

National Aeronautics and
Space Administration



NASA's Multicenter Planetary Protection Metagenomics Study

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NASA Standard Spore Assay (NSA)

- Fast, Reproducible, Easy, Quantifiable

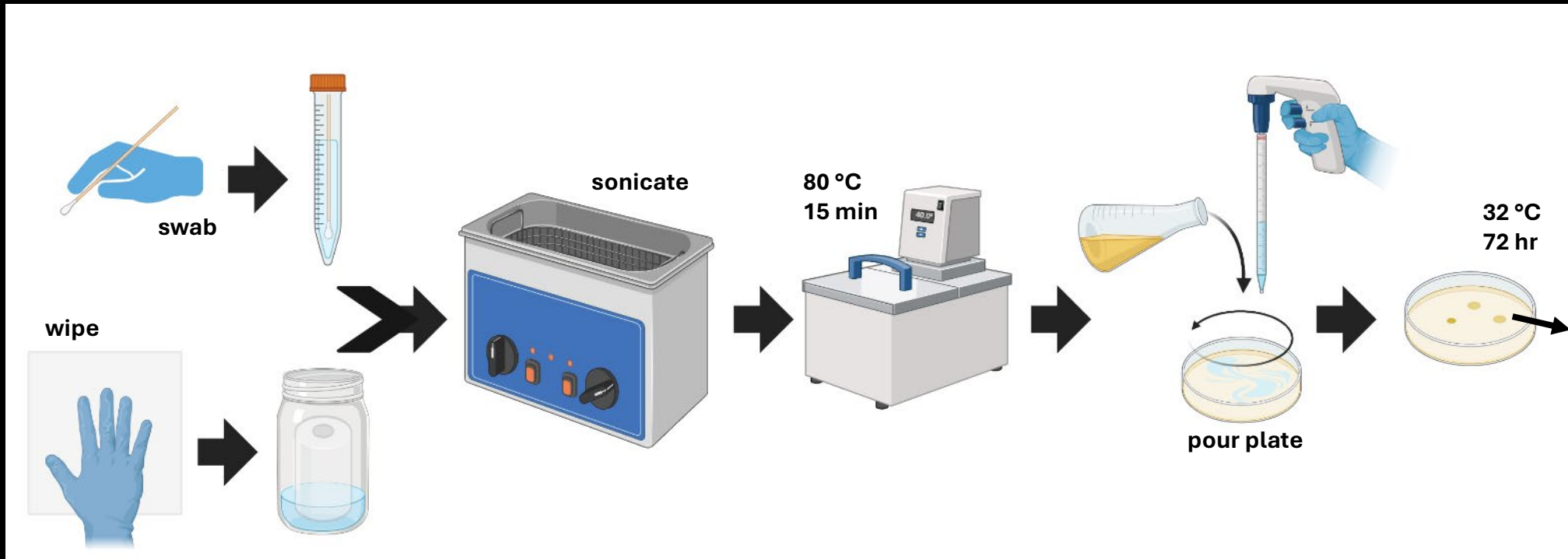
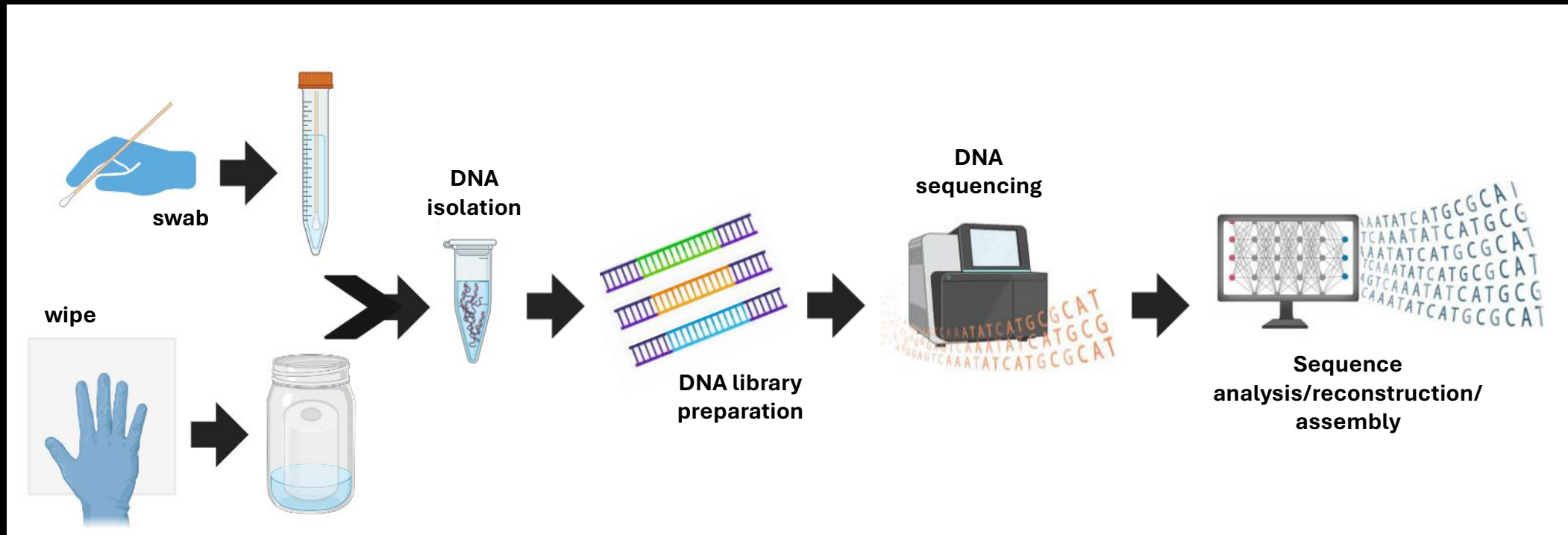


Image created with BioRender.com

Modernization of PP Methods

- **Metagenomics** is the study of all genetic material (DNA) from all organisms in a particular environment.





Details and Objectives

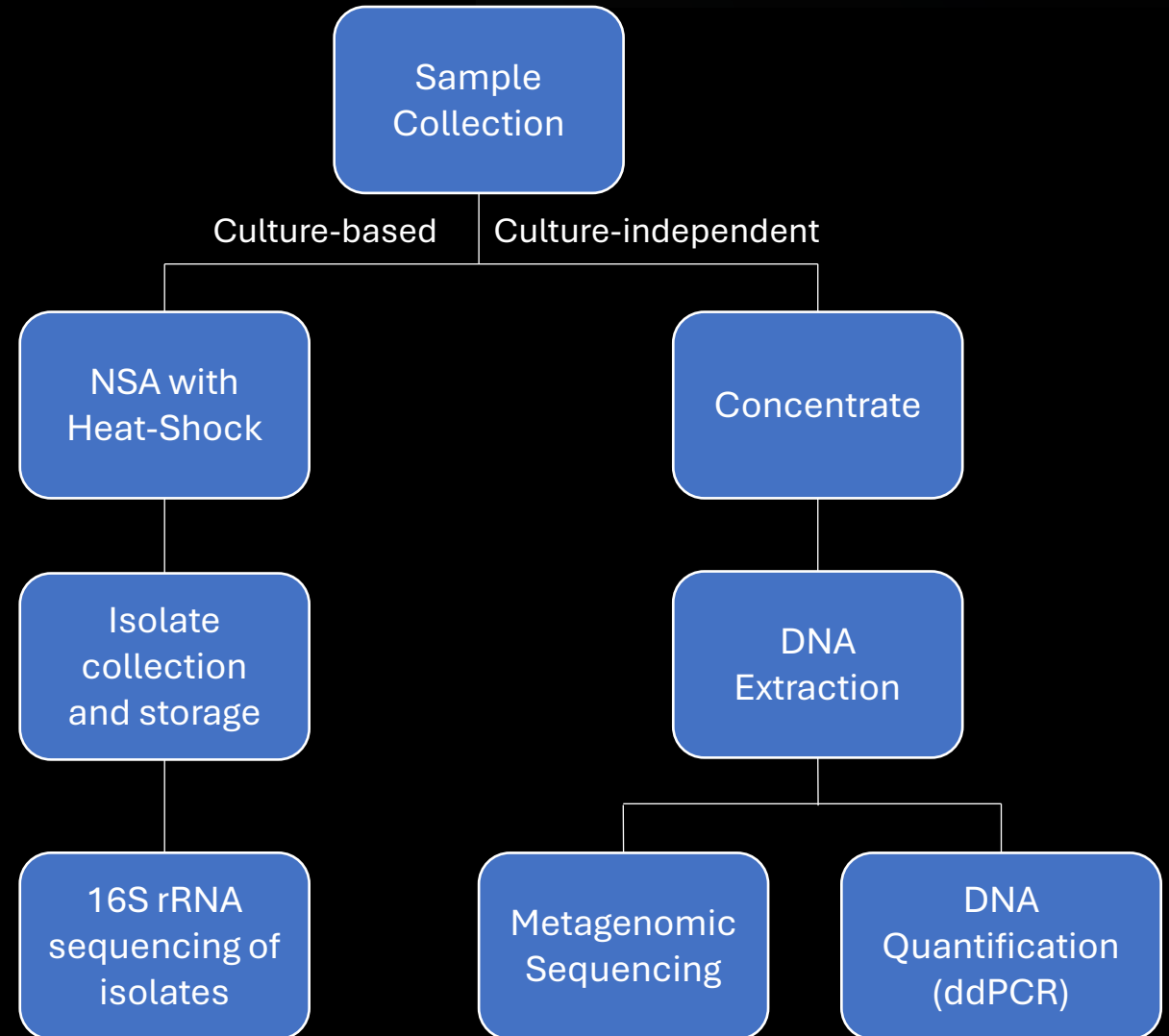
- Timeframe: September 2024 – ongoing
- What / Vision
 - To collect cleanroom environmental samples from multiple NASA Centers to compare metagenomic analysis with NASA Standard Spore Assay (NSA)
- Objectives
 - Evaluate cleanrooms across NASA Centers using metagenomics and the NSA
 - Strengthen NASA's PP SME community and build collaboration
 - Publication of findings
 - Eventual: collaborate with the international community on an international standard



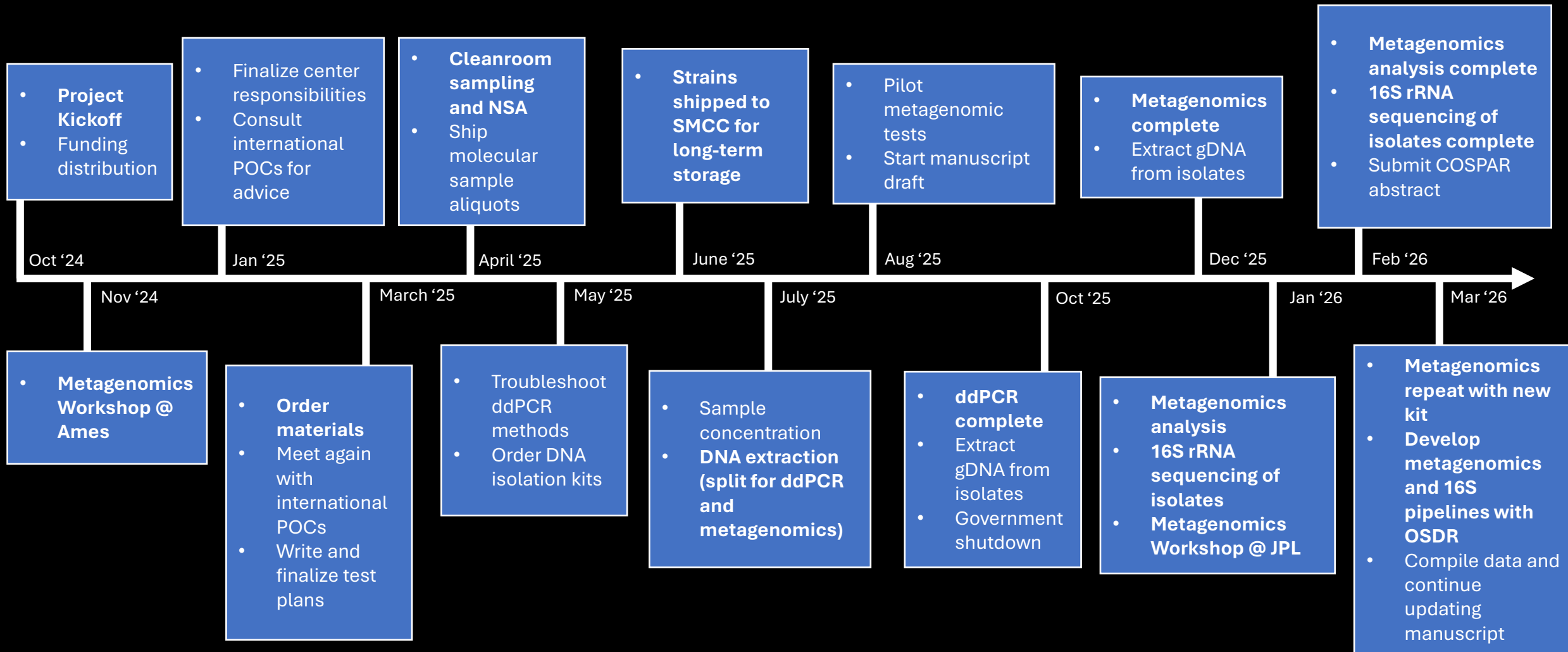


Key Tasks

- Develop/edit test plan (including shipment procedures)
- Cleanroom wipe sampling
- Sample processing
- NSA
- Isolate archiving
- Isolate ID (amplicon sequencing)
- Sample concentration
- DNA extraction
- ddPCR
- Metagenomics
- Bioinformatics



Workflow Timeline

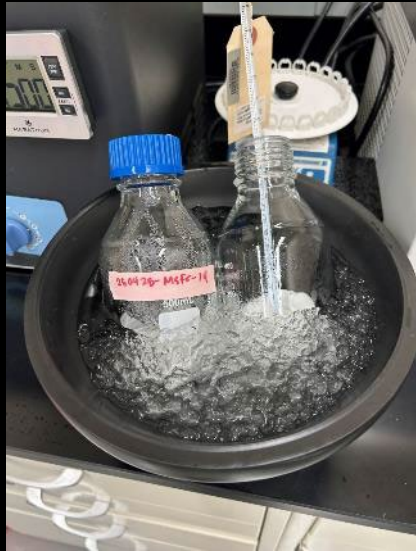


Test Plan Development and Cleanroom Sampling



- Developed test plan and procedure
- 3 centers selected a cleanroom for sampling:
 - 4 heavy traffic walkways
 - 2 minor trafficked areas
 - 1 corner
 - 1 surface or table
 - 2 field blank
 - 2 DNA Spike controls (Zymo D6321 Mock Community)

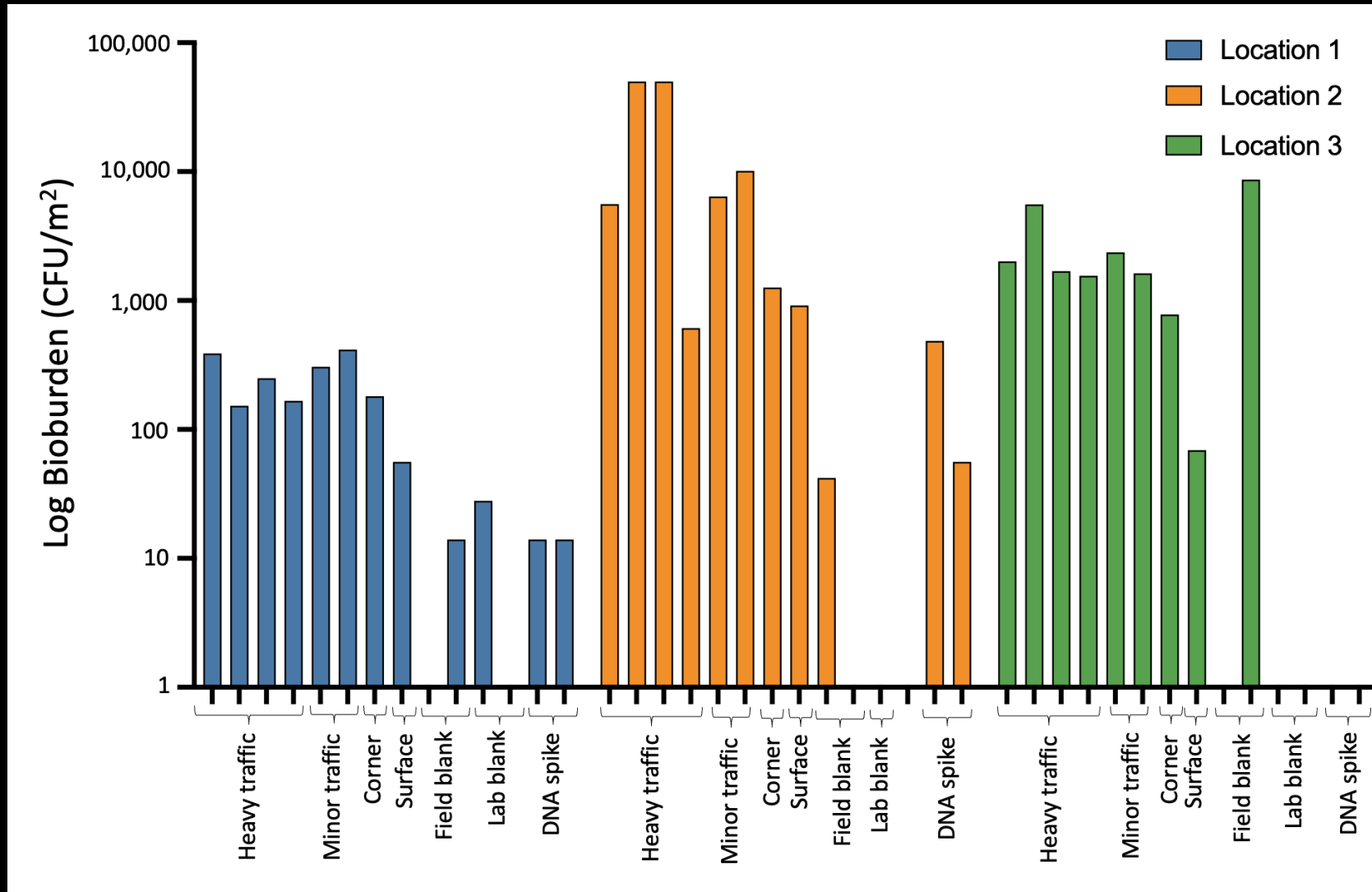
Sample Processing and NSA



- Samples were processed at two different centers
- Prior to heat shock, 100 ml of each sample was removed for molecular analysis and frozen at $-80\text{ }^{\circ}\text{C}$
- Remaining 100 ml was processed via NSA
- After 72 hours of incubation, representative isolates were streaked

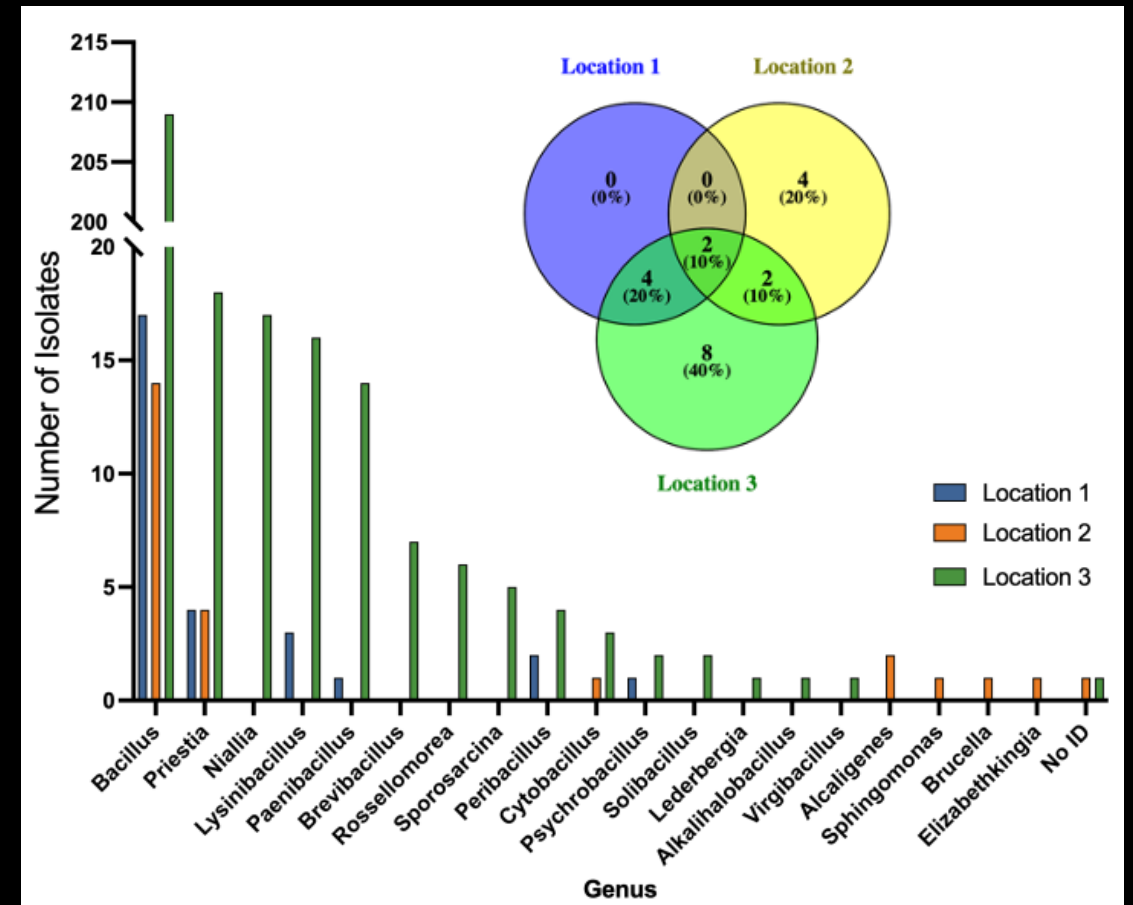


Sample Processing and NSA



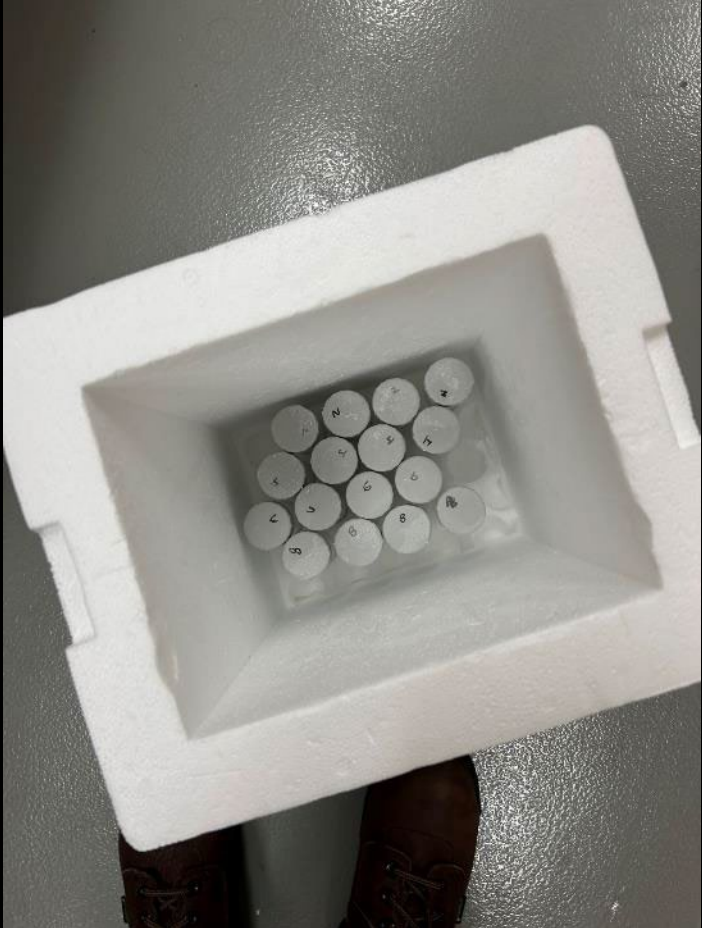
Isolate Archiving and Identification

- Pure cultures were collected and frozen at -80 °C
- Over 360 isolates were shipped to ARC's Space Microbial Culture Collection (SMCC)
- Sequenced 16S rRNA gene to identify
 - Working with OSDR to develop standardized pipeline for taxonomy calling



Oliveros, J.C. (2007-2015) Venny. An interactive tool for comparing lists with Venn's diagrams. <https://bioinfogp.cnb.csic.es/tools/venny/index.html>

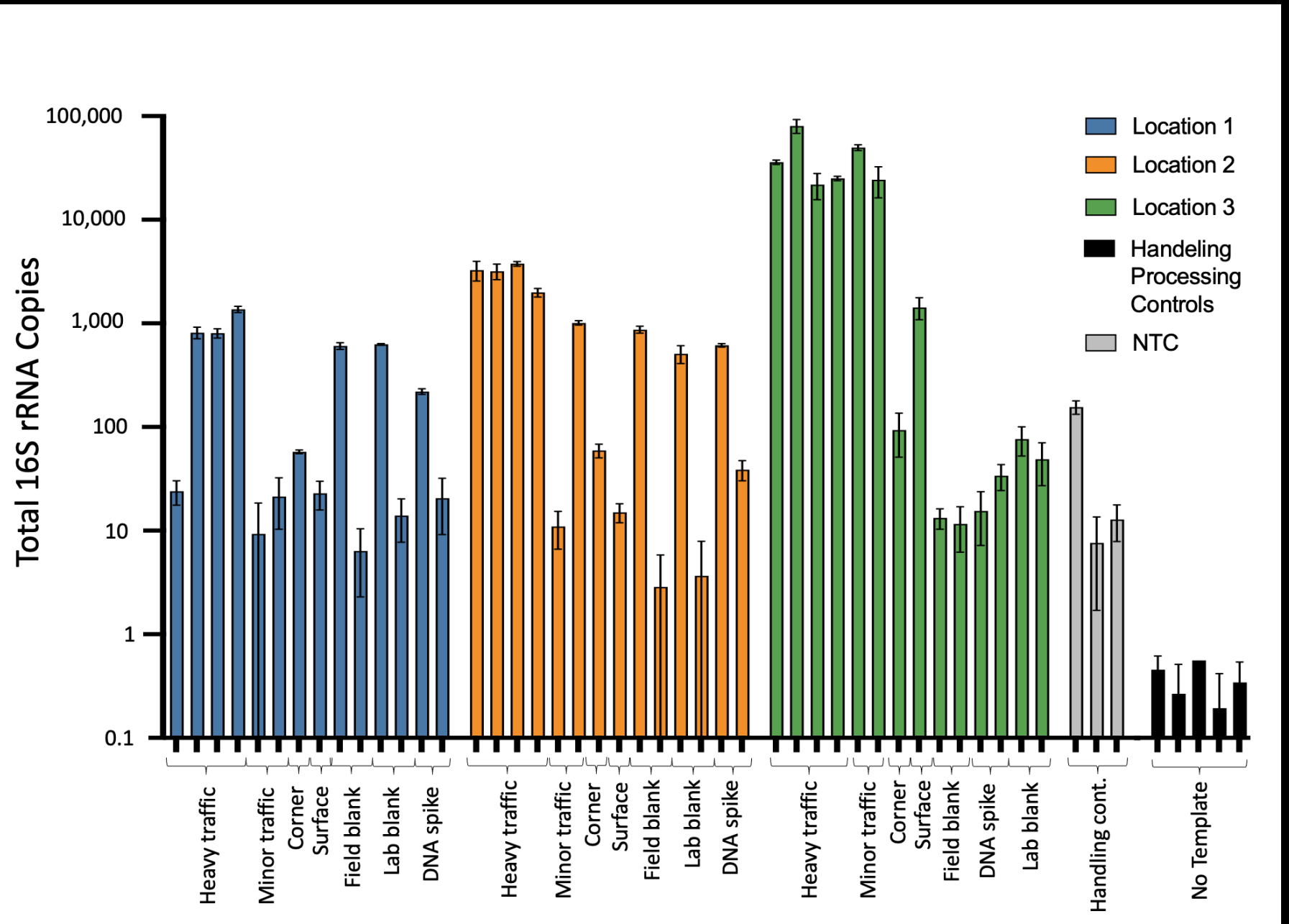
Sample Concentration and DNA Extraction



- 100 ml aliquots saved from samples were shipped blind on dry ice
- Samples were concentrated using an InnovaPrep FluidPrep Concentrating Pipette System
- DNA was extracted from concentrated samples using the ZymoBIOMICS DNA Microprep Kit
- All DNA was shipped for ddPCR or metagenomics

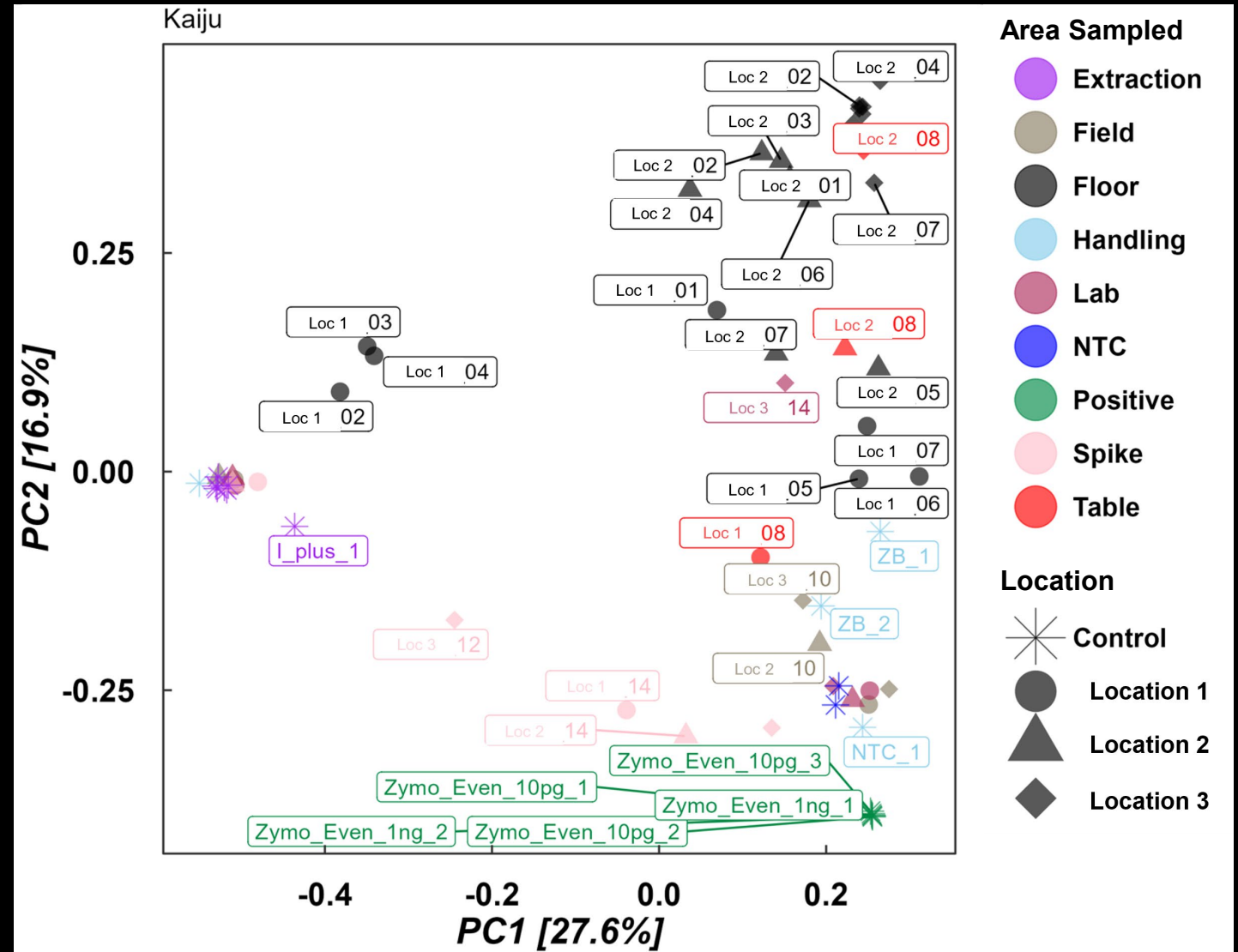
ddPCR

- ddPCR of 16S rRNA performed on BioRad QX200 with Manual Droplet Generator



Metagenomics and Bioinformatics

- TapeStation & Qubit showed that the DNA concentration was below detection for all samples
- Libraries were constructed using the Illumina DNA Prep Kit and pooled.
- Sequencing was performed on an Illumina NovaSeq 6000 using a SP 500 cycle flow cell
- Area sampled and location may explain some clustering

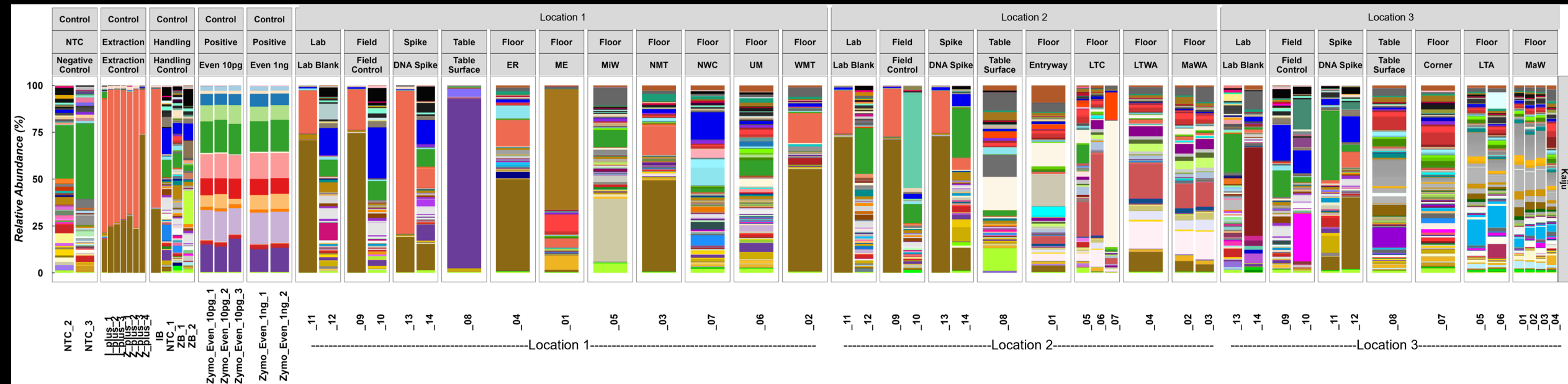


Metagenomics and Bioinformatics

- ME - Main entrance floor
- WMT - Floor west of main traffic flow
- NMT - Floor north of main traffic flow
- ER - Floor on east side of room
- MiW - Minor walkway floor, between hardware cages

- UM - Upper mezzanine floor
- NWC - Floor of northwest corner
- Entryway - Entryway floor
- MaWA - Major work area floor
- LTWA - Less trafficked work area floor

- LTC - Less trafficked corner floor
- MaW - Major walkway floor
- LTA - Less trafficked area floor
- Corner - Corner floor



- Cleanroom samples were dominated by extremophiles, human associated microbes, and plants
- Microbial composition was different depending on area sampled and location (appears that ISO 7 has less diversity than the ISO 8 cleanrooms with Location 3 having the most)
- The dominant species were different for each location

Metagenomics and Bioinformatics

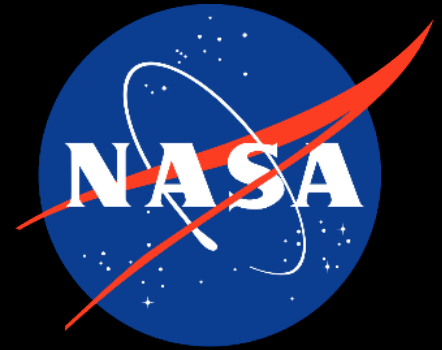
- Still interpreting the data, but our use of blanks, and controls is allowing us to identify steps in the process where contamination is occurring
- Variable read depth and quality for samples using Illumina library prep. kit.
- Trying a Takara DNA Prep Kit currently
- Data from both kit runs will be available publicly in OSDR
- Pipelines for metagenomic data analysis in preparation

Takeaways and Next Steps

- Sample processing for molecular analysis matters
 - Variability between centers
 - Assessing differences in Takara vs. Illumina library prep kits
- Working with OSDR to develop standardized pipeline for metagenomics analysis and 16S taxonomy calling to increase efficiency and decrease data processing timeline
- Manuscript in preparation to submit soon
- Submitted abstract to present this work at COSPAR (1 – 9 August)
- Open to additional follow-up studies and collaborations
 - Strengthen statistical considerations and use of data for decision making

We have a great team!

Limited resources, backorders, layoffs/personnel changes, new administration, 4 parental leave considerations, Government shutdown, Software installation delays, Shipment constraints, etc.



- **NASA MSFC**

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- Carly Parker

- **NASA LaRC**

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- **NASA GSFC**

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- Dr. Richard Davis

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- Dr. Jonathan Galazka
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