### Genetically Engineered Primate Models for Brain Disorder Research



#### **Guoping Feng, PhD**

McGovern Institute for Brain Research, Massachusetts Institute of Technology Stanley Center for Psychiatric Research Broad Institute of Harvard and MIT

# The need for better models in brain disorder research

#### Genetic engineering in mice has revolutionized biomedical research. However, its impact on developing treatments for brain disorders is limited.

#### Limitations of rodent models for brain disorders:

- 1. Huge differences in both structure and function between rodent and human brain.
- 2. Evolutional divergence in behaviors and underlying circuits
- 3. Numerous failures in translating preclinical success in rodents to clinical trials in humans



The lack of good animal models is considered one of bottlenecks to the development of new drugs for brain disorders

## Better animal models for prefrontal cortex and developmental studies



## Feasibility: New genome-editing technology—CRISPR/Cas9 and TALEN



- 1. Highly efficient (heterozygous and homozygous mutations)
- 2. Can make multiple mutations in parallel
- 3. Direct embryo injection leads to genetically engineered animals

#### Resource

#### Generation of Gene-Modified Cynomolgus Monkey via Cas9/RNA-Mediated Gene Targeting in One-Cell Embryos

Yuyu Niu,<sup>1,5,7</sup> Bin Shen,<sup>2,7</sup> Yiqiang Cui,<sup>3,7</sup> Yongchang Chen,<sup>1,5,7</sup> Jianying Wang,<sup>2</sup> Lei Wang,<sup>3</sup> Yu Kang,<sup>1,5</sup> Xiaoyang Zhao,<sup>4</sup> Wei Si,<sup>1,5</sup> Wei Li,<sup>4</sup> Andy Peng Xiang,<sup>6</sup> Jiankui Zhou,<sup>2</sup> Xuejiang Guo,<sup>3</sup> Ye Bi,<sup>3</sup> Chenyang Si,<sup>1,5</sup> Bian Hu,<sup>2</sup> Guoying Dong,<sup>3</sup> Hong Wang,<sup>1,5</sup> Zuomin Zhou,<sup>3</sup> Tianqing Li,<sup>1,5</sup> Tao Tan,<sup>1,5</sup> Xiuqiong Pu,<sup>1,5</sup> Fang Wang,<sup>1,5</sup> Shaohui Ji,<sup>1,5</sup> Qi Zhou,<sup>4</sup> Xingxu Huang,<sup>2,\*</sup> Weizhi Ji,<sup>1,5,\*</sup> and Jiahao Sha<sup>3,\*</sup>

<sup>1</sup>Yunnan Key Laboratory of Primate Biomedical Research, Kunming 650500, China

#### Cell Stem Cell Brief Report



#### TALEN-Mediated Gene Mutagenesis in Rhesus and Cynomolgus Monkeys

Hailiang Liu,<sup>1,2,9</sup> Yongchang Chen,<sup>1,3,4,9</sup> Yuyu Niu,<sup>1,3,4,9</sup> Kunshan Zhang,<sup>2,9</sup> Yu Kang,<sup>1,3</sup> Weihong Ge,<sup>6</sup> Xiaojing Liu,<sup>2</sup> Enfeng Zhao,<sup>2,6</sup> Chencheng Wang,<sup>2</sup> Shaoyun Lin,<sup>2</sup> Bo Jing,<sup>2</sup> Chenyang Si,<sup>1,3</sup> Quan Lin,<sup>2,6</sup> Xiaoying Chen,<sup>2,6</sup> Haijun Lin,<sup>2</sup> Xiuqiong Pu,<sup>1,3</sup> Yingying Wang,<sup>2</sup> Binlian Qin,<sup>2</sup> Fang Wang,<sup>1,3</sup> Hong Wang,<sup>1,3,4</sup> Wei Si,<sup>1,3,4</sup> Jing Zhou,<sup>2</sup> Tao Tan,<sup>1,3,4</sup> Tianqing Li,<sup>1,3,4</sup> Shaohu Ji,<sup>1,3,4</sup> Zhigang Xue,<sup>2</sup> Yuping Luo,<sup>2</sup> Liming Chen,<sup>2</sup> Qi Zhou,<sup>6</sup> Siguang Li,<sup>2,\*</sup> Yi Eve Sun,<sup>1,2,6,\*</sup> and Weizhi Ji<sup>1,3,7,\*</sup>

<sup>1</sup>Yunnan Key Laboratory of Primate Biomedical Research, Kunming, 650500, China

#### Cell Stem Cell Resource



#### Generation of a Nonhuman Primate Model of Severe Combined Immunodeficiency Using Highly Efficient Genome Editing

Kenya Sato,<sup>1</sup> Ryo Oiwa,<sup>1</sup> Wakako Kumita,<sup>1</sup> Rachel Henry,<sup>2</sup> Tetsushi Sakuma,<sup>3</sup> Ryoji Ito,<sup>1</sup> Ryoko Nozu,<sup>1</sup> Takashi Inoue,<sup>1</sup> Ikumi Katano,<sup>1</sup> Kengo Sato,<sup>4</sup> Norio Okahara,<sup>1</sup> Junko Okahara,<sup>1</sup> Yoshihisa Shimizu,<sup>1</sup> Masafumi Yamamoto,<sup>1</sup> Kisaburo Hanazawa,<sup>5</sup> Takao Kawakami,<sup>6</sup> Yoshie Kametani,<sup>7</sup> Ryuji Suzuki,<sup>8</sup> Takeshi Takahashi,<sup>1</sup> Edward J. Weinstein,<sup>2</sup> Takashi Yamamoto,<sup>3</sup> Yasubumi Sakakibara,<sup>4</sup> Sonoko Habu,<sup>9</sup> Jun-ichi Hata,<sup>1</sup> Hideyuki Okano,<sup>10,\*</sup> and Erika Sasaki<sup>1,11,\*</sup>

## Genome-editing in primates to generate better disease models?



<u>Macaque (Cynomolgus)</u> Large (7-20lb), 30 years of life span, native to Southeast Asia Mature in 4 years, 180 days gestation. Very similar to rhesus monkey Similar brain structure to humans Great for study of higher brain function



Common marmoset

Small (~350g), 12-16 years of life span, New World monkey Short reproduction cycle, mature in 1 year, 150 days gestation Give birth twice a year and produce twins each birth. Very social and communicative

# **Comparison of reproductive characteristics**

	Mouse	Marmoset	Macaque
Age of puberty	8 weeks	1 year old	3-4 years
Litter size	6-8	2-3	1
Gestation period	18 days	~140 days	~160 days
Delivery interval	20-28 days	~150 days	~550 days
offspring/year	36-48	4-6	<1



**Fast reproduction**: Faster to germline; large numbers of animals for behavioral testing; large numbers of animals for brain tissues.

## Precise engineering of human genetic mutations in primates



#### **Transgenic primates**

Disease models (knockouts and knockins):

#### **Gene deletions**

**Gene insertions** 

**Point mutations** 

**Conditional mutations** 

Genetic tools (knockins and transgenes): Cell type-specific Cre lines Cell type-specific indicators

**Cell type-specific actuators** 

# Genetically engineered marmosets as a primate model for studying brain disorders—a collaborative project at MIT and Broad Institute

#### Long term goal:

Establish genetically engineered primates as a genetic model for brain disorder research and new drug discovery, using CRISPR system for targeted genetic mutations. Program will serve as a hub for academic and commercial research activities nationwide.

## Current Status:

- 1. Established marmoset colonies
- 2. Establishing a genetic engineering platform—in collaboration with Jim Pickel and Afonso Silva at NIMH and Hideyuki Okano and Erika Sasaki at Keio University and CIEA.
- 3. Generated a better marmoset reference sequence.
- 4. Developed hardware and software for automated and quantitative vocal communication analysis—with MIT Lincoln Laboratory
- 5. Developing hardware and software for automated and quantitative behavioral analysis with MIT Computer Science and Artificial Intelligence Laboratory (CSAIL).
- 6. Establishing brain imaging methods—in collaboration with Martino Center for Biomedical Imaging at MGH
- 7. Single ell transcriptome analysis using Dropseq—with Steve McCarroll at Harvard.

# Shank3 mutations cause autism spectrum disorder



## Marmoset embryos injected with Shank3 Cas9/sgRNA

(In collaboration with Jim Pickel at NIMH)



Embryo # 1: at least one allele carrying large deletion (43bp), will induce frameshift and loss of shank3 protein, as revealed by direct sequencing from PCR product



Embryo # 27: at least one allele carrying insertion (85bp), will induce frameshift and loss of shank3 protein, as revealed by direct sequencing from PCR product

	203	210	220	230	240	250	260	270	280	290	300	310	320	330	340	350
305Pickel Embryo27 Shk3F2R2 31 NUMHMarmoselSbk3F2B2W1	203 CA(	CAGGCTGGGGG	CCGAAGAGGA	CGGCGGCAG	AGGCCAGC	CGGCAGAGGC		TGCTGGGCCA	TAAGCTCGCTG	CCGAGGAC	TGTTTCAGCA		GAAGGTCA	CCCCGGAGTTGG		CATGCAGI
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# **Developing behavioral testing and monitoring systems**

#### **Vocal behavior**



## **Vocal behavior of adults**



# **Vocal behavior**

#### Automatic call detection





## Vocal interactions Influence modeling



## **Developing behavioral testing and monitoring systems**

# Automated tracking and behavior recognition using depth cameras and machine learning



**Estimated Gaze Direction Over Time** 

## Cynomolgus macaque embryos injected with Shank3 Cas9/sgRNA (In collaboration with Shihua Yang and Xiang Peng in Guangzhou, China)



Modified embryos: #4, #5, #12, #14

Targeting rate: ~30%

## Toward development of macaque models of Shank3 mutations

#### Macaque model

- Established a collaborative project with Shenzhen Institute of Advanced Technology: Liping Wang, Huihui Zhou and the team.
- Active collaboration with Andy (Peng) Xiang at Sun Yat-sen University and Shihua Yang at Southern China Agricultural University to generate Shank3 mutant Cynomolgus macaques
- Generated several Shank3 mutant macaque monkeys
- Behavioral testing is in progress



Heterozygous Shank3 deletion

Homozygous Shank3 deletion



# **Looking Forward**

- 1. Generating multiple monogenic disease models for studying neurobiological mechanisms.
- 2. Focus on prefrontal cortex, developmental process and longitudinal molecular and cellular changes.
- 3. Improve CRISPR/Cas9-mediated homologous recombination efficiency.
- 4. Develop cell type-specific Cre lines
- 5. Mapping disease relevant cell type-specific molecular changes.
- 6. Identify cell type-specific targets for correcting circuit dysfunction.

# **Our Primate Genetic Engineering Team**

<u>Guoping Feng</u>, (MIT and Stanley Center for Psychiatric Research at Broad Institute)
<u>Feng Zhang</u>, (MIT and Broad Institute)
<u>Robert Desimone</u> and McGovern Institute for Brain Research
<u>Steve Hyman</u> and the Stanley Center for Psychiatric Research
<u>Steve McCarroll</u>, (Harvard Medical School and Stanley Center for Psychiatric Research)
<u>Laura Brattain, Thomas Quatieri, Edward Wack</u> and MIT Lincoln Lab
<u>Yang Zhou, Rogier Landman, Julia Hyman, Jitendra Sharma, Charles Jennings, Martin Wienisch</u>
<u>Alan Wisler, Karthik Srinivasan, Ricardo del Rosario</u> (MIT and Broad)
<u>James Fox, Robert Marini and Monika Burns,</u> (Division of Comparative Medicine, MIT)
<u>Many others at MIT and Stanley Center/Broad Institute</u> (Jacquelyn Stathopoulos)

#### **Key Collaborators:**

Jim Pickel, (Director of NIMH Transgenic Core)
<u>Afonso Silva, (Investigator, NIMH)</u>
Hideyuki Okano and Erika Sasaki (Professors at Keio University and CIEA.
<u>Shihua Yang</u> (Professor, South China Agricultural University)
<u>Peng Xiang</u> (Professor, Sun Yat-sen University)
<u>Jon Hennebold</u> (Director of Reproduction lab, Oregon Primate Center)
<u>Liping Wang</u> (Professor, Shenzhen Institute of Advanced Technology, CAS)
<u>Huihui Zhou</u> (Professor, Shenzhen Institute of Advanced Technology, CAS)
<u>SIAT Peacock Team</u>: Xingtian Wu, Fuqiang Xu, Yu Chen, Yang Zhan

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