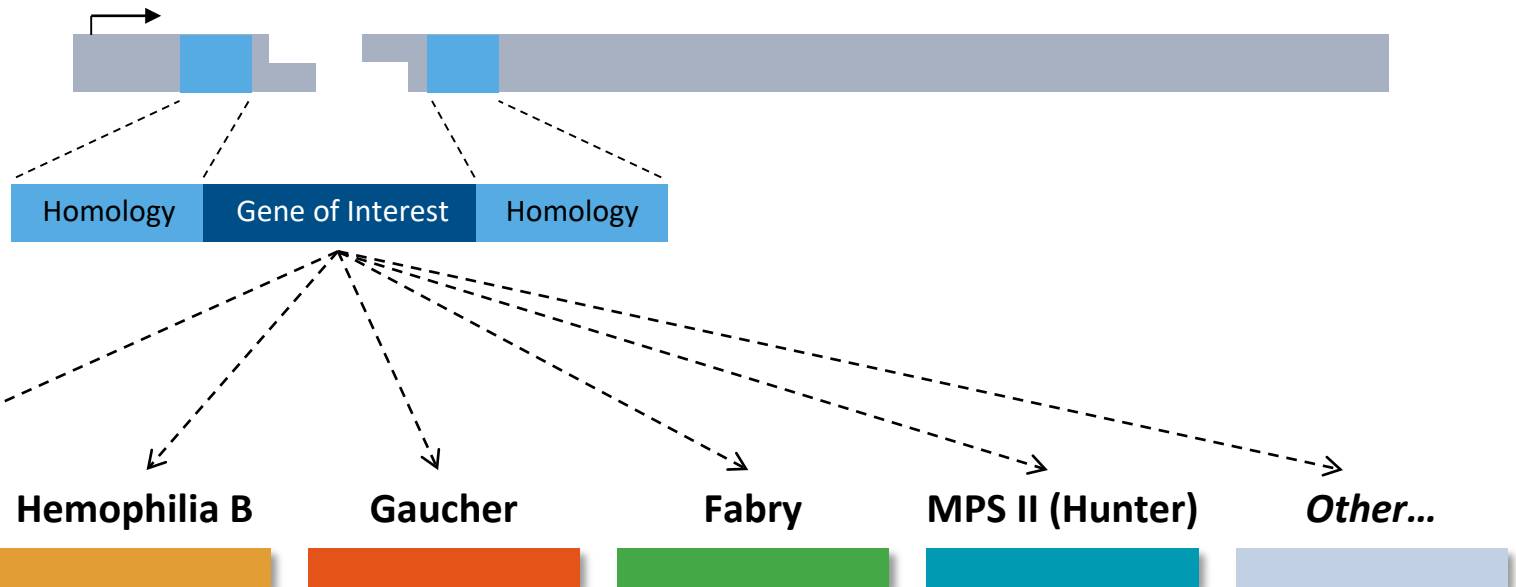
Sangamo's *In Vivo* Protein Replacement Platform

Broadly Leverageable

Albumin Gene:

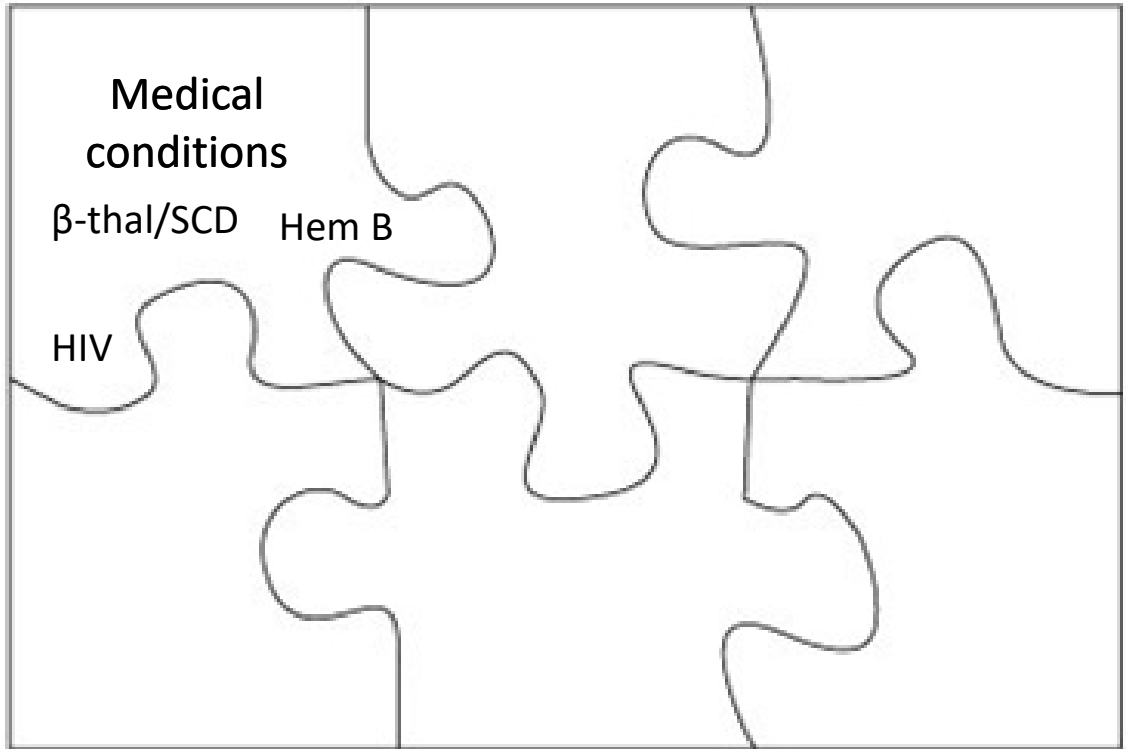


Donor:



A quote from Sydney Brenner, and a look to the future

“Progress in science depends on new technologies, new ideas, and new discoveries
– **in that order**” Sydney Brenner

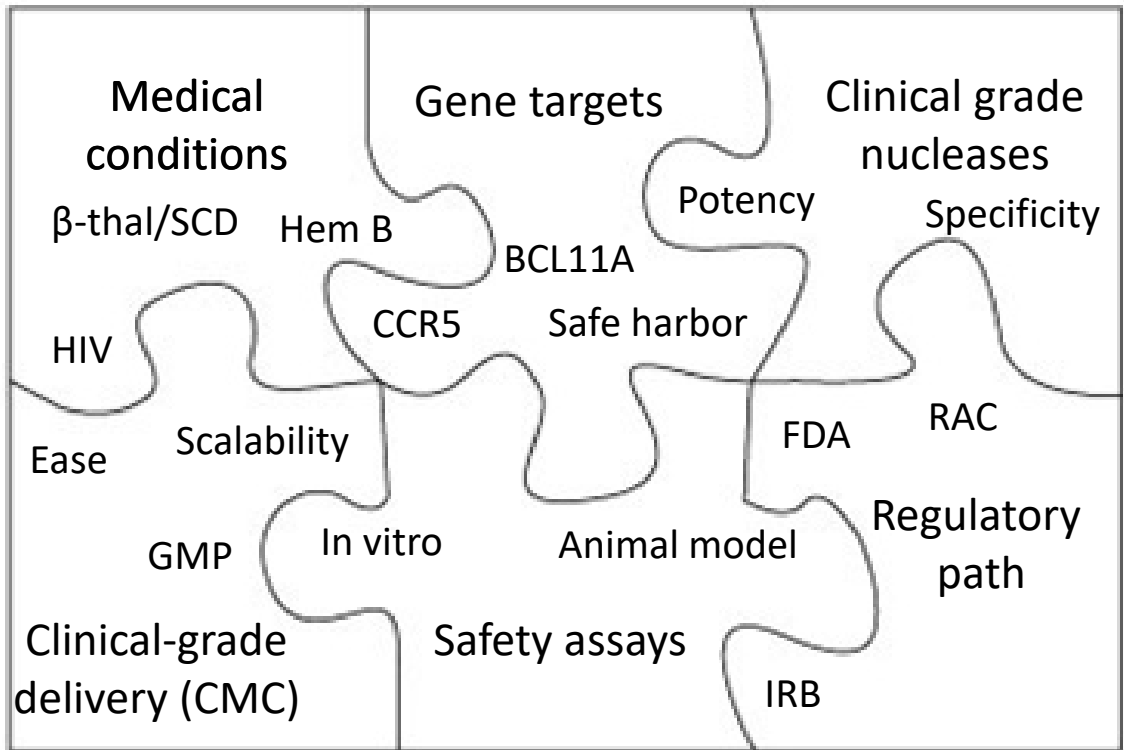


A quote from Sydney Brenner, and a look to the future

“Progress in science depends on new technologies, new ideas, and new discoveries
– **in that order**” Sydney Brenner

“Pharmacological-grade” genome editing:

- Potency
- Specificity
- Delivery
- Regulatory path



Debt of gratitude: “academic” research

Zinc finger design

Rebar and Pabo *Science* 1994

Isalan Klug *Nature Biotech* 2001

Editing

Rouet and Jasin MCB, PNAS 1994

Bibikova and Carroll MCB 2001, Genetics
2002, Science 2003

Porteus and Baltimore Science 2003

Debt of gratitude: collaborators and colleagues

Collaborators

HIV: Carl June, Pablo Tebas, Bruce Levine (Penn); G. Blick, J. Lalezari.

β -thalassemia: George Stamatoyannopoulos, Thalia Papayannopoulou, Kai-Hsin Chang (UW) Jeff Vierstra, John Stamatoyannopoulos (UW)

Stuart Orkin, Daniel Bauer (Boston Children's/DFCI)

Haiyan Jiang, Siyuan Tan, Kasra Kasraian (Biogen)

Sangamo

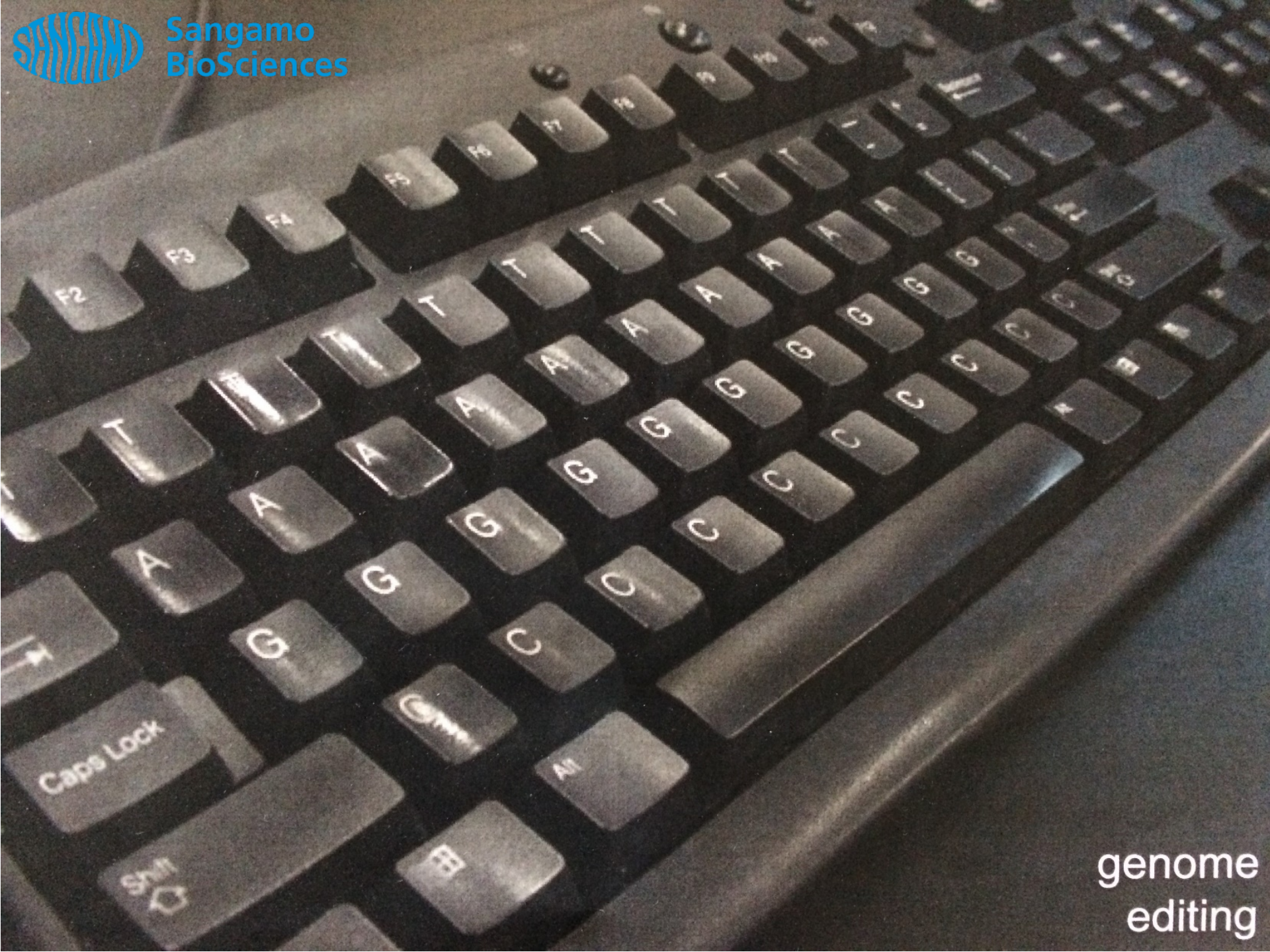
Edward Rebar, Jeffrey Miller, Lei Zhang
Andreas Reik

Lisa Fox

Michael Holmes, Thomas Wechsler

Philip Gregory

Dale Ando, Geoff Nichol



genome
editing